



B.S. Abdur Rahman™

**Crescent**

Institute of Science & Technology

Deemed to be University u/s 3 of the UGC Act, 1956

*Regulations 2022  
Curriculum and Syllabi  
(Updated upto April 2023, as per  
20<sup>th</sup> Academic Council)*

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**M.Sc. (Microbiology)**



**REGULATIONS 2022  
CURRICULUM AND SYLLABI  
(Updated upto April 2023, as per 20<sup>th</sup> Academic Council)**

**M.SC. MICROBIOLOGY**



## **VISION AND MISSION OF THE INSTITUTION**

### **VISION**

B.S. Abdur Rahman Crescent Institute of Science and Technology aspires to be a leader in Education, Training and Research in multidisciplinary areas of importance and to play a vital role in the Socio-Economic progress of the Country in a sustainable manner.

### **MISSION**

- To blossom into an internationally renowned Institute.
- To empower the youth through quality and value-based education.
- To promote professional leadership and entrepreneurship.
- To achieve excellence in all its endeavors to face global challenges.
- To provide excellent teaching and research ambience.
- To network with global Institutions of Excellence, Business, Industry and Research Organizations.
- To contribute to the knowledge base through Scientific enquiry, Applied Research and Innovation.



## **SCHOOL OF LIFE SCIENCES**

### **VISION AND MISSION**

#### **Vision**

To attain new heights in biotechnology education and research, shaping life sciences into a premier precision tool for the future for creation of wealth and ensuring social justice-specially for the welfare of the socially weaker group

#### **Mission**

- The mission of the school of life sciences is to maximize the benefits of Biotechnology to the Institute, the nation and the globe
- Being an excellent quality, comprehensive, multidisciplinary school that supports, coordinates, disseminates knowledge to the community
- Apply biotechnology in the areas of social welfare and entrepreneurship



**PROGRAMME EDUCATIONAL OBJECTIVES AND OUTCOMES****M.Sc. (MICROBIOLOGY)****PROGRAMME EDUCATIONAL OBJECTIVES:**

The course aims to provide an advanced understanding of the core principles and topics of Microbiology and their experimental basis to enable students in acquiring a specialized knowledge by means of dedicated lecture series and subject-oriented research project. Hence, the main objectives of the program are:

- To communicate the basic knowledge in general microbiology with detailed subdivision of microbiology.
- For the basic understanding this course includes microbial biochemistry, physiology and molecular Biology to give basic understanding of the microbiology.
- Further bacteriology, virology, mycology give individual sections of microbiology with detailed information on economic importance of microbiology.
- Finally this course explains the advanced sections of microbiology like Immunology, Microbial genetics, food microbiology, medical microbiology, Environmental microbiology, industrial microbiology and bioinformatics.
- This course provides necessary theoretical and practical experience in all divisions of microbiology to become an effective professional.
- This course provides broad exposure to various communities, ecological and commercial issues in the field microbiology.



**PROGRAMME OUTCOMES:**

After successfully completing this course, the student should be able to:

- On completion of the program the graduates will have applied knowledge in microbiology and the subdivision of microbiology.
- Microbial Biochemistry, Physiology and Molecular Biology will give basic understanding of the microbiology.
- Individual sections like bacteriology, virology, mycology give will give knowledge detailed information on economic importance of microbiology. develop ability to independently carry out a complete scientific work process, such as understanding of theoretical background, hypothesis generation, collection and analysis of data, and interpretation and presentation of results.
- Assess and predict the technological, ethical and social effects of their own work /disciplines and of microbiology.

**B.S. ABDUR RAHMAN CRESCENT INSTITUTE OF SCIENCE AND  
TECHNOLOGY, CHENNAI – 600 048.**

**REGULATIONS 2022**

**M.Tech. / MCA / M.Sc. / M.Com. / M.A. DEGREE PROGRAMMES**

***(Under Choice Based Credit System)***

**1.0 PRELIMINARY DEFINITIONS AND NOMENCLATURE**

In these Regulations, unless the context otherwise requires:

- i) **"Programme"** means post graduate degree programme (M.Tech. / MCA / M.Sc. / M.Com. / M.A.)
- ii) **"Branch"** means specialization or discipline of programme like M.Tech. in Structural Engineering, Food Biotechnology etc., M.Sc. in Physics, Chemistry, Actuarial Science, Biotechnology etc.
- iii) **"Course"** means a theory / practical / laboratory integrated theory / mini project / seminar / internship / project and any other subject that is normally studied in a semester like Advanced Concrete Technology, Electro Optic Systems, Financial Reporting and Accounting, Analytical Chemistry, etc.
- iv) **"Institution"** means B.S. Abdur Rahman Crescent Institute of Science and Technology.
- v) **"Academic Council"** means the Academic Council, which is the apex body on all academic matters of this Institute.
- vi) **"Dean (Academic Affairs)"** means the Dean (Academic Affairs) of the Institution who is responsible for the implementation of relevant rules and regulations for all the academic activities.
- vii) **"Dean (Student Affairs)"** means the Dean (Students Affairs) of the Institution who is responsible for activities related to student welfare and discipline in the campus.
- viii) **"Controller of Examinations"** means the Controller of Examinations of the Institution who is responsible for the conduct of examinations and declaration of results.
- ix) **"Dean of the School"** means the Dean of the School of the department concerned.
- x) **"Head of the Department"** means the Head of the Department concerned.

## 2.0 PROGRAMMES OFFERED AND ADMISSION REQUIREMENTS

### 2.1 Programmes Offered

The various programmes and their mode of study are as follows:

Degree	Mode of Study
M.Tech.	Full Time
MCA	
M.Sc.	
M.Com.	
M.A.	

### 2.2 ADMISSION REQUIREMENTS

**2.2.1** Students for admission to the first semester of the Master's Degree Programme shall be required to have passed the appropriate degree examination as specified in the clause 3.2 [Eligible entry qualifications for admission to programmes] of this Institution or any other University or authority accepted by this Institution.

**2.2.2** The other conditions for admission such as class obtained, number of attempts in the qualifying examination and physical fitness will be as prescribed by the Institution from time to time.

### 3.0 DURATION, ELIGIBILITY AND STRUCTURE OF THE PROGRAMME

**3.1.** The minimum and maximum period for completion of the programmes are given below:

Programme	Min. No. of Semesters	Max. No. of Semesters
M.Tech.	4	8
MCA	4	8
M.Sc.	4	8
M.Com.	4	8
M.A.	4	8

**3.1.1** Each academic semester shall normally comprise of 90 working days. Semester end examinations shall follow within 10 days of the last Instructional day.

**3.1.2** Medium of instruction, examinations and project report shall be in English.

### **3.2 ELIGIBLE ENTRY QUALIFICATIONS FOR ADMISSION TO PROGRAMMES**

<b>Sl. No.</b>	<b>Name of the Department</b>	<b>Programmes offered</b>	<b>Eligibility for Admission in M.Tech. / MCA / M.Sc. / M.Com. / MA Programmes</b>
1.	Aeronautical Engineering	M.Tech. (Avionics)	B.E. / B.Tech. in Aeronautical Engineering / Aerospace Engineering / Mechanical Engineering / Mechatronics / EEE / ECE / EIE / or Equivalent degree in relevant field.
2.	Civil Engineering	M.Tech. (Structural Engineering)	B.E. / B.Tech. in Civil Engineering / Structural Engineering or Equivalent degree in relevant field.
		M. Tech. (Construction Engineering and Project Management)	B.E. / B.Tech. in Civil Engineering / Structural Engineering / B.Arch. or Equivalent degree in relevant field.
3.	Mechanical Engineering	M.Tech. (CAD/CAM)	B.E. / B.Tech. in Mechanical / Automobile / Manufacturing / Production / Industrial / Mechatronics / Metallurgy / Aerospace / Aeronautical / Material Science / Polymer / Plastics / Marine Engineering or Equivalent degree in relevant field.
4.	Electrical and Electronics Engineering	M.Tech. (Power Systems Engineering)	B.E. / B.Tech. in EEE / ECE / EIE / ICE / Electronics / Instrumentation Engineering or Equivalent degree in relevant field.
5.	Electronics and Communication Engineering	M.Tech. (VLSI and Embedded Systems)	B.E. / B.Tech. in ECE / EIE / ICE / EEE / IT or Equivalent degree in relevant field.
6.	Computer Science and Engineering	M.Tech. (Computer Science and Engineering)	B.E. / B.Tech. in CSE / IT / ECE / EEE / EIE / ICE / Electronics Engineering / MCA or Equivalent degree in relevant field.
		M.Tech. (Artificial Intelligence and Data Science)	B.E. / B.Tech. in CSE / IT / ECE / EEE / EIE / ICE / Electronics Engineering / MCA or Equivalent degree in relevant field.
7.	Information Technology	M.Tech. (Information Technology)	B.E. / B.Tech. in IT / CSE / ECE / EEE / EIE / ICE / Electronics Engineering / MCA or Equivalent degree in relevant field.

Sl. No.	Name of the Department	Programmes offered	Eligibility for Admission in M.Tech. / MCA / M.Sc. / M.Com. / MA Programmes
8.	Computer Applications	MCA	BCA / B.Sc. Computer Science / B.E. / B.Tech. / B.Sc. Mathematics, B.Sc. Physics / Chemistry / B.Com. / BBA / B.A. with Mathematics at graduation level or at 10 + 2 level or equivalent degree in relevant field.
9.	Mathematics	M.Sc. (Actuarial Science)	Any under graduate degree with Mathematics / Statistics as one of the subjects of study at 10 + 2 level.
10.	Physics	M.Sc.(Physics)	B.Sc. in Physics / Applied Science / Electronics / Electronics Science / Electronics & Instrumentation or Equivalent degree in relevant field.
11.	Chemistry	M.Sc.(Chemistry)	B.Sc. in Chemistry / Applied Science or Equivalent degree in relevant field.
12.	Life Sciences	M.Sc. Biochemistry & Molecular Biology	B.Sc. in Biotechnology / Biochemistry / Botany / Zoology / Microbiology / Molecular Biology / Genetics or Equivalent degree in relevant field.
		M.Sc. Biotechnology	B.Sc. in Biotechnology / Biochemistry / Botany / Zoology / Microbiology / Molecular Biology / Genetics or Equivalent degree in relevant field.
		M.Sc. Microbiology	B.Sc.in Biotechnology / Biochemistry / Botany / Zoology / Microbiology / Molecular Biology / Genetics or Equivalent degree in relevant field.
		M.Tech. Biotechnology	B.Tech. / B.E. in Biotechnology or Equivalent degree in relevant field.
		M.Tech. Food Biotechnology	B.E. / B.Tech. in Biotechnology / Food Biotechnology / Chemical Engineering / Biochemical Engineering / Industrial Biotechnology or Equivalent degree in relevant field.
13.	Commerce	M.Com	B.Com. / BBA

Sl. No.	Name of the Department	Programmes offered	Eligibility for Admission in M.Tech. / MCA / M.Sc. / M.Com. / MA Programmes
14.	Arabic and Islamic Studies	M.A. Islamic Studies	B.A. in Islamic Studies / Arabic (or) Afzal-ul-Ulama (or) Any under graduate degree with Part 1 Arabic (or) Any under graduate degree with Aalim Sanad / Diploma / Certificate in Arabic or Islamic Studies.

### 3.3. STRUCTURE OF THE PROGRAMME

**3.3.1** The PG. programmes consist of the following components as prescribed in the respective curriculum:

- i. Core courses
- ii. Elective courses
- iii. Laboratory integrated theory courses
- iv. Project work
- v. Laboratory courses
- vi. Open elective courses
- vii. Seminar
- viii. Mini Project
- ix. Industry Internship
- x. MOOC courses (NPTEL-Swayam, Coursera etc.)
- xi. Value added courses

**3.3.2** The curriculum and syllabi of all programmes shall be approved by the Academic Council of this Institution.

**3.3.3** For the award of the degree, the student has to earn a minimum total credits specified in the curriculum of the respective specialization of the programme.

**3.3.4** The curriculum of programmes shall be so designed that the minimum prescribed credits required for the award of the degree shall be within the limits specified below:

Programme	Range of credits
M.Tech.	76 -80
MCA	86
M.Sc.	77 - 85
M.Com.	88

M.A.	72
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**3.3.5** Credits will be assigned to the courses for all programmes as given below:

- ❖ One credit for one lecture period per week or 15 periods of lecture per semester.
- ❖ One credit for one tutorial period per week or 15 periods per semester.
- ❖ One credit each for seminar/practical session/project of two or three periods per week or 30 periods per semester.
- ❖ One credit for 160 hours of industry internship per semester for all programmes (except M.Com.)
- ❖ Four credits for 160 hours of industry internship per semester for M.Com.

**3.3.6** The number of credits the student shall enroll in a non-project semester and project semester is as specified below to facilitate implementation of Choice Based Credit System.

Programme	Non-project semester	Project semester
M.Tech.	9 to 32	18 to 26
MCA	9 to 32	18 to 26
M.Sc.	9 to 32	10 to 26
M.Com.	9 to 32	16 to 28
M.A.	9 to 32	NA

**3.3.7** The student may choose a course prescribed in the curriculum from any department offering that course without affecting regular class schedule. The attendance will be maintained course wise only.

**3.3.8** The students shall choose the electives from the curriculum with the approval of the Head of the Department / Dean of School.

**3.3.9** Apart from the various elective courses listed in the curriculum for each specialization of programme, the student can choose a maximum of two electives from any other similar programmes across departments, aliter to open electives, during the entire period of study, with approval of Head of the department offering the course and parent department.

### **3.4. ONLINE COURSES**

**3.4.1** Students are permitted to undergo department approved online courses under SWAYAM up to 40% of credits of courses in a semester excluding project semester (in case of M.Tech. M.Sc. & MCA programmes) with the recommendation of the Head of the Department / Dean of School and with the prior approval of Dean Academic Affairs during his/ her period of study. The credits earned through online courses shall be transferred following the due approval procedures. The online courses can be considered in lieu of core courses and elective courses.

**3.4.2** Students shall undergo project related online course on their own with the mentoring of the project supervisor.

### **3.5 PROJECT WORK**

**3.5.1** Project work shall be carried out by the student under the supervision of a faculty member in the department with similar specialization.

**3.5.2** A student may however, in certain cases, be permitted to work for the project in an Industry / Research organization, with the approval of the Head of the Department/ Dean of School. In such cases, the project work shall be jointly supervised by a faculty of the Department and an Engineer / Scientist / Competent authority from the organization and the student shall be instructed to meet the faculty periodically and to attend the review meetings for evaluating the progress.

**3.5.3** The timeline for submission of final project report / dissertation is within 30 calendar days from the last instructional day of the semester in which project is done.

**3.5.4** If a student does not comply with the submission of project report / dissertation on or before the specified timeline he / she is deemed to have not completed the project work and shall re-register in the subsequent semester.

## **4.0 CLASS ADVISOR AND FACULTY ADVISOR**

### **4.1 CLASS ADVISOR**

A faculty member shall be nominated by the HOD/ Dean of School as Class Advisor for the class throughout their period of study.



The class advisor shall be responsible for maintaining the academic, curricular and co-curricular records of students of the class throughout their period of study.

#### **4.2 FACULTY ADVISOR**

To help the students in planning their courses of study and for general counseling, the Head of the Department / Dean of School of the students shall attach a maximum of 20 students to a faculty member of the department who shall function as faculty advisor for the students throughout their period of study. Such faculty advisor shall guide the students in taking up the elective courses for registration and enrolment in every semester and also offer advice to the students on academic and related personal matters.

#### **5.0 COURSE COMMITTEE**

**5.1** Each common theory / laboratory course offered to more than one group of students shall have a "Course Committee" comprising all the teachers handling the common course with one of them nominated as course coordinator. The nomination of the course coordinator shall be made by the Head of the Department / Dean (Academic Affairs) depending upon whether all the teachers handling the common course belong to a single department or from several departments. The Course Committee shall meet as often as possible to prepare a common question paper, scheme of evaluation and ensure uniform evaluation of the assessment tests and semester end examination.

#### **6.0 CLASS COMMITTEE**

**6.1** A class committee comprising faculty members handling the classes, student representatives and a senior faculty member not handling the courses as chairman will be constituted in every semester:

**6.2** The composition of the class committee will be as follows:

- i) One senior faculty member preferably not handling courses for the concerned semester, appointed as chairman by the

Head of the Department

- ii) Faculty members of all courses of the semester
- iii) All the students of the class
- iv) Faculty advisor and class advisor
- v) Head of the Department – Ex officio member

**6.3** The class committee shall meet at least three times during the semester. The first meeting shall be held within two weeks from the date of commencement of classes, in which the nature of continuous assessment for various courses and the weightages for each component of assessment shall be decided for the first and second assessment. The second meeting shall be held within a week after the date of first assessment report, to review the students' performance and for follow up action.

**6.4** During these two meetings the student members, shall meaningfully interact and express opinions and suggestions to improve the effectiveness of the teaching-learning process, curriculum and syllabi of courses.

**6.5** The third meeting of the class committee, excluding the student members, shall meet within 5 days from the last day of the semester end examination to analyze the performance of the students in all the components of assessments and decide their grades in each course. The grades for a common course shall be decided by the concerned course committee and shall be presented to the class committee(s) by the concerned course coordinator.

## **7.0 REGISTRATION AND ENROLLMENT**

**7.1** The students of first semester shall register and enroll at the time of admission by paying the prescribed fees. For the subsequent semesters registration for the courses shall be done by the student one week before the last working day of the previous semester.

### **7.2 Change of a Course**

A student can change an enrolled course within 10 working days from the commencement of the course, with the approval of the Dean (Academic Affairs), on the recommendation of the Head of the Department of the student.

### 7.3 Withdrawal from a Course

A student can withdraw from an enrolled course at any time before the first continuous assessment test for genuine reasons, with the approval of the Dean (Academic Affairs), on the recommendation of the Head of the Department of the student.

7.4 A student can enroll for a maximum of 32 credits during a semester including Redo / Predo courses.

### 8.0 BREAK OF STUDY FROM PROGRAMME

8.1 A student may be allowed / enforced to take a break of study for two semesters from the programme with the approval of Dean (Academic Affairs) for the following reasons:

8.1.1 Medical or other valid grounds

8.1.2 Award of 'I' grade in all the courses in a semester due to lack of attendance

8.1.3 Debarred due to any act of indiscipline

8.2 The total duration for completion of the programme shall not exceed the prescribed maximum number of semesters (vide clause 3.1).

8.3 A student who has availed a break of study in the current semester (odd/even) can rejoin only in the subsequent corresponding (odd/even) semester in the next academic year on approval from the Dean (Academic affairs).

8.4 During the break of study, the student shall not be allowed to attend any regular classes or participate in any activities of the Institution. However, he / she shall be permitted to enroll for the 'I' grade courses and appear for the arrear examinations.

### 9.0 MINIMUM REQUIREMENTS TO REGISTER FOR PROJECT WORK

9.1 A student is permitted to register for project semester, if he/she has earned the minimum number of credits specified below:

Programme	Minimum no. of credits to be earned to enroll for project semester
M.Tech.	18
MCA	22

M.Sc.	18
M.Com	NA
M.A.	NA

- 9.2** If the student has not earned minimum number of credits specified, he/she has to earn the required credits, at least to the extent of minimum credits specified in clause 9.1 and then register for the project semester.

## **10.0 ATTENDANCE REQUIREMENT AND SEMESTER / COURSE REPETITION**

- 10.1** A student shall earn 100% attendance in the contact periods of every course, subject to a maximum relaxation of 25% to become eligible to appear for the semester end examination in that course, failing which the student shall be awarded "I" grade in that course.
- 10.2** The faculty member of each course shall cumulate the attendance details for the semester and furnish the names of the students who have not earned the required attendance in the concerned course to the class advisor. The class advisor shall consolidate and furnish the list of students who have earned less than 75% attendance, in various courses, to the Dean (Academic Affairs) through the Head of the Department / Dean of the School. Thereupon, the Dean (Academic Affairs) shall officially notify the names of such students prevented from writing the semester end examination in each course.
- 10.3** If a student secures attendance between 65% and less than 75% in any course in a semester, due to medical reasons (hospitalization / accident / specific illness) or due to participation in the institution approved events, the student shall be given exemption from the prescribed attendance requirement and the student shall be permitted to appear for the semester end examination of that course. In all such cases, the students shall submit the required documents immediately after joining the classes to the class advisor, which shall be approved by the Head of the Department / Dean of the School. The Vice Chancellor, based on the recommendation of the Dean (Academic Affairs) may approve the condonation of attendance.

- 10.4** A student who has obtained an “I” grade in all the courses in a semester is not permitted to move to the next higher semester. Such students shall repeat all the courses of the semester in the subsequent academic year. However, he / she is permitted to redo the courses awarded with 'I' grade / arrear in previous semesters. They shall also be permitted to write arrear examinations by paying the prescribed fee.
- 10.5** The student awarded “I” grade, shall enroll and repeat the course when it is offered next. In case of “I” grade in an elective course either the same elective course may be repeated or a new elective course may be taken with the approval of the Head of the Department / Dean of the School.
- 10.6** A student who is awarded “U” grade in a course shall have the option to either write the semester end arrear examination at the end of the subsequent semesters, or to redo the course when the course is offered by the department. Marks scored in the continuous assessment in the redo course shall be considered for grading along with the marks scored in the semester end (redo) examination. If any student obtains “U” grade in the redo course, the marks scored in the continuous assessment test (redo) for that course shall be considered as internal mark for further appearance of arrear examination.
- 10.7** If a student with “U” grade, who prefers to redo any particular course, fails to earn the minimum 75% attendance while doing that course, then he / she is not permitted to write the semester end examination and his / her earlier “U” grade and continuous assessment marks shall continue.

### **11.0 REDO COURSES**

- 11.1** A student can register for a maximum of two redo courses per semester without affecting the regular semester classes, whenever such courses are offered by the department concerned, based on the availability of faculty members, and subject to a specified minimum number of students registering for each of such courses.
- 11.2** The number of contact hours and the assessment procedure for any redo course shall be the same as regular courses, except

there is no provision for any substitute examination and withdrawal from a redo course.

## **12.0 ASSESSMENT PROCEDURE AND PERCENTAGE WEIGHTAGE OF MARKS**

**12.1** Every theory course shall have a total of three assessments during a semester as given below:

<b>Assessments</b>	<b>Weightage of Marks</b>
Continuous Assessment 1	25%
Continuous Assessment 2	25%
Semester End Examination	50%

### **12.2 Theory Course**

Appearing for semester end theory examination for each course is mandatory and a student shall secure a minimum of 40% marks in each course in semester end examination for the successful completion of the course.

### **12.3 Laboratory Course**

Every practical course shall have 75% weightage for continuous assessments and 25% for semester end examination. However, a student shall have secured a minimum of 50% marks in the semester end practical examination for the award of pass grade.

### **12.4 Laboratory Integrated Theory Courses**

For laboratory integrated theory courses, the theory and practical components shall be assessed separately for 100 marks each and consolidated by assigning a weightage of 75% for theory component and 25% for practical component. Grading shall be done for this consolidated mark. Assessment of theory components shall have a total of three assessments with two continuous assessments carrying 25% weightage each and semester end examination carrying 50% weightage. The student shall secure a separate minimum of 40% in the semester end theory examination. The evaluation of practical components shall be through continuous assessment.

**12.5** The components of continuous assessment for theory/practical/laboratory integrated theory courses shall be finalized in the first class committee meeting.

### **12.6 Industry Internship**

In the case of industry internship, the student shall submit a report, which shall be evaluated along with an oral examination by a committee of faculty members constituted by the Head of the Department. The student shall also submit an internship completion certificate issued by the industry / research / academic organisation. The weightage of marks for industry internship report and viva voce examination shall be 60% and 40% respectively.

### **12.7 Project Work**

In the case of project work, a committee of faculty members constituted by the Head of the Department / Dean of the School will carry out three periodic reviews. Based on the project report submitted by the students, an oral examination (viva voce) shall be conducted as semester end examination by an external examiner approved by the Controller of Examinations. The weightage for periodic reviews shall be 50%. Of the remaining 50%, 20% shall be for the project report and 30% for the viva voce examination.

**12.8** The assessment of seminar course including its component and its weightage shall be decided by a committee of faculty members constituted by the Head of the Department. This committee shall ensure the conduct of assessment of components and award marks accordingly.

**12.9** For the first attempt of the arrear theory examination, the internal assessment marks scored for a course during first appearance shall be used for grading along with the marks scored in the arrear examination. From the subsequent appearance onwards, full weightage shall be assigned to the marks scored in the semester end examination and the internal assessment marks secured during the course of study shall become invalid.

In case of laboratory integrated theory courses, after one regular and one arrear appearance, the internal mark of theory component is invalid and full weightage shall be assigned to the

marks scored in the semester end examination for theory component. There shall be no arrear or improvement examination for lab components.

### **13.0 SUBSTITUTE EXAMINATIONS**

**13.1** A student who is absent, for genuine reasons, may be permitted to write a substitute examination for any one of the two continuous assessment tests of a course by paying the prescribed substitute examination fee. However, permission to take up a substitute examination will be given under exceptional circumstances, such as accidents, admission to a hospital due to illness, etc. by a committee constituted by the Head of the Department / Dean of School for that purpose. However, there is no substitute examination for semester end examination.

**13.2** A student shall apply for substitute exam in the prescribed form to the Head of the Department / Dean of School within a week from the date of assessment test. However, the substitute examination will be conducted only after the last working day of the semester and before the semester end examination.

### **14.0 SUPPLEMENTARY EXAMINATION**

**14.1** Final Year students can apply for supplementary examination for a maximum of three courses thus providing an opportunity to complete their degree programme. Likewise, students with less credit can also apply for supplementary examination for a maximum of three courses to enable them to earn minimum credits to move to higher semester. The students can apply for supplementary examination within three weeks of the declaration of results in both odd and even semesters.

### **15. PASSING, DECLARATION OF RESULTS AND GRADE SHEET**

**15.1** All assessments of a course shall be made on absolute marks basis. However, the Class Committee without the student members shall preferably meet within 5 days after the semester end examination and analyze the performance of students in all



assessments of a course and award letter grades. The letter grades and the corresponding grade points are as follows:

Letter Grade	Grade Points
S	10
A	9
B	8
C	7
D	6
E	5
U	0
I	0

“I” denotes inadequate attendance and hence prevented from appearing for semester end examination

“U” denotes unsuccessful performance in the course.

- 15.2** A student who earns a minimum of five grade points ('E' grade) in a course is declared to have successfully completed the course. Such a course cannot be repeated by the student for improvement of grade.
- 15.3** The results, after awarding of grades, shall be signed by the Chairman of the Class Committee and Head of the Department/Dean of School and it shall be declared by the Controller of Examinations.
- 15.4** Within one week from the date of declaration of result, a student can apply for revaluation of his / her semester end theory examination answer scripts of one or more courses, on payment of prescribed fees to the Controller of Examinations. Subsequently the Head of the Department/ Dean of School offered the course shall constitute a revaluation committee consisting of Chairman of the Class Committee as convener, the faculty member of the course and a senior faculty member knowledgeable in that course as members. The committee shall meet within a week to re-evaluate the answer scripts and submit its report to the Controller of Examinations for consideration and decision.

- 15.5** After results are declared, grade sheets shall be issued to each student, which contains the following details: a) list of courses enrolled during the semester including redo courses / arrear courses, if any; b) grades scored; c) Grade Point Average (GPA) for the semester and d) Cumulative Grade Point Average (CGPA) of all courses enrolled from first semester onwards.

GPA is the ratio of the sum of the products of the number of credits of courses registered and the grade points corresponding to the grades scored in those courses, taken for all the courses, to the sum of the number of credits of all the courses in the semester.

If  $C_i$ , is the number of credits assigned for the  $i^{\text{th}}$  course and  $GP_i$  is the Grade Point in the  $i^{\text{th}}$  course

$$GPA = \frac{\sum_{i=1}^n (C_i)(GP_i)}{\sum_{i=1}^n C_i}$$

Where  $n$  = number of courses

The Cumulative Grade Point Average (CGPA) is calculated in a similar manner, considering all the courses enrolled from first semester.

"I" grade is excluded for calculating GPA.

"U" and "I" grades are excluded for calculating CGPA.

The formula for the conversion of CGPA to equivalent percentage of marks is as follows:

Percentage Equivalent of Marks = CGPA X 10

- 15.6** After successful completion of the programme, the Degree shall be awarded upon fulfillment of curriculum requirements and classification based on CGPA as follows:

Classification	CGPA
First Class with Distinction	8.50 and above and passing all the courses in first appearance and completing the programme within the minimum prescribed period.
First Class	6.50 and above and completing the programme within a minimum prescribed period plus two semesters.
Second Class	Others

**15.6.1 Eligibility for First Class with Distinction**

- A student should not have obtained 'U' or 'I' grade in any course during his/her study
- A student should have completed the PG programme within the minimum prescribed period of study (except clause 8.1.1)

**15.6.2 Eligibility for First Class**

A student should have passed the examination in all the courses not more than two semesters beyond the minimum prescribed period of study (except clause 8.1.1)

**15.6.3** The students who do not satisfy clause 15.6.1 and clause 15.6.2 shall be classified as second class.

**15.6.4** The CGPA shall be rounded to two decimal places for the purpose of classification. The CGPA shall be considered up to three decimal places for the purpose of comparison of performance of students and ranking.

**16.0 DISCIPLINE**

**16.1** Every student is expected to observe discipline and decorum both inside and outside the campus and not to indulge in any activity which tends to affect the reputation of the Institution.

**16.2** Any act of indiscipline of a student, reported to the Dean (Student Affairs), through the HOD / Dean shall be referred to a Discipline and Welfare Committee constituted by the Registrar for taking appropriate action.

**17.0 ELIGIBILITY FOR THE AWARD OF THE MASTER'S DEGREE**

**17.1** A student shall be declared to be eligible for the award of the Master's Degree, if he/she has:

- i. Successfully acquired the required credits as specified in the curriculum corresponding to his/her programme within the stipulated time.
- ii. No disciplinary action is pending against him/her.
- iii. Enrolled and completed at least one value added course.
- iv. Enrollment in at least one MOOC / SWAYAM course (non-credit) before the final semester.

**17.2** The award of the degree must have been approved by the Institute.

**18.0 POWER TO MODIFY**

Notwithstanding all that have been stated above, the Academic Council has the right to modify any of the above regulations from time to time.

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**B.S. ABDUR RAHMAN CRESCENT INSTITUTE OF SCIENCE AND  
TECHNOLOGY**

**M.Sc.MICROBIOLOGY**

**CURRICULUM FRAMEWORK, REGULATIONS 2022**

*(Choice Based Credit System)*

**SEMESTER I**

<b>Sl. No.</b>	<b>Course Group</b>	<b>Course Code</b>	<b>Course Title</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
1.	CCE	ENE 6182	English for Career Development	2	1	0	3
2.	PCC	LSE 6121	Microbial Genetics	4	0	0	4
3.	PCC	LSE 6103	Principles of Microbiology	3	0	0	3
4.	PCC	LTE 6105	Cell and Molecular Biology	3	0	0	3
5.	PCC	LSE 6122	Laboratory I (Genetics/ Microbiology/ Cell and Molecular Biology)	0	0	4	2
6.	PEC		Professional Elective I	3	0	0	3
7.	PEC		Professional Elective II	3	0	0	3
<b>Credits</b>							<b>21</b>

**SEMESTER II**

<b>Sl. No.</b>	<b>Course Group</b>	<b>Course Code</b>	<b>Course Title</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
1.		GEE 6202	Research Methodology and IPR	3	0	0	3
2.	PCC	LSE 6221	Environmental and Medical Microbiology	3	0	0	3
3.	PCC	LSE 6241	Bioinformatics	4	0	0	4
5.	PCC	LSE 6222	Bioprocess and Fermentation Technology	3	0	0	3
6.	PEC	LSE 6223	Laboratory II(Medical Microbiology/ Bioinformatics/ Bioprocess Technology)	0	0	4	2
7.	PEC		Professional Elective III	3	0	0	3
8.	PEC		Professional Elective IV	3	0	0	3
<b>Credits</b>							<b>21</b>

**SEMESTER III**

Sl. No.	Course Group	Course Code	Course Title	L	T	P	C
1.	OEC		Open Elective I	3	0	0	3
2.	PCC	LSE 7121	Industrial, Food and Agricultural Microbiology	4	0	0	4
3.	PCC	LSE 7122	Immunology	4	0	0	4
5.	PCC	LSE 7123	Laboratory III (Industrial, Food and Agricultural Microbiology/ Immunology)	0	0	4	2
6.	PEC		Professional Elective V	3	0	0	3
7.	PEC		Professional Elective VI	3	0	0	3
8.	PCC	LSE 7211	Project Work Phase I **	0	0	12	4
9.	PCC		MOOC course (related to the project)	0	0	0	0
		LSE 7124	Industry Internship	0	0	2	2
<b>Credits</b>							<b>21</b>

**SEMESTER IV**

Sl. No.	Course Group	Course Code	Course Title	L	T	P	C
1.	PCC	LSE 7211	Project Work Phase II	0	0	32	16
<b>Total Credits</b>						<b>4 + 16= 20</b>	

**Overall Total Credits – 83**

# Industrial training will be undertaken during first year summer vacation for 30 days. The credit will be awarded in the 3rd Semester.

\*\* Credits for project work phase I in III semester to be accounted along with project work phase II in IV semester

**LIST OF PROFESSIONAL ELECTIVE COURSES**

Sl. No.	Course Group	Course Code	Course Title	L	T	P	C
<b>Semester I</b>							
1.	PEC	LSEY101	Biostatistics	3	0	0	3
2.	PEC	LSEY102	Biosafety, Bioethics, Bioentrepreneurship and IPR)	3	0	0	3
	PEC	LSEY103	Recombinant DNA Technology	3	0	0	3
3.	PEC	LSEY121	Microbial Diversity and Extremophiles	3	0	0	3
4.	PEC	LSEY122	Microbial Physiology and Metabolism	3	0	0	3
5.	PEC	LSEY123	Parasitology	3	0	0	3
<b>Semester II</b>							
1.	PEC	LSEY201	Molecular Diagnostics	3	0	0	3
2.	PEC	LSEY202	Plant and Animal Biotechnology	4	0	0	4
3.	PEC	LSEY203	Protein Engineering	3	0	0	3
4.	PEC	LSEY211	Molecular Microbiology	3	0	0	3
5.	PEC	LSEY212	Fermentation Technology	3	0	0	3
6.	PEC	LSEY213	Host-microbe Interactions	3	0	0	3
<b>Semester III</b>							
1.	PEC	LSEY111	Nanobiotechnology	3	0	0	3
2.	PEC	LSEY112	Medical Coding	3	0	0	3
3.	PEC	LSEY113	Gene Manipulation	3	0	0	3
4.	PEC	LSEY131	Antibiotics	3	0	0	3
5.	PEC	LSEY132	Microbial Systems Biology	3	0	0	3
6.	PEC	LSEY133	Metagenomics	3	0	0	3



**LIST OF OPEN ELECTIVE COURSES OFFERED TO M.S.C.  
PROGRAMMES UNDER REGULATIONS 2022**

Sl. No.	Course Code	Course Title	L	T	P	C	Offering Department / School
1.	OEEY 731	Advanced Materials for Energy Applications	3	0	0	3	Physics
2.	OEEY 732	Alternative Energy Resources	3	0	0	3	Chemistry
3.	OEEY 701	Analytical Techniques	3	0	0	3	Chemistry
4.	OEEY 733	Biomass for Energy Applications	3	0	0	3	Chemistry
5.	OEEY 703	Biomaterials	3	0	0	3	Physics
6.	OEEY 704	Biomedical Instrumentation	3	0	0	3	Physics
7.	OEEY 705	Biophotonics	3	0	0	3	Physics
8.	OEEY 734	Corrosion and Corrosion Control	3	0	0	3	Chemistry
9.	OEEY 735	Corrosion Science and Technology	3	0	0	3	Physics
10.	OEEY 736	Environmental Chemistry	3	0	0	3	Chemistry
11.	OEEY 737	Fuel Cells for Sustainable Energy Production	3	0	0	3	Chemistry
12.	OEEY 738	Green and Sustainable Chemistry	3	0	0	3	Chemistry
13.	OEEY 739	Industrial Pollution Control	3	0	0	3	Chemistry
14.	OEEY 740	Introduction to Embedded System	3	0	0	3	ECE
15.	OEEY 741	Matlab Programming	3	0	0	3	ECE
16.	OEEY 710	Nanotechnology and Catalysis	3	0	0	3	Chemistry
17.	OEEY 715	Structural Interpretation of Materials	3	0	0	3	Chemistry
18.	OEEY 742	Surface Coating Technology	3	0	0	3	Chemistry
19.	OEEY 743	Thin Film Science and Technology	3	0	0	3	Physics

**SEMESTER I**

<b>ENE 6182</b>	<b>PROFESSIONAL COMMUNICATION</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4 and 8</b>		<b>2</b>	<b>1</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To enhance the Employability and Career Skills of students

**COB2:** To orient the students towards grooming as a professional

**COB3:** To make them Employability Graduates

**COB4:** To train students making effective presentations and discussion on various topics.

**MODULE I COMMUNICATION AT WORKPLACE 6+1**

Language and communication-Communication at the workplace- Formal and informal communication- Direction of flow of communication- Non-verbal communication- Communication and organizational culture-Communication and inter- personal relations- Importance of the 'U' in communication

**MODULE II PRESENTATION SKILLS 6+5**

(This module focuses more on the practical aspects of communication for career development.)

Importance of presentation skills-Overcoming the fear of public speaking towards making effective presentations- A step-by-step approach to presentations –planning the presentation-Gathering feedback- Making the presentation

**MODULE III CORRESPONDENCE AT WORK 6+3**

Importance of workplace correspondence-Types of correspondence- Mechanics of effective business correspondence-Tips for effective correspondence-The seven Cs of communication- Writing effective emails- Email etiquette-Personal touch in business communication

**MODULE IV TEAMWORK 6+5**

(This module focuses more on the practical aspects of communication for career development.)

Importance of team work-Understanding team behavior-Team as an employability skill- Team formation and development- Pooling competencies in a team- Significance of team spirit-How to be an effective team player – Group Discussion

**MODULE V      WORKPLACE ETIQUETTE****6+1**

Etiquette in modern workplace- Workplace etiquette- global and local Culture sensitivity-Gender sensitivity- importance of grooming-Etiquette in interaction- Netiquette

**L - 30, T - 15, TOTAL HOURS - 45****REFERENCES:**

1. Butterfield, Jeff Soft Skills for Everyone. Cengage Learning: New Delhi, 2015
2. Interact English Lab Manual for Undergraduate Students, Orient Black Swan: Hyderabad, 2016.
3. E. Suresh Kumar et al. Communication for Professional Success. Orient Blackswan: Hyderabad, 2015
4. Raman, Meenakshi and Sangeeta Sharma. Professional Communication. Oxford University Press: Oxford, 2014
5. S. Hariharan et al. Soft Skills. MJP Publishers: Chennai, 2010.

**COURSE OUTCOMES:****CO1:** Identify the flows of communication**CO2:** Make effective presentations**CO3:** Write effective business correspondences.**CO4:** Participate in group discussions and teamwork confidently.**CO5:** Follow appropriate workplace etiquette**Board of Studies (BoS) :**

15<sup>th</sup> BoS of the Department of English held on 14.6.2022

**Academic Council:**

19<sup>th</sup> Meeting of the Academic Council held on 29.09.2022

**SDG 4:** Ensure inclusive and equitable quality education and promote lifelong learning opportunities for all

**SDG 8:** Promote sustained, inclusive and sustainable economic growth, full and productive employment and decent work for all

**Statement:** This course ensures that the students acquire quality education and are also made eligible to obtain productive and decent employment.

<b>LSE 6121</b>	<b>MICROBIAL GENETICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3,15</b>		<b>4</b>	<b>0</b>	<b>0</b>	<b>4</b>

**COURSE OBJECTIVES:**

**COB1:** To get overview on microbial genetics, concepts, theories and genetic tools.

**COB2:** To learn about the genetics of bacteriophage

**COB3:** To get knowledge on fungal and viral genetics.

**COB4:** To understand the concept of operon gene expression and different types of operons.

**COB5:** To understand regulation of gene expression

**MODULE I BACTERIOPHAGE 12**

Structure, Life Cycle and Lytic-Lysogenic Options of A Bacteriophage. The lambda Lifecycle, Genes involved in lysogeny and lytic cycle of lambda phage. Lifecycle Of M13-M13 Adsorption and Injection, Protection of The M13 Genome. Lifecycle of P1, Adsorption, Injection and P1 Transducing Particles. Lifecycle of T4-T4 Adsorption and Injection.

**MODULE II METHODS OF GENE TRANSFER IN BACTERIA 12**

Genetics of Bacteria, Mutations and growth in bacteria. Mechanisms of Genetic Exchange in Bacteria- Transformation, conjugation, Properties of the F plasmid, F+ x F - mating, F' x F- conjugation. Hfr bacteria and chromosome mapping, Transduction: Generalized and specialized transduction.

**MODULE III TRANSPOSABLE GENETIC ELEMENTS 12**

Transposable elements in bacteria and yeast, Insertion sequences-structure and occurrence, Mechanism of transpositions –Replicative, Non replicative, regulation and activation. Conjugative transpositions, Phase variation.

**MODULE IV FUNGAL GENETICS 12**

Fungi as Model for genetic studies, Structure and Organization of Fungal Genome, Life Cycles, Mating Systems and Genetic Exchange in Fungi: Life cycles and sexual process, Mating system and promotion of outcrossing, Systems restricting outcrossing, Sexual differentiation and relationship to outcrossing, Heterokaryosis, the parasexual cycle and Vegetative incompatibility, Role and consequences of Vegetative incompatibility, Sex

and Outcrossing, Neurospora and Classical Genetics, Applied molecular genetics of fungi.

**MODULE V REGULATION OF GENE EXPRESSION 12**

Constitutive, Inducible, and Repressible Gene expression, positive and negative control of gene expression operons: coordinately regulated units of gene expression. The lactose operon in e. coli: induction and catabolite repression. The tryptophan operon in E. coli: repression and attenuation. Translational control of gene expression, posttranslational regulatory mechanisms

**L – 60; TOTAL HOURS –60**

**TEXT BOOKS:**

1. Snyder L and Champness W. Molecular Genetics of Bacteria. 3rd Ed, ASM Press, Washington, 2002.
2. Baumberg S. Prokaryotic gene expression. Oxford University Press 2002.
3. Watson J. D, Hopkins N. H, Roberts J. W, Steitz J. A and Weiner A. M. Molecular Biology of the Gene, 4th Ed., Benjamin/Cummings, 1987.
4. Streips U. N, Yasbin R. E. Modern Microbial Genetics, 2<sup>nd</sup> Ed., Wiley-Liss, Inc. 2002.

**COURSE OUTCOMES:**

**CO1:** Understand the structure of viruses, know about the different forms based on life cycle

**CO2:** Introduced to the concept of transfer of genetic information between viruses and host cells

**CO3:** Will get information on jumping genes, their occurrence in different life forms

**CO4:** Understand the use of fungus in carrying out the genetic studies

**CO5:** Also learn about genetic regulation of essential life processes.

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on  
20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	-	-	-	-	-	-
CO2	H	L	-	-	-	-	-	-	-	-	-	-
CO3	H	L	-	-	-	-	-	-	-	-	-	-
CO4	H	L	-	-	-	-	-	-	-	-	-	-
CO5	H	L	-	-	-	-	-	-	-	-	-	-

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of microbial genetics we can understand the genetic basis of underlying diseases, that will help in good health and well being.

### SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to microbial geneticists that gives knowledge about relation with all the levels of life in the earth.

<b>LSE 6103</b>	<b>PRINCIPLES OF</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>MICROBIOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** Describe how microorganisms are used as model systems to study basic biology, genetics, metabolism, and ecology.

**COB2:** Identify ways microorganisms play an integral role in disease, and microbial and immunological methodologies are used in disease treatment and prevention.

**COB3:** To provide an introduction to the science of microbiology, particularly medical microbiology, to the student with both limited background in the biological sciences and limited interest in pursuing this field further.

**COB4:** To provide concepts of microbial metabolism, growth, and control of microbes.

**COB5:** Describe the opportunity available in applied & industrial microbiology scope through the different areas of applications.

**MODULE I INTRODUCTION TO MICROBIOLOGY 9**

Types of microorganisms. A brief history of microbiology. Types of Microorganisms- Viruses, Bacteria, Protozoa, Fungi, Algae, Archaea and differences. Classification of microorganism and methods of classifying and identification of microorganisms. Size, shape, and arrangement of bacterial cells. Bacterial cell structures, Structures external to the cell wall, structures internal to the cell wall.

**MODULE II VIRUS, FUNGI, ALGAE AND PROTOZOA 9**

General Characteristics of Viruses, Isolation, Cultivation, and Identification of Viruses - Growing Bacteriophages in the Laboratory, Growing Animal Viruses in the Laboratory, Viral Identification; Viral Multiplication- Multiplication of Bacteriophages, Multiplication of Animal Viruses; Fungi – vegetative structures, Life cycle, Zygomycota, Microsporidia, Ascomycota, Basidiomycota, Economic importance of Fungi and Pathogenic fungi. Algae – Characteristics, The life cycle of the unicellular green alga Chlamydomonas. Protozoa- Characteristics, Ameba, Apicomplexa, and Plasmodium life cycle.

**MODULE III OBSERVING MICROORGANISMS THROUGH A MICROSCOPE 7**

Types of Microscopy-Light Microscopy, Two-Photon Microscopy, Scanning Acoustic Microscopy, Electron Microscopy, Confocal Microscopy, Scanned-

Probe Microscopy; Preparation of Specimens for Light Microscopy- Preparing Smears for Staining, Simple Stains, Differential Stains, Special Stains.

**MODULE IV                                  MICROBIAL NUTRITION, GROWTH AND 10 CONTROL**

Bacterial nutrition: Basic nutritional requirements, growth factors, nutritional categories, physical requirements of bacterial growth. Bacteriological media: types (complex, synthetic, differential, enrichment and selective media) and their uses, culture characteristics of bacteria on different media. Bacterial growth: growth kinetics, growth curve. Batch, continuous and synchronous culture. Measurement of growth and influence of environmental factors affecting growth. Control of Microbial Growth, Action of microbial control agents, physical and chemical methods of microbial control.

**MODULE V                                  SCOPE OF MICROBIOLOGY                                  10**

The cycle of matter in nature. Microbial interactions- mutualism, symbiosis, commensalisms, predation, parasitism, amensalism, competition, bioluminescence, biodegradation, biofilms. Cleaning oil spills, microbes in composting, biopesticides, bioremediation, bioleaching, SCP, microbial enzymes and fermented foods. Human diseases and their causative agents. Definition of aeromicrobiology, airborne pathogens, and allergens, Phytopathogenic bacteria: Angular leaf spot of cotton, crown galls, bacterial cankers of citrus. Diseases caused by Phytoplasmas: Aster yellow, citrus stubborn.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Microbiology: An Introduction: Tortora, Funke & Case. 7th edition, 2001
2. Microbiology: Davis, Dulbecco, Eisen and Ginsburg.
3. Introduction to Microbiology: Ross  
General Microbiology: Stainier, Adelberg and Ingraham.

**COURSE OUTCOMES:**

**CO1:** Demonstrate a broad understanding of the diversity and range of microorganisms, the interactions between humans and microorganisms, the role of microorganisms in industrial and environmental processes, and their role in the development of the techniques that underpin modern molecular biology

**CO2:** Demonstrate proficiency in a set of core microbiological and molecular biological technical methods, including both an understanding of the



principles of the methods and their utilization in laboratory settings

**CO3:** Demonstrate familiarity with the risk assessment process, and use this information to operate safely in the laboratory environment

**CO4:** Collect, organize, analyze, evaluate and interpret experimental data using appropriate quantitative, technological and critical thinking skills

**CO5:** Critically evaluate relevant scientific data and literature and comprehend the nature and scope of the scientific literature in microbiology and related areas

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
CO1	H	L	-	-	-	-	-	-	-	-	-	-
CO2	H	L	-	-	-	-	-	-	-	-	-	-
CO3	H	L	-	-	-	-	-	-	-	-	-	-
CO4	H	L	-	-	-	-	-	-	-	-	-	-
CO5	H	L	-	-	-	-	-	-	-	-	-	-

**Note:**L- Low Correlation M -Medium Correlation H -High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of plant and animal biotechnology can help in the development of better crops and livestock that can contribute to good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the ways to create better plant, crop varieties and livestock that can contribute all the levels of life in the earth.

<b>LTE 6105</b>	<b>CELL AND MOLECULAR BIOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To get overview of classes of cells and structural and function aspects of plasma membrane and cell organelle.

**COB2:** To develop skill to understand molecular aspects of cell cycle and cell division.

**COB3:** To get familiar with transcription and translation in details.

**COB4:** To understand the signaling pathways in cell functioning

**COB5:** To understand energy conservation and conversion phenomenon in cells

**MODULE I INTRODUCTION TO CELL 9**

Basic properties of cell, Different classes of cell: Prokaryotic, animal and plant cell. Plasma membrane- structure and function, Chemical composition of membranes, membrane lipids and proteins, fluid mosaic model, Transport across the membranes- diffusion, osmosis, facilitated diffusion, passive and active transport; membrane potential and nerve impulses.

**MODULE II MEMBRANE TRANSPORT 9**

Endoplasmic Reticulum, Golgi complex- glycosylation, Vesicle transport- COPI and COPII; Lysosomes-autophagy; Endocytic pathway- endocytosis and phagocytosis, transport of proteins into peroxisomes, mitochondria and chloroplast;

**MODULE III ENERGY CONVERSION**

Structure of mitochondria and organization of respiratory chain; Proton Pump and ATP generation in mitochondria; Structure of chloroplast and Photosynthesis, photorespiration; Genetic system of mitochondria and chloroplast.

**MODULE IV BASIC GENETIC MECHANISMS 9**

The structure and function of DNA, DNA packaging and Chromosomes, chromatin structure and function, DNA replication mechanisms, DNA damage and repair and homologous recombination and transposable elements, Telomeres, telomerase and end replication. Role of telomerase in aging and cancer.

**MODULE V TRANSCRIPTION AND TRANSLATION 9**

Transcription- Prokaryotic and eukaryotic Transcription- RNA polymerases- general and specific transcription factors- regulatory elements- mechanism of transcription, Transcription termination Post transcriptional modification- splicing- editing- nuclear export of mRNA- mRNA stability; Translation- Genetic code, Mechanism of initiation- elongation and termination- Regulation of translation.

**L – 45; TOTAL HOURS –45****TEXT BOOKS:**

1. Molecular Biology of Cell by Alberts et.al. John Wiley & Sons, 6Ed, 2015
2. The Cell by Cooper. ASM Press, 4Ed, 2007
3. Cell and Molecular Biology by Karp. John Wiley & Sons, 7Ed, 2013
4. Lodish H. F. Cell and Molecular Biology. W.H. Freeman & Co Ltd, 7Ed, 2000.

**COURSE OUTCOMES:****CO1:** Appreciate the basic organization of organisms and living being**CO2:** Understand the machinery of the cell that is ultimately responsible for various daily activities**CO3:** Understand the basic organization of DNA**CO4:** Appreciate the core genetic process of synthesis of mRNA and proteins**CO5:** Acquire knowledge about biological problems that requires engineering expertise to solve them**Board of Studies (BoS) :**9<sup>th</sup>BoS of SLS held on 20.08.2022**Academic Council:**19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	M	H	M	L	L	-	-	-	-	-	-
CO2	H	M	H	M	L	L	-	-	-	-	-	-
CO3	H	M	H	M	L	L	-	-	-	-	-	-
CO4	H	M	H	M	L	L	-	-	-	-	-	-
CO5	H	M	H	M	L	L	-	-	-	-	-	-

**Note:** L - Low Correlation M - Medium Correlation H - High Correlation

SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of cell biology live organisms can help in maintain systems to promote good health and well being.

<b>LSE 6122</b>	<b>LABORATORY I</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3,15</b>	<b>(GENETICS/ MICROBIOLOGY/ CELL AND MOLECULAR BIOLOGY)</b>	<b>0</b>	<b>0</b>	<b>4</b>	<b>2</b>

### **COURSE OBJECTIVES:**

**COB1:** To learn basic techniques in biochemistry, molecular biology, microbiology, plant and animal physiology

**COB2:** To study and to characterize biomolecules, extraction, identification and quantification

**COB3:** To learn the preliminary methods in biochemistry as well microbiology by preparing buffer and adjusting pH.

**COB4:** To estimate various biomolecules by biochemical assays

**COB5:** To estimate the effect of hormones on the biochemical assays

### **EXPERIMENTS**

1. Laboratory safety guidelines
2. To determine an unknown protein concentration by plotting a standard graph of BSA using UV-Vis Spectrophotometer and validating the Beer- Lambert's Law.
3. To prepare an Acetic-Na Acetate Buffer system and validate the Henderson-Hasselbach equation.
4. Effect of temperature on enzyme activity.
5. Preparation of competent cell by calcium chloride method and checking its efficiency
6. Preparation of slides from onion root tip for mitosis.
7. Separation techniques for amino acids and sugar: (a) paper chromatography (b) thin layer chromatography.
8. Separation of proteins by native and SDS-PAGE.
9. Isolation & Purification of genomic DNA from bacteria.
10. Isolation & Purification of plasmid DNA.
11. Isolation of RNA.
12. Agarose gel electrophoresis of chromosomal & plasmid DNA.
13. Restriction Digestion of chromosomal & plasmid DNA.
14. Isolation of DNA fragments from agarose gel.
15. Single and double radial immunodiffusion.
16. Double diffusion, Immuno-electrophoresis and Radial Immunodiffusion.
17. Blood group mapping.
18. Competent cell preparation.

19. Polymerase Chain Reaction.  
20. Isolation of Genomic DNA from Plants.

**P – 60; TOTAL HOURS –60**

**TEXTBOOKS:**

1. Michel R. G and Sambrook J. Molecular Cloning- A laboratory manual. Cold spring harbor laboratory press, 2012.
2. Laboratory Exercises in Microbiology, Fifth Edition by Harley–Prescott, The McGraw–Hill Companies, 2002.
3. Wilson K and Walker J, Principles and Techniques in Practical Biochemistry, 5th Ed., Cambridge University Press, 2000.
4. Holtzhauser M, Basic Methods for the Biochemical Lab, Springer, 2006.
5. Nigam, Lab Manual in Biochemistry: Immunology and Biotechnology, Tata McGraw-Hill Education, 2007.

**COURSE OUTCOMES:**

**CO1:** To understand the importance of laboratory safety and standard operating procedures of common laboratory equipment

**CO2:** The students will be trained in performing routine molecular microbial techniques

**CO3:** Students will be able to isolate culture and identify microbes and also to efficiently use a light microscope

**CO4:** The students will be trained in studying both plant and animal physiology

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
CO1	H	M	H	M	L	L	-	-	-	-	-	-
CO2	H	M	H	M	L	L	-	-	-	-	-	-
CO3	H	M	H	M	L	L	-	-	-	-	-	-
CO4	H	M	H	M	L	L	-	-	-	-	-	-
CO5	H	M	H	M	L	L	-	-	-	-	-	-

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of cell biology live organisms can help in maintain systems to promote good health and well being.

**SEMESTER II**

<b>GEE 6202</b>	<b>RESEARCH METHODOLOGY AND IPR</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4, 9, 11 &amp; 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

Students will be trained to

**COB1:** Basic concepts of Research.

**COB2:** Select and Define a research problem

**COB3:** Analyze and Interpret the Results

**COB4:** write Scientific and Technical reports & thesis

**COB5:** Apply the Copyrights, Patents and Intellectual Property Rights.

**MODULE I INTRODUCTION TO RESEARCH METHODOLOGY 9**

Research: Objectives, Motivation and types - Approaches, Significance of Research, Research process, Criteria of good research, Problems encountered by researchers - Introduction to ethics, scientific conduct and misconduct, misconduct and why it occurs, fabrication, authorship issues, The investigation and punishment of scientific misconduct (Erratum).

**MODULE II RESEARCH FORMULATION AND DESIGN 9**

Defining and formulating the research problem, selecting the problem, necessity of defining the problem, importance of literature review in defining a problem, literature review-primary and secondary sources, reviews, monograph, patents, research databases, web as a source, identifying gap areas from literature and research database, development of working hypothesis. Definition and importance of Journal Impact factor, Cite Scores and Citation Indexes.

**MODULE III DATA COLLECTION, ANALYSIS AND INTERPRETATION OF DATA 9**

Observation and Collection of data, methods of data collection, sampling methods, data processing, analysis strategies and tools, data analysis with statistical tools (Sigma STAT, SPSS student, ANOVA), hypothesis testing. Importance and scientific methodology in recording results, importance of negative results, conceptions of error of measurement - absolute and relative errors, true score theory and generalisability theory. Measures of central tendency – mean median and mode.

**MODULE IV SCIENTIFIC AND TECHNICAL WRITING 9**

Different types of scientific and technical publications in the area of research - Technical writing skills for report, synopsis and thesis – organisation of contents and layout of the research reports, oral presentation, mechanics of writing a research report, precautions for writing research reports, conclusions. Preparing papers for international journals - software for paper formatting like LaTeX/MS Office, Grammarly - reference management software – Mendeley and detection of similarity index / plagiarism by Turnitin.

**MODULE V INTELLECTUAL PROPERTY RIGHTS 9**

The concept, Intellectual Property system in India, development of TRIPS complied regime in India, Patents Act, 1970, Trade Mark Act, 1999, The Designs Act, 2000, Commercialization, Copy Right, Royalty, Trade related aspects of Intellectual Property Rights (TRIPS); Geographical indications, Industrial designs, Enforcement of Intellectual Property Rights, Function of UNSECO in IPR maintenance. Patents, Patentable subject matter, Rights conferred, Exceptions, Term of protection, Conditions on Patent applicants, Process patents.

**L – 45; TOTAL HOURS – 45**

**TEXT BOOKS:**

1. Cooper Donald R, Schindler Pamela S and Sharma JK., 2012. “Business Research Methods”, Tata McGraw Hill Education, 11e.
2. Kothari C.R., “Research Methodology, Methods and Techniques”, Wiley Eastern Ltd., New Delhi, 1991.

**REFERENCES:**

1. Anthony, M., Graziano, A.M. and Raulin, M.L., 2009. Research Methods: A Process of Inquiry, Allyn and Bacon.
2. Garg, B.L., Karadia, R., Agarwal, F. and Agarwal, U.K., 2002. An introduction to Research Methodology, RBSA Publishers.
3. Day, R.A., 1992. How to Write and Publish a Scientific Paper, Cambridge University Press.
4. Sinha, S.C. and Dhiman, A.K., 2002. Research Methodology, Ess Ess Publications. 2 volumes .
5. Essentials of Research Design and Methodology Geoffrey R. Marczyk, David DeMatteo, David Festinger, 2005. John Wiley & Sons Publishers, Inc
6. Biochemical Calculations: How to Solve Mathematical Problems in General Biochemistry, 2nd Edition, Irwin H. Segel, 1976. John Wiley & Sons Publishers, Inc
7. R Arora. Encyclopaedia of Research Methodology in Biological Sciences., Anmol Publishing, 2004.

8. CoghilM.andGardsonL.R.,TheACSStyleGuideEffectiveCommunicationofScientificInformation,3rdEdn.,OxfordUniversityPress,2006.

### COURSE OUTCOMES:

The students will be able to

**CO1:** recognize the basic concepts of research and its methodologies

**CO2:**select and define appropriate research problem and parameters

**CO3:** apply packages for data collection, analyze and interpretation of data into reports.

**CO4:** write scientific report as journal article, thesis and technical proposal for funding.

**CO5:** propose research findings as publications, copyrights, trademarks and IPR.

### Board of Studies (BoS) :

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

### Academic Council:

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO1 1	PO 12	PSO 1	PSO 2	PSO 3
CO1													M		
CO2			H					M						M	
CO3		H			M										
CO4													H		M
CO5										H			H		

**Note:** L- Low Correlation M - Medium Correlation H -High Correlation

SDG 4 : Quality Education

SDG 9 : Build resilient infrastructure, promote inclusive and sustainable industrialization and foster innovation

SDG 11 : Sustainable Cities and Communities

SDG 15 : Life on Land

Statement: The understanding of concepts of high quality research, innovative thinking, knowledge on sustainable development and service to the society and mankind through quality research.



<b>LSE 6221</b>	<b>ENVIRONMENTAL AND MEDICAL</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>MICROBIOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To learn the basic principles of environment and medical microbiology and be able to apply these principles to understanding and solving problems in water quality.

**COB2:** To become familiar with current research in environmental microbiology and its applications.

**COB3:** Develop knowledge of microbial organisms and their relevance of infectious diseases.

**COB4:** Understand the principles of prevention and treatment of pathogenic microorganism infection in humans.

**COB5:** Understand the principles of pathological progression of infections and mechanism of drug action.

<b>MODULE I</b>	<b>INTRODUCTION TO ENVIRONMENTAL MICROBIOLOGY</b>	<b>9</b>
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Environmental Microbiology: Waste treatment - Wastes - types and characterization. Treatment of solid wastes - composting, vermiform composting, silage, pyrolysis and saccharifications. Treatment of liquid wastes - primary, secondary (anaerobic and aerobic) - trickling, A-9 21 activated sludge, oxidation pond, and oxidation ditch-tertiary - disinfection.

<b>MODULE II</b>	<b>DEGRADATION USING MICROBES</b>	<b>9</b>
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Degradation of of Xenobiotic compounds: Simple aromatics, chlorinated polyaromatic petroleum products, pesticides and surfactants. Biodeterioration of materials - paper, leather, wood, textiles and paint. Metal corrosion - Bioaccumulation of heavy metals. Biofouling and Bioleaching.

<b>MODULE III</b>	<b>MICROBIAL ASSESMENT OF WATER</b>	
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Physical, chemical and microbial assessment of water and potability test for water. Physical and chemical - colour, pH, alkalinity, acidity, COD, BOD, anions and cations. Microbiological - MPN index - presumptive, completed and confirmatory tests.

<b>MODULE IV</b>	<b>EPIDEMIOLOGY AND PATHOGENECITY OF DISEASES</b>	<b>9</b>
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Epidemiology, pathogenicity and treatment of diseases caused by Haem. influenzae, C. diphtheriae, E. coli, Pseudomonas, Bacillus anthracis, Cl. tetani,

Cl. walchi, Leptospira icterohaemorrhagiae, N. gonorrhoea, Mycoplasma, Compylobacter. Rickettsia -cox, burneti, Chlamidiae -trachoma, New Bacterial diseases- Helicobacter, Lyme diseases, Legionella.

## MODULE V INFECTION & DRUGS 9

Infections in Vulnerable hosts, antimicrobial drugs, Bauer Kirby test, Broth dilution test, minimum inhibitory and lethal concentrations. Mechanism of action and activity spectrum of penicillin, streptomycin, tetracycline, sulfonamides, rifampicin, polymyxin - B, Amphotericin – B. Drug resistance.

**L – 45; TOTAL HOURS –45**

### TEXT BOOKS:

1. Ashok Kumar Chauhan, Ajit Varma, Microbes: Health and Environment (Microbiology), Anshan Publishing; 1 edition, 2006.
2. Ian Pepper Charles Gerba Jeffrey Brendecke, Environmental Microbiology, Elsevier, 1995.
3. Patrick Murray Ken Rosenthal Michael Pfaller, Medical Microbiology, Elsevier, 2015

### COURSE OUTCOMES:

**CO1:** The concept and principles of waste management, types and uses

**CO2:** The concept and principles of physical, chemical and microbial analysis of water bodies

**CO3:** The concept and principles of disease epidemiology

**CO4:** The concept and principles infectious diseases and development of vaccines

**CO5:** The concept and principles of infection spread and development of drugs

### Board of Studies (BoS) :

9<sup>th</sup>BoS of SLS held on 20.08.2022

### Academic Council:

19<sup>th</sup> AC held on 29.09.2022

CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of microbial genetics we can understand the genetic basis of underlying diseases, that will help in good health and well being

SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to microbial geneticists that gives knowledge about relation with all the levels of life in the earth.

<b>LSE 6241</b>	<b>BIOINFORMATICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>4</b>	<b>0</b>	<b>0</b>	<b>4</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the programming languages applied in computational biology.

**COB2:** To understand the methods and applications for sequence analysis, Phylogenetics and Protein modelling.

**COB3:** Understanding of alignment tools and techniques

**COB4:** Understanding of Phylogenetic analysis methods

**COB5:** Understanding of Predictive models and methods

**MODULE I INTRODUCTION TO BIOINFORMATICS AND DATABASES 10**

Introduction- scope- Historical account, Database Management Systems- Primary and Secondary databases- Genbank-EMBL- DDBJ -Sequence retrieval, file formats and conversion tools, metabolic pathway databases Primer designing-tools and applications.

**MODULE II MOLECULAR SEQUENCE ALIGNMENT 13**

Pair wise Alignment- Global Alignment- Local Alignment- Multiple Sequence Alignment methods. Phylogenetic Analysis: Construction of Phylogenetic trees - Distance Methods- Maximum Parsimony Method- Maximum likelihood method.

**MODULE III OMICS, TYPES, METHODS AND APPLICATIONS 13**

Omics-Importance and applications Functional genomics- assigning the function, applications-Glycomics-databases and tools used and applications Lipidomics-role in disease and stress predictions, tools and applications Proteomics, metabolomics-tools and applications in bioinformatics

**MODULE IV MOLECULAR MODELING AND DRUG DESIGNING 12**

Introduction to Protein Structure Prediction- Rational drug discovery- Recent advances in drug design methodologies- Structure-based drug design- Drug-receptor interactions- Structure-Activity Relationships.

**MODULE V RECENT ADVANCES IN COMPUTATIONAL BIOLOGY 12**

Metagenomics-methods, prediction of new species, NGS-methods, applications and advantages, Cancer informatics-diagnostics and treatment of cancer Gene ontology-methods, databases, classifications, systems biology-introduction, databases and applications, Microarray-Types, methods, applications, databases and tools

**L – 60; TOTAL HOURS –60**

**TEXT BOOKS:**

1. Claverie J. M, Notredame C. Bioinformatics, Wiley Publishing, Inc. 2003
2. Dan.E.K, Michael L. R. Fundamental concepts in Bioinformatics. 1st Ed., Pearson Education. 2006.
3. David Mount. Bioinformatics: Sequence and Genome Analysis. CSHL Publisher, 2001.
4. Andreas D. Baxevanis & B.F. Francis Ouellette. Bioinformatics. A Practical Guide to the analysis of genes and Proteins, John Wiley & Sons, UK, 1998.
5. Higgins. D and Taylor W. Bioinformatics Sequence, Structure databanks. OUP Oxford, 2000.
6. Silberschatz A, Korth H. F, Sudarshan S. Database System Concepts. 3rd Ed., mcgraw-Hill, 2010.

**COURSE OUTCOMES:**

On completion of the course, the students will be able to understand the

**CO1:** history and importance of bioinformatics

**CO2:** Biological Databases and Data Retrieval,

**CO3:** Molecular Sequence Alignment

**CO4:** Grasp the principle involved Gene Prediction

**CO5:** Phylogenetic Analysis and Molecular Modeling and Drug Designing.

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-			-	-	-	
CO2	H	L	-	-	-	-			-	-	-	
CO3	H	L	-	-	-	-			-	-	-	
CO4	H	L	-	-	-	-			-	-	-	
CO5	H	L	-	-	-	-			-	-	-	

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of bioinformatics we can carry out data mining gene and protein expression patterns and modelling cellular interactions and processes, that will help in good health and well being

### SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to computer scientists in bioinformatics gives knowledge about relation with all the levels of life in the earth.

<b>LSE 6222</b>	<b>BIOPROCESS AND</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3,15</b>	<b>FERMAENTATION TECHNOLOGY</b>	<b>4</b>	<b>0</b>	<b>0</b>	<b>4</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the design and operation of fermenters and types of fermentation processes

**COB2:** To acquire knowledge about the formulation of medium and its prerequisites

**COB3:** To interpret stoichiometry and energetics of cell growth and product formation

**COB4:** To analyze the modes of operation of bioreactor and its design equations

**COB5:** To evaluate the kinetics and mechanism of microbial growth by using various models

**MODULE I BIOPROCESS TECHNOLOGY 12**

Design features of bioreactors / fermenters, Fundamentals of bioprocess technology, Principles underlying product formation, Principles underlying product recovery and purification, Large scale production of fermentation products, Fermentation kinetics: Reaction kinetics, Scale up of fermentation process, Downstream processing, Biosynthetic pathways for some secondary metabolites.

**MODULE II MODELING AND DESIGN OF FERMENTATION PROCESSES 12**

Principles of model building for biotechnological processes, modeling of recombinant systems. biomass growth and product formation, Kinetics of substrate utilization, inhibition on cell growth and product formation. Design and operation of continuous cultures, chemostat in series, batch and fed batch cultures, total cell retention cultivation, Case studies on Production of green chemicals, algal biofuels, recombinant Insulin. Case studies should deal with medium design, reactor design & process optimization etc.

**MODULE III DOWNSTREAM PROCESSING 12**

Introduction-downstream processing, biomolecules and bioprocesses, biomass removal and disruption technique- centrifugation, sedimentation, flocculation, microfiltration, sonication, Homogenizers, chemical lysis, enzymatic lysis , pretreatment and stabilisation of bioproducts.

**MODULE IV SEPARATION AND ISOLATION 12**

Unit operations for solid-liquid separation - filtration and centrifugation. Membrane

based purification: Ultrafiltration; Reverse osmosis; Dialysis ; Diafiltration ; Pervaporation; Perstraction Adsorption and chromatography: size, charge, shape, hydrophobic interactions, Biological affinity; Process configurations (packed bed, expanded bed, simulated moving beds).

## **MODULE V PRODUCT PURIFICATION AND FORMULATION 12**

Ammonium Sulfate-precipitation, solvent), Chromatography, principles, instruments and practice, adsorption, reverse phase, ionexchange, size exclusion, hydrophobic interaction, bioaffinity and pseudo affinity chromatographic techniques. Crystallization, drying and lyophilization in final in product formulation.

**L – 60; TOTAL HOURS –60**

### **TEXT BOOKS:**

1. Shuler, M.L. and Kargi, F. Bioprocess Engineering : Basic concepts, 2<sup>nd</sup> ed., Prentice-Hall, 2002.
2. Doran Pauline M, Bioprocess Engineering Principles, Academic Press, 1995 to the Analysis of Genes and Proteins, 2<sup>nd</sup> ed., John Wiley, 2002
3. Nielsen, J. and Villadsen, J. "Bioreaction Engineering Principles". Springer, 2007.
4. P.A. Belter, E.L. Cussler And Wei-Houhu – Bioseparations – Downstream Processing For Biotechnology, Wiley Interscience Pub. (1988).
5. Blanch, H.W and Clark D.S., "Biochemical Engineering", Marcel Dekker, 1997.
6. R.O. Jenkins, (Ed.) – Product Recovery In Bioprocess Technology – Biotechnology By Open Learning Series, Butterworth-Heinemann (1992).

### **COURSE OUTCOMES:**

**CO1:** Understand the design and operation of fermenter and types of fermentation process

**CO2:** Acquire knowledge about formulation of medium and its prerequisites•

**CO3:** Interpret stoichiometry and energetics of cell growth and product formation

**CO4:** Analyze the modes of operation of bioreactor and its design equations

**CO5:** Evaluate the kinetics and mechanism of microbial growth by using various models

### **Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

### **Academic Council:**

19<sup>th</sup> AC held on 29.09.2022



	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of bioprocesswe can carry out data mining gene and protein expression patterns and modelling cellular interactions and processes, that will help in good heath and well being.

### SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to computer scientists in fermentation gives knowledge aboutrelation with all the levels of life in the earth

<b>LSE 6223</b>	<b>LABORATORY II(MEDICAL</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>MICROBIOLOGY/ BIOINFORMATICS/ BIOPROCESS TECHNOLOGY)</b>	<b>0</b>	<b>0</b>	<b>4</b>	<b>2</b>

**COURSE OBJECTIVES:**

**COB1:** To inculcate skill of plasmid construction, mappings and analysis.

**COB2:** To train the students in protein structure prediction, sequence homology mapping

**COB3:** To gain the working knowledge of enzyme immobilization used in industries

**COB4:** To elucidate the importance of microorganisms in biomedical and industrial effluents treatment

**COB5:** To gain hands-on skills to perform upstream and downstream processes

**PRACTICALS**

List of Experiments:

1. Plasmid Construction/Restriction Mapping
2. PCR Primer Designing
3. Sequence Retrieval and Format Conversion
4. Homology Search/ Multiple Sequence Alignment
5. Motif finding in DNA and Protein Sequences
6. Protein Secondary Structure Prediction
7. Sauerkraut fermentation
8. Effect of substrate concentration on biomass yield
9. Solvent extraction techniques for product recovery
10. Enzyme immobilized by alginate gel method
11. Isolation of antibiotic resistant mutants by chemical mutagenesis
12. Physical analysis of sewage/industrial effluent by measuring total solids, total dissolved solids and total suspended solids
13. Determination of indices of pollution by measuring BOD/COD of different effluents
14. Microbial dye decolourization/adsorption
15. Comparison of ethanol production using various Organic wastes /raw Material [Free cells/ immobilized cells.

**P – 60; TOTAL HOURS –60**

**REFERENCES:**

1. Michael Doyle, Francisco Diez-Gonzalo, Food Microbiology: Fundamentals and Frontiers , American Society for Microbiology, 2013

2. Baxevanis A. D, Ouellette B. F. F. Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins. 3rd edition Wiley, John & Sons, Incorporated, 2004
3. Rashidi H, Buehler L. K. Bioinformatics Basics: Applications in Biological Science and Medicine. 2nd Ed., CRC Press, 2005.

### COURSE OUTCOMES:

**CO1:** Students will be trained with various soft skills/tool used in bioinformatics

**CO2:** They will also become skilled in analyzing and interpolating data from PCR primer designing to structure predictions.

**CO3:** They will gain the working knowledge of enzyme immobilization used in industries

**CO4:** The students will be able to apply various microorganisms to treat biomedical and industrial effluents.

**CO5:** Students will gain hands-on skills to perform upstream and downstream processes.

### Board of Studies (BoS) :

9<sup>th</sup>BoS of SLS held on 20.08.2022

### Academic Council:

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of techniques we can carry out data mining gene and protein expression patterns and modelling cellular interactions and processes, that will help in good health and well being.

### SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to lab techniques gives knowledge about relation with all the levels of life in the earth.

**SEMESTER III**

<b>LSE 7121</b>	<b>INDUSTRIAL, FOOD, AND</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>AGRICULTURAL MICROBIOLOGY</b>	<b>4</b>	<b>0</b>	<b>0</b>	<b>4</b>

**COURSE OBJECTIVES:**

**COB1:** To explore about food process and technology.

**COB2:** To get an overview of the processing of various types of food

**COB3:** To expose themselves to storage and handling of food and food products

**COB4:** To get an understanding the of fermentation process and its industrial application

**COB5:** To understand the properties and reactions of various microorganisms and their various development strategies with a clear understanding in terms of agricultural and food Biotechnology.

<b>MODULE I</b>	<b>INDUSTRIAL BIOTECHNOLOGY- AN</b>	<b>12</b>
	<b>INTRODUCTION</b>	

Exploitation of microorganisms and their products, screening, strain development strategies, immobilization methods, fermentation media, raw material used in media production, antifoaming agents, buffers, downstream processing.

<b>MODULE II</b>	<b>FERMENTATION</b>	<b>11</b>
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Fermentation equipment and its uses, fermentor design, Types of fermentors and fermentations- single, batch, continuous, multiple, surface, submerged and solid state.

<b>MODULE III</b>	<b>MICROORGANISMS IN AGRICULTURE AND</b>	<b>14</b>
	<b>PLANT DISEASES</b>	

Microorganisms as biofertilizers (Rhizobial, Cyanobacterial, Mycorrhizal, Azotobacter): production and application of Microbial biopesticides, recombinant pesticides, GMO and their impact. Microbial diseases of crops: transmission of pathogens, Citrus canker, little leaf of brinjal, red rot of sugarcane, brown rot of potato, black rot, mosaic virus, tomato spot, early and late blight, wilt disease. Control of plant diseases.

**MODULE IV                      MICROORGANISMS IN FOOD                      13**

Microorganisms important in food microbiology – molds, yeasts, bacteria, principles of food preservation – high and low temperatures, drying, chemical preservatives, food additives. Food spoilage and food borne infections, general principles underlying food spoilage and contamination, canned food spoilage, spoilages of vegetables, fruits, meat and meat products, milk and milk products, fish, seafood and poultry.

**MODULE V                      FOOD PRODUCTS                      10**

Food produced by microbes: bread, cheese, fermented dairy products, microbial cells as food – single cell proteins, mushroom, fermented Indian foods.

**L – 60; TOTAL HOURS –60**

**TEXT BOOKS:**

1. Fennema Karrel, Principles of Food Science, Vol-I, 1985.
2. Modern Dairy Products, Lampert LH; 1970, Chemical Publishing Company
3. Casida, L.E. "Industrial Microbiology", New Age International (P) Ltd, 1968.
4. Prescott, S.C. and Cecil G. Dunn, "Industrial Microbiology", Agrobios (India), 2005.
- 5.

**COURSE OUTCOMES:**

**CO1:** Know the equipments and their preliminary operations in food processing

**CO2:** Understand the physical principles involved in the food processing techniques and the equipments used.

**CO3:** Equip themselves to trouble shoot the problems arises in drying process to preserve the foods

**CO4:** Familiarize with preservation of foods at low temperature, prevention of growth of microorganisms in food products

**CO5:** Familiarize with alternative sources of food

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	-	-	-	-	-	-
CO2	H	L	-	-	-	-	-	-	-	-	-	-
CO3	H	L	-	-	-	-	-	-	-	-	-	-
CO4	H	L	-	-	-	-	-	-	-	-	-	-
CO5	H	L	-	-	-	-	-	-	-	-	-	-

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of agriculture we can carry out data mining gene and protein expression patterns and modelling cellular interactions and processes, that will help in good health and well being

### SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to microbiology gives knowledge about relation with all the levels of life in the earth

<b>LSE 7122</b>	<b>IMMUNOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>4</b>	<b>0</b>	<b>0</b>	<b>4</b>

**COURSE OBJECTIVES:**

**COB1:** An understanding of immunity, history of immunology, cells and organ involved in immune system

**COB2:** An understanding of Antigen-Antibody interaction

**COB3:** An understanding of cytokines and complement system involved in immune system

**COB4:** An understanding of receptors, MHC class of molecules and regulation of immune response

**COB5:** Learning different techniques in immunology

**MODULE I OVERVIEW OF IMMUNE SYSTEM 12**

Innate, adaptive and Comparative Immunology, Immune dysfunction and its consequences, Cells & Tissues of Immune System: Hematopoiesis, Apoptosis and Necrosis, systemic function of Immune system, organs of immune systems, Lymphoid cells and organs Evolutionary comparison. Cytokines- Properties of Cytokines, Cytokine Receptors, Cytokine Antagonists, Cytokine Secretion by TH1 and TH2 Subsets, Cytokine-Related Diseases, Therapeutic Uses of Cytokines and Their Receptors, Cytokines in Hematopoiesis.

**MODULE II MOLECULAR IMMUNOLOGY 12**

Immunogenicity Versus Antigenicity, Factors that influence immunogenicity, Epitopes, Haptens and the Study of Antigenicity, Pattern-Recognition Receptors, drug allergies-when medicine become immunogens, Molecular structure of antibody, Obstacles to Antibody Sequencing, Immunoglobulin Fine Structure, Antibody-Mediated Effector Functions, Antibody Classes and Biological Activities, Antigenic Determinants on Immunoglobulins, The B-Cell Receptor, The Immunoglobulin Superfamily, Monoclonal Antibodies.

**MODULE III ORGANIZATION AND EXPRESSION OF IMMUNOGLOBULIN GENES 12**

Genetic Model Compatible with Ig Structure, Multigene Organization of Ig Genes, Variable-Region Gene Rearrangements, Mechanism of Variable-Region DNA Rearrangements, Generation of Antibody Diversity, Class Switching among Constant-Region Genes, Expression of Ig Genes,

Synthesis, Assembly, and Secretion of Immunoglobulins, Regulation of Ig-Gene Transcription, Antibody Genes and Antibody Engineering.

**MODULE IV ANTIGEN PROCESSING AND PRESENTATION 12**

General organization and inheritance of the major histocompatibility complex (MHC), MHC molecules and genes, detailed genomic map of MHC genes, cellular distribution of MHC molecules, regulation of MHC expression, MHC and immune responsiveness, MHC and disease susceptibility self-MHC restriction of T cells, role of antigen-presenting cells, evidence for two processing and presentation pathways, endogenous antigens: the cytosolic pathway, exogenous antigens: the endocytic pathway presentation of nonpeptide antigens

**MODULE V GENERATION OF T AND B CELL RESPONSE 12**

T-Cell Receptor, Early Studies of the T-Cell Receptor and  $\alpha\beta$  and  $\gamma\delta$  T-Cell Receptors: Structure and Roles, Organization and Rearrangement of TCR Genes, T-Cell Receptor Complex: TCR-CD3, T-Cell Accessory Membrane Molecules, Three-Dimensional Structures of TCR-Peptide-MHC Complexes, Alloreactivity of T Cells, T-Cell Maturation and the Thymus, Thymic Selection of the T-Cell Repertoire,  $T_H$ -Cell Activation, T-Cell Differentiation, Cell Death and T-Cell Populations Peripheral  $\gamma\delta$  T-Cells, B-Cell Maturation, B-Cell Activation and Proliferation, The Humoral Response, In Vivo Sites for Induction of Humoral Responses, Germinal Centers and Antigen-Induced B-Cell Differentiation, Regulation of B-Cell Development, Regulation of the Immune Effectors Response.

**L – 60; TOTAL HOURS –60**

**TEXT BOOKS:**

1. Kuby, RA Goldsby, Thomas J. Kindt, Barbara, A. Osborne Immunology, 6th Edition, Freeman, 2002.
2. Brostoff J, Seaddin JK, Male D, Roitt IM., Clinical Immunology, 6th Edition, Gower Medical Publishing, 2002.
3. Janeway et al., Immunobiology, 4th Edition, Current Biology publications., 1999.
4. Paul, Fundamental of Immunology, Lippincott Williams & Wilkins; 4th edition, 1999.



**COURSE OUTCOMES:**

**CO1:** be introduced to the science of immunology and a detailed understanding of various types of immune cells, immune systems and their classification, structure.

**CO2:** They will also become skilled to be able to analyze and interpolate data starting from PCR primer designing to structure predictions.

**CO3:** have an understanding of antibody structure, the origin of variations in its structure and role in imparting immunity.

**CO4:** get a thorough understanding for the mechanisms involved in mounting the immune response. .

**CO5:** edge of the cellular and molecular basis for autoimmune disease and allergies.

**Board of Studies (BoS) :**9<sup>th</sup>BoS of SLS held on 20.08.2022**Academic Council:**19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of immunology we can understand interactions and processes, that will help in good health and well being.

**SDG15: Life on Earth**

Statement: This course makes aware of the range of technologies available to computer scientists in immunology gives knowledge about relation with all the levels of life in the earth

<b>LSE 7123</b>	<b>LABORATORY III</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>(INDUSTRIAL, FOOD AND AGRICULTURAL MICROBIOLOGY/ IMMUNOLOGY)</b>	<b>0</b>	<b>0</b>	<b>4</b>	<b>2</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the principles of the underlying the properties and reactions of various microorganisms.

**COB2:** To learn the basic techniques in immunology and be able to apply these techniques to understanding and solving problems in antigen and antibody interactions.

**COB3:** To understand the various development strategies with a clear understanding in terms of food and industrial microbiology Biotechnology and to update students' knowledge of new developments in biology of industrial relevance.

**COB4:** To become familiar with current research in immunotechniques and its applications in various disease diagnosis

**COB5:** To become familiar with the food testing techniques

**PRACTICALS**

1. Isolation of food poisoning bacteria from contaminated foods and from fermented milk products-yogurt, curd
2. Extraction and detection of aflatoxins from contaminated food products
3. Preservation of potato/onion by UV radiation
4. Reductase test for milk – Methylene Blue/ Resazurin.
5. Microbial production, extraction, purification and confirmation of alpha amylase/ Lipase.
6. Physical analysis of sewage/industrial effluent by measuring total solids, total dissolved solids and total suspended solids
7. Blood group mapping
8. Double diffusion, Immuno-electrophoresis and Radial Immuno diffusion.
9. Preparation of antigens from pathogens
10. Antibody titre by ELISA method.
11. ELISA for detection of antigens and antibodies-DOT ELISA
12. Immunoblotting and Immunofluorescence technique

**P – 60; TOTAL HOURS –60**

**REFERENCES:**

1. Martin Adams, Maurice Boss, Food Microbiology. 2nd Edition, 2000.
2. Banwart George J, Basic Food Microbiology, Springer, 1989.
3. Michael Doyle, Francisco Diez-Gonzalo, Food Microbiology: Fundamentals and Frontiers, American Society for Microbiology, 2013
4. V. K. Joshi, Biotechnology: Food Fermentation Microbiology, Biochemistry and Technology. Volume 2, 2009.
5. J. B. Prajapati, Fundamentals of Dairy Microbiology, Nadiad Akta Prakashan, 1995.
6. Brian J. Wood, Microbiology of Fermented Foods. Volume II and I, Springer US, 1998.
7. John C. Ayres. J. Orwin Mundt. William E. Sandinee, Microbiology of Foods by. W. H. Freeman and Co., 1980.
8. Rose et al., Manual of Clinical laboratory Immunology, 6th Ed ASM Publications, 2002.
9. Lefkovic and Pernis. Immunological methods. Academic Press, 1978.
10. Hudson L. and Hay F.C. Practical Immunology. Black Well publishers, 1989

**COURSE OUTCOMES:**

**CO1:** Know the equipments and their preliminary operations in food processing in industrial settings

**CO2:** Understand the physical principles involved in the food processing techniques and the equipments used.

**CO3:** The concept and principles of physical, chemical and microbial analysis of water bodies

**CO4:** They will also learn to perform the qualitative and quantitative analysis using antibodies.

**CO5:** Understand the antigenic expression by probing specific antibodies

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of immunology we can understand interactions and processes, that will help in good health and well being

SDG15: Life on Earth

Statement: This course makes aware of the range of technologies available to computer scientists in immunology gives knowledge about relation with all the levels of life in the earth

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<b>LSE 7211</b>	<b>PROJECT WORK (PHASE 1)</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
		<b>0</b>	<b>0</b>	<b>12</b>	<b>4</b>

**COURSE OBJECTIVES:**

**COB1:**To learn and experiments handled on their own by the students to exhibit their capacity in executing a project work and provide a fruitful solution to a research problem or improving the health and wealth of human beings in the field of Microbiology.

**GENERAL GUIDELINES:**

- ❖ At post-graduate level, project work shall be carried out by the student individually
- ❖ Student shall select a project topic of his/her interest relevant to Microbiology and approach any faculty member of the School of Life Sciences with expertise in that field and get his willingness to supervise the project.
- ❖ Students are permitted to carry out their project in an Industry / Research organization, with the approval of the Dean of the School of Life Sciences. In such cases, the project work shall be jointly supervised by a faculty of the school and a professor/ Scientist from the organization. Proper permission and approvals should be obtained from the industry and documented.
- ❖ The information related to the proposed topic and the faculty member willing to act as a guide shall be informed to the project coordinator within 15 days from the commencement of the semester.
- ❖ Supervisor identified by the student shall be approved by the dean the of School of Life Sciences considering the guidelines followed in the School of Life Sciences to allot supervisor for student projects.
- ❖ The project coordinator, in consultation with the Professor in-charge shall give initial approval to start the project.
- ❖ A project review team comprising minimum of two senior faculty members of the department preferably doctorates shall be

appointed by the Dean of the School of Life Sciences.

- ❖ Project review schedules, weightage for each review, and rubrics for evaluation will be prepared by the project coordinator in line with the academic calendar and informed to the students in advance. A minimum of three reviews shall be conducted to evaluate the progress of the students. All the members of the review committee shall evaluate the students individually and the mean value shall be taken for grading.
- ❖ Students should meet the supervisor periodically and attend the review committee meetings for evaluating the progress. Proper documents shall be maintained by the supervisor to ensure the attendance and progress of the students.
- ❖ In the project phase I, students are expected to identify a suitable topic, draw the need for present study and scope of the investigation, review at least 25 journal papers in the related field, formulate the experimental/analytical methodology and conduct preliminary studies.
- ❖ At the end of project work phase I, students should submit a report based on the preliminary studies and the future work to be carried out.

### **COURSE OUTCOMES:**

Students will be able to

**CO1:** Apply their practical knowledge and skill in Microbiology with a specialization in solving real time problems

**CO2:** Prepare an appropriate documentation

**MOOC COURSE****L T P C**

0 0 0 0

**COURSE OBJECTIVES:**

**COB1:** To learn the basic principles and concepts of the topic in which a project work is undertaken by the student.

**GENERAL GUIDELINES:**

- ❖ Students shall identify a MOOC course related to his/her project topic in consultation with the project supervisor.
- ❖ Student shall register for a MOOC course with minimum two credit offered by any recognized organization during the project phase I.
- ❖ Selection and completion of MOOC course by the students shall be endorsed by Head/Dean of the Department.

**COURSE OUTCOMES:**

Students will be able to

**CO1:** Familiarize the basic principles and concepts related to the topic of his/her project work.

**CO2:** Utilize the knowledge gained in the field of study to perform literature review with ease.

**CO3:** Formulate the experimental / analytical methodology required for the project work

<b>LSE 7124</b>	<b>INDUSTRY INTERNSHIP</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
		<b>0</b>	<b>0</b>	<b>2</b>	<b>2</b>

**GENERAL GUIDELINES:**

- It is one credit for four weeks of internship.
- Internship shall be of not less than two weeks duration and shall be
- organized by the Dean of the Department.
- Students should choose preferably, government agencies/ Central Government research Institutes/DBT/ICAR/IISER/CSIR/ICMR /IIT's/ NITs/major industries in their specialization to do their internship
- At the end of the industrial internship, the student shall submit a certificate and feedback from the organization. Students should also submit a brief report.
- The evaluation will be made based on this report and a Viva-Voce
- Examination, conducted internally by a Departmental Committee
- constituted by the Dean of the School of Life Sciences.



**SEMESTER IV**

<b>LSE 7211</b>	<b>PROJECT WORK (PHASE 2)</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
		0	0	32	16

**COURSE OBJECTIVES:**

**COB1:**To learn and experiments handled on their own by the students to exhibit their capacity in executing a project work and provide a fruitful solution to a research problem or improving the health and wealth of human beings in the field of Microbiology.

**GENERAL GUIDELINES:**

- ❖ Project work phase II is a continuation of phase I following the same guidelines.
- ❖ The project co-ordinator shall arrange to conduct three reviews to ascertain the progress of the work and award the marks based on the performance.
- ❖ Detailed experimental investigation / in-depth analytical study /
- ❖ Preparation of specimens / testing has to be performed in-line with the scope of the investigation.
- ❖ The students are expected to analyse the obtained results and
- ❖ Elaborately discuss the same by preparing necessary Figures/Graphs/Tables/Illustrations/images to get an inference.
- ❖ The important conclusions need to be drawn and scope for further research also to be highlighted.
- ❖ The outcome of project work shall be published in journals /
- ❖ conference of National or International importance.
- ❖ At the end, students should submit a report covering the various
- ❖ aspects of the Project work.
- ❖ The typical components of the project report are the Introduction, Need for present study, Scope of the Investigation, Literature review,
- ❖ Methodology / Experimental investigation/development of software packages, Results & discussion of experimental and analytical work, Conclusions, References etc.
- ❖ The deadline for submission of final Project Report / Thesis /
- ❖ Dissertation is within 30 calendar days from the last Instructional day of the semester.
- ❖ The project co-ordinator, in consultation with the head of the department and controller of examination, shall arrange for an external expert member to conduct the final viva-voce examination to ascertain the overall

performance of the students in Project work.

**COURSE OUTCOMES:**

Students will be able to

**CO1:** Apply their practical knowledge and skill in Microbiology with specialization in to solve real time problems

**CO2:** Prepare an appropriate documentation

**PROFESSIONAL ELECTIVE COURSES****SEMESTER I**

<b>LSEY101</b>	<b>BIOSTATISTICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** make informed decisions based on data.

**COB2:** correctly apply a variety of statistical procedures and tests

**COB3:** know the uses, capabilities and limitations of various statistical procedures.

**COB4:** interpret the results of statistical procedures and tests

**COB5:** understand sampling techniques

**MODULE I INTRODUCTION TO STATISTICS 9**

Exploratory Data Analysis - Motivation, Population vs Sample," Scientific Method" - Definitions, Examples, Medical Study Designs – Graphical Displays: Dot plots, Stem plots, Pie chart, Histograms - Summary Statistics: Measures of Central tendency.

**MODULE II MEASURES OF DISPERSION 9**

Range, Quartile deviation, Mean deviation, Standard deviation, Variance, Coefficient of Dispersion: coefficient of variation, Moments: Relationship between raw and central moments, Effect of change of Origin and Scale, Pearson beta and gamma coefficients, Skewness: Measures of Skewness, Kurtosis.

**MODULE III CORRELATION AND REGRESSION**

Bi-variate data – Correlation and Regression coefficients and their relation, properties - Effect of change of origin and scale on correlation coefficient, Linear regression, Association and Independence of attributes.

**MODULE IV PROBABILITY AND ITS DISTRIBUTIONS 9**

Events - exhaustive, mutually exclusive and equally likely - Baye's theorem (without proof) - Binomial, Poisson, Exponential and Normal distributions - Simple properties of the above distributions (without derivation).

**MODULE V                      SAMPLING TECHNIQUES                      9**

Concept of population and sample, Random sample, Methods of taking a simple random sample, Tests of Significance: Sampling distribution of mean and standard error, Large sample tests (test for an assumed mean and equality of two population means with known S.D.); small sample tests (t-test for an assumed mean and equality of means of two populations when sample observations are independent, Paired and unpaired t-test for correlation and regression coefficients, t-test for comparison of variances of two populations, Chi-square test for independence of attributes, Goodness of fit.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Norman T J Bailey, "Statistical Methods in Biology " (3rd Edition), Cambridge University Press 1995
2. Gerald van Belle, L.D.Fisher, P.J.Heagerty, and T.Lumney, "Introduction to Biostatistics" Second Edition, John Wiley & Sons, New Jersey 2004
3. Wong Limsoon, "Essence of biostatistics" , NUS Lecture Notes Series 2003.
4. Gupta.S.C and V.K.Kapoor, "Fundamentals of Mathematical Statistics", Sultan Chand & Sons, New Delhi 2002.
5. Gupta.S.C., "Fundamentals of Applied Statistics", Sultan Chand & Sons, NewDelhi 2014.
6. Ross,S.M., "Probabilty and Statistics for Engineers and Scientists" John Wiley & Sons, New Jersey 2007

**COURSE OUTCOMES:**

**CO1:** The students will understand the basics of statistics

**CO2:** The students will gain knowledge of measures of dispersion

**CO3:** The students will gain knowledge of correlation and regression

**CO4:** The students will gain knowledge of probability

**CO5:** The students will gain knowledge of sampling techniques

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

SDG 4: Ensure inclusive and equitable quality education and promote lifelong opportunities for all.

Statement: Learning of various mathematical tools like Matrices, Calculus and Numerical methods will lead to knowledge of applications in biological fields

<b>LSEY102</b>	<b>BIO SAFETY, BIOETHICS,</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>BIOENTREPRENEURSHIP AND</b>				
	<b>IPR)</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** provide an understanding of the ethical issues underlying biotechnology research and innovation in addition to protection of the acquired intellectual property

**COB2:** The student will gain an understanding research methodology, the ethical issues underlying biotechnology research and the importance of protection of intellectual property

**COB3:** get an understanding of patents

**COB4:** interpret marketing strategies

**COB5:** understand intellectual property and laws pertaining to it

**MODULE I ETHICS IN BIOLOGY 9**

The legal and socioeconomic impacts of biotechnology - Public education of the processes of biotechnology involved in generating new forms of life for informed decision making - Biosafety regulation and national and international guidelines - rDNA guidelines

**MODULE II BIOSAFETY 9**

Experimental protocol approvals - levels of containment - Environmental aspects of biotech applications - Use of genetically modified organisms and their resistance in environment - Special procedures for r-DNA based product production

**MODULE III MARKETING 9**

Assessment of market demand for potential product(s) of interest; Market conditions, segments; Prediction of market changes; Identifying needs of customers including gaps in the market, packaging the product; Market linkages, branding issues; Developing distribution channels; Pricing/Policies/Competition; Promotion/Advertising; Services Marketing

**MODULE IV INTELLECTUAL PROPERTY RIGHTS 9**

Intellectual property rights - TRIP International conventions patents and methods of application of patents - Legal implications - Biodiversity and farmers rights - Beneficial applications' and development of research focus to the need of the poor - Identification of directions for yield effect in agriculture,

aquaculture Bioremediation etc.

## **MODULE V PATENT SYSTEM 9**

Objectives of the patent system - basic principles and general requirements of patent law - biotechnological inventions and patent law - legal development - patentable subjects and protection in biotechnology - The patentability of microorganisms - IPR and WTO regime - consumer protection and IPR - IPR and plant genetic resources - GATT and TRIPS.

**L – 45; TOTAL HOURS –45**

### **TEXT BOOKS:**

1. Beier, F.K., Crespi, R.S. and Straus, J. Biotechnology and Patent protection-Oxford and IBH Publishing Co. New Delhi, 1985.
2. Sasson A, Biotechnologies and Development, UNESCO Publications, 1988.
3. Singh K, Intellectual Property rights on Biotechnology, BCIL, New Delhi, 1993.

### **COURSE OUTCOMES:**

**CO1:** understand the nature of hazards related to biotechnology and the importance of biosafety in research.

**CO2:** debate on ethical issues related to biotechnology research.

**CO3:** understand methods used in scientific research and to emphasize on the importance of statistical concepts.

**CO4:** realize the importance of intellectual property and its protection under the constitution.

**CO5:** The students will understand patent system

### **Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

### **Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.



<b>LSEY103</b>	<b>RECOMBINANT DNA</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>TECHNOLOGY</b>				
		3	0	0	3

**COURSE OBJECTIVES:**

**COB1:** To introduce the students to different vectors for genetic manipulation of cells

**COB2:** To give a working knowledge for techniques involved in DNA extraction, purification and manipulation

**COB3:** Make the students understand the principle of techniques used in the creation of recombinant DNA molecules and the selection of the cells harbouring them

**COB4:** To learn applications of Recombinant DNA Technology

**COB5:** To become theoretically familiar with rDNA techniques

**MODULE I CLONING & CLONING VECTOR 9**

Types of cloning vectors viz. Plasmids, cosmids, ssDNA Phages, Yeast cloning vectors, Animal viruses, Ti plasmids and Cauliflower Mosaic Virus. Structural and Functional Organization of Plasmids, Plasmid Replication, Stringent and Relaxed Plasmids, Incompatibility of Plasmid Maintenance.

**MODULE II MANIPULATION OF PURIFIED DNA 9**

Enzymes involved in DNA Manipulation- Nucleases, Ligases, Polymerases and DNA modifying enzymes, Restriction endonucleases-Types, Blunt and sticky ends, Ligation- Mode of action of DNA Ligase.

**MODULE III CONSTRUCTION OF RECOMBINANT DNA 9**

Preparation of competent cell-Transformation, transfection – Recombinant selection and screening- Genomic DNA library- cDNA synthesis strategies - Linkers - Adapters - Homopolymer tailing- Making genomic and cDNA libraries in plasmids and phages. PCR product cloning (TA cloning). Cloning strategies in yeast, E. coli and B. subtilis.

**MODULE IV HYBRIDIZATION TECHNIQUES & MUTAGENESIS 9**

DNA hybridization, colony hybridization and in-situ hybridization (Southern, Northern and Dot blots and immunological techniques Western blotting), Mutagenesis - Deletion mutagenesis, Oligonucleotide derived mutagenesis, Site directed mutagenesis - Its applications- Applications of rDNA technology

in Diagnostics.

## **MODULE V APPLICATIONS OF rDNA TECHNOLOGY 9**

Gene Cloning and DNA analysis in Agriculture, Forensic Science and Medicine- Production of Recombinant pharmaceuticals, identification of genes responsible for human disease, Genetic Finger printing, Gene Therapy, Plant Genetic engineering, Problems with Genetically modified plants.

**L – 45; TOTAL HOURS –45**

### **TEXT BOOKS:**

1. James D. Watson, Recombinant DNA, 2<sup>nd</sup> Edition, Scientific American; Second Edition edition, 1998.
2. T. A. Brown, Gene Cloning and DNA analysis: An Introduction, 7<sup>th</sup> edition, Willey-Blackwell, 2016.

### **COURSE OUTCOMES:**

**CO1:** Familiarize with the basic concepts and principles of utilization of different expression vectors for cloning in prokaryotic and eukaryotic organisms

**CO2:** Understand the different strategies of gene cloning and construction of genomic and cDNA libraries for applications of recombinant DNA technology

**CO3:** Familiarize the concepts of structural and functional genomics

**CO4:** Understand utilization and principle of mutagenesis studies and hybridization probes

**CO5:** will be skilled enough to use these techniques in different fields, such as forensic science, agriculture, medicine, industry, etc.

### **Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

### **Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation M - Medium Correlation H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of rDNA technology we can modify the live organisms that can help in maintains systems to promote good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY121</b>	<b>MICROBIAL DIVERSITY AND</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>EXTREMOPHILES</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To impart knowledge about the taxonomical classification of Microorganisms.

**COB2:** To understand the Bergey's manual of systematic bacteriology

**COB3:** To elucidate the physiological structure, characters and economical important of Algae

**COB4:** To study the salient features about fungi, protozoa and virus.

**COB5:** To understand the structure and functions of extremophiles

**MODULE I DIVERSITY OF LIFE 9**

Prokaryotic and Eukaryotic microorganisms. Classification of microorganisms. Binomial Nomenclature. Taxonomy: Principles- numerical- genetic- serotaxonomy and chemotaxonomy -Binomial Nomenclature. Kingdom concept: Hackel three kingdom -Whittaker's five kingdom. Bergy's systemic classification of bacteria.

**MODULE II BACTERIA AND FUNGI 9**

General characteristics of bacteria and Archea bacteria. Structure of bacterial cell. Economic importance of bacteria. General characters of Actinomycetes and their importance. General characteristics and classification of fungi. Fungal cell structure. Mode of reproduction – *Mucor*, *Aspergillus*, *Penicillium* and *Rhizopus*. Economic importance of fungi.

**MODULE III ALGAE AND CYANOBACTERIA 9**

General characteristics and classification of Algae. Morphology of *Chlamydomonas*, *Oedogonium* and *Gracillaria*. Cyanobacteria – salient features and its importance. Heterocyst – structure of *Oscillatoria* and *Anabena*..

**MODULE IV VIRUS AND PROTOZOA 9**

General characteristics and structure of animal virus (Adeno virus), plant virus (TMV), bacteriophage (T4), insect virus (npv). General characters of Protozoa- Structure of *Euglena* and *paramecium*. Medically important parasite..

**MODULE V EXTREMOPHILES****9**

Extremophiles – Types, Classification, Molecular Evolution of Extremophiles, Strategies for the Isolation and Cultivation of Halophilic Microorganisms. Biotechnological Applications of Cold-Adapted Bacteria. Extremophiles and their Application.

**L – 45; TOTAL HOURS –45****TEXT BOOKS:**

1. Prescott L M, J P Harley and D A Klein (2005). Microbiology. Sixth edition, International edition, McGraw Hill.
2. Dubey RC and Maheswari DK (2012). A text of Microbiology (Revised edition). S.Chand and Company Ltd., New Delhi.
3. Geeta Sumbali and Mehrotra RS (2009). Principles of Microbiology. First edition, Tata McGraw Hill P.Ltd., New Delhi.
4. Pelczar TR M J Chan ECS and Kreig N R (2006). Microbiology. Fifth edition, Tata McGraw-Hill INC. New York.
5. Robert F Boyd (1984). General microbiology. Times mirror/Mosby college publishers.
6. Jagadish Chander (1996). A textbook of Medical Mycology. Interprint, New Delhi.
7. Atlas and Atlas. Microbiology. Pearson publications. 4TH edition.

**COURSE OUTCOMES:**

**CO1:** Understand the knowledge about Prokaryotic and Eukaryotic microorganisms.

**CO2:** Acquire knowledge about classification of microorganisms (Bergey's manual),

**CO3:** Created an understanding about salient features and economic importance of Algae

**CO4:** Created an understanding about salient features and economic importance of fungi

**CO5:** Retain knowledge about extrempphiles

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on  
20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of this course we can modify the live organisms that can help in maintains systems to promote good health and well being.

### SDG15: Life on Earth

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY122</b>	<b>MICROBIAL PHYSIOLOGY AND</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>METABOLISM</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

After completing this course student should be able to:

**COB1:** Distinguish between the eukaryotic and prokaryotic cell structure and describe the bacterial cell components and its function role in the cell

**COB2:** Define basic concepts of microbial physiology and explain microbial growth, growth kinetics and factors affecting growth.

**COB3:** Evaluate the importance of central pathways of carbohydrate metabolism for microbial physiology.

**COB4:** Cell enzymes and its role in nutrition • Illustrate macromolecular synthesis and processing.

**COB5:** Link the microbial physiology to the genomics of cells.

**MODULE I INTRODUCTION TO METABOLISM 9**

Microbial enzymes: Structure and Classification- Mechanism of Enzyme actions: Lock and key model, induced fit theory- Factors affecting rates of enzyme-mediated reactions- The role of ATP in metabolism

**MODULE II GROWTH AND STRESS MANAGEMENT 9**

Definitions of growth and generation time, measurement of microbial growth and specific growth rate- Batch and Continuous culture- Phases and types of growth curve and its industrial application- Microbial growth in response to temperature, pH, solute and water activity, oxygen, pressure and radiation- Oxidative stress- Thermal stress- Starvation stress and stringent response- Aerobic to anaerobic transitions

**MODULE III NUTRIENTS AND PATHWAYS 9**

Classification of bacteria based on nutrients-Membranes of microorganisms, Ion channels- Passive and facilitated diffusion, Primary and secondary active transport, concept of uniport, symport and antiport- Group translocation and Iron uptake-Photosynthetic pigments and apparatus in bacteria-Photophosphorylation-C<sub>3</sub> and C<sub>4</sub> pathways- Difference between oxygenic and anoxygenic photosynthesis -Mode of nutrition in Hydrogen and Nitrifying bacteria- Mode of nutrition in Purple sulfur bacteria, Non-sulfur bacteria and Green sulfur bacteria-Mode of nutrition in methylophs and methanogens- Utilization of light energy by

halobacteria

**MODULE IV                      AEROBIC/ANAEROBIC                      METABOLISMS                      9**  
**CONCEPTS**

Concept of aerobic respiration, anaerobic respiration and fermentation- Sugar degradation pathways i.e., EMP, ED and Pentose phosphate pathway- TCA cycle and Electron transport chain- Comparison of mitochondrial and bacterial ETC, electron transport phosphorylation, uncouplers and inhibitors

**MODULE V                      FERMENTATION                      AND                      INDUSTRIAL                      9**  
**APPLICATIONS**

Fates of pyruvate, Pasteur effect and industrial importance of fermentation- Alcohol fermentation- Lactate fermentation (homo fermentative and hetero fermentative pathways)- Concept of linear and branched fermentation pathways- Utilization of Lactose and Galactose- Utilization of Maltose and Mannitol- Degradation of cellulose, starch and glycogen- Conversion of biomass to energy using microorganisms - Mechanism of nitrogen fixation- Symbiotic and non-symbiotic nitrogen fixation- Biosynthesis of amino acids- Degradation amino acids

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Moat A.G., Foster J.W. and Spector M.P. 2002. Microbial Physiology, 4th edition. A Johan Wiley and sons inc., publication.
2. Kim B.H. and Gadd G.M. 2008. Bacterial physiology and metabolism. Cambridge University Press, Cambridge.
3. Gilbert H.F. 2000. Basic concepts in biochemistry: A student's survival guide. Second Edition. Mc-Graw-Hill Companies, health professions Division, New York.
4. Madigan M.T., Martinko J.M., Stahl D.A. and Calrk D.P. 2012. Brock Biology of Microorganisms. 13th ed. Pearson Education Inc.

**COURSE OUTCOMES:**

Upon successful completion of this course The student will be able to

**CO1:** Define basic concepts of microbial physiology and list the factors effect on the physiological microorganisms

**CO2:** Evaluate the importance of central pathways of carbohydrate metabolism for microbial physiology

**CO3:** Define types of membrane transport for nutrient uptake and protein



excretion and diagram biosynthesis pathway for amino acids, fatty acids, organic acids synthesis.

**CO4:** Link the microbial physiology to the genomics of cells and explain how some microorganisms can degrade the macromolecule polymers.

**CO5:** Calculate the kinetic energy in aerobic and anaerobic microbes and explain how the extremophiles live in their conditions

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of this course we can use knowledge to the life of other organisms that can help in maintains systems to promote good health, wellbeing and ecosystem.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and nonliving and relation with all the levels of life in the earth.

<b>LSEY123</b>	<b>PARASITOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To understand animal association, parasitic diseases, lifecycle and mode infection.

**COB2:** To get an overview on important parasites and characteristics.

**COB3:** To understand survival adaptations and significance of important parasites

**COB4:** To understand immunology related to parasites

**COB5:** To understand laboratory handling of parasites

**MODULE I      FUNDAMENTALS OF PARASITOLOGY      9**

Introduction to parasitology; type of parasites, animal associations and host – parasite relationship; distribution of diseases and Zoonoses caused by animal parasites.

**MODULE II      COMMON PARASITES      9**

Plasmodium - morphology, life-cycle and mode of infection. Molecular biology of Plasmodium – drug targets, mechanism of drug resistance, vaccine strategies and proteomic approaches; Leishmania - morphology, life-cycle and mode of infection. Molecular biology of Leishmania – drug targets, drug resistance and vaccine strategies. Entamoeba - morphology, biology, life-cycle and mode of infection. Giardia - morphology, biology, life-cycles, mode of infection. Gastro-intestinal nematodes - morphology, biology, life-cycles and modes of entry of Schistosoma, Wuchereria, Brugia, Ancylostoma, Trichinella and Dracanculus; molecular biology of nematodes and vaccine strategies.

**MODULE III      LABORATORY TECHNIQUES IN PARASITOLOGY**

Examination of faeces, ova and cysts – worm burden – concentration method – Flotation / Sedimentation technique – Staining Iron haetoxylin, Blood smear examinations – thick / thin smears – cultivation of protozoan parasites

**MODULE IV      PARASITE IMMUNOLOGY      9**

Immune response and self-defence mechanisms, immune evasion- Parasite causing infections in immunocompromised hosts and in AIDS- Biochemical adaptations of parasites; parasites of veterinary importance.

**MODULE V INSECT PARASITES****9**

Parasites of insects and their significance; nematode parasites of plants, morphology, biology, lifecycle and infection of crop plants by plant parasitic nematodes – Arthropod vectors Ticks, mites, flea, mosquitoes transmitting diseases in man and animals – vector control measures.

**L – 45; TOTAL HOURS –45****TEXT BOOKS:**

1. Roberts L.S. and Janovy J. Foundations of Parasitology. McGraw-Hill Publishers, New York, 2008.
2. Cox F. E. G. Modern Parasitology: A Textbook of Parasitology, FEG Cox., WileyBlackwell, 1993
3. Parija S. C. (1996). Text Book of Medical Parasitology. Orient Longmans.
4. Chatterjee (1986). Medical Parasitology. Tata McGraw Hill.

**COURSE OUTCOMES:**

**CO1:** Understand the basic concepts of parasitology as a field of science

**CO2:** Get knowledge of some common parasites, their structure, life cycle and associated diseases

**CO3:** Understand the basic immune response mounted by organisms as defence against parasites

**CO4:** Get knowledge about insect parasites, lifecycle, survival adaptations and also to prevention and treatment strategies.

**CO5:** Get knowledge about parasites and laboratory handling.

**Board of Studies (BoS) :**9<sup>th</sup>BoS of SLS held on 20.08.2022**Academic Council:**19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
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CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation M - Medium Correlation H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of this course we can modify the live organisms that can help in maintains systems to promote good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

**SEMESTER II**

<b>LSEY201</b>	<b>MOLECULAR DIAGNOSTICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** Developing the basic concept of molecular diagnostics

**COB2:** Understanding the common procedures and which are used in disease diagnosis

**COB3:** To be familiar with various types of diseases diagnosis methods and progression of diagnosed disease.

**COB4:** Understand the concepts of different types of vaccines

**COB5:** Apply the new technologies in healthcare settings

**MODULE I INTRODUCTION TO MOLECULAR DIAGNOSTICS 9**

Collection, preservation and storage of clinical samples, biopsy, Principles, application and limitations of biological assays used in diagnosis- PCR, ELISA, FISH, gene sequencing, microarrays, protein arrays. GLP, SOP and ethics in molecular diagnostics.

**MODULE II INFECTIONS 9**

Infection and mode of transmission, types of infectious diseases- bacterial and fungal infections, diagnosis of infections caused by Streptococcus, Coliforms, Salmonella, Shigella, Vibrio, and Mycobacterium- diagnosis of fungal infections, major fungal diseases, Dermatophytoses, Candidiosis and Aspergillosis. Diagnosis of DNA and RNA viruses- pox virus, rhabdo virus, hepatitis; virus diagnosis of protozoan diseases- amoebiosis, malaria, trypanosomiosis, leishmaniasis- study of helminthic diseases- Fasciola hepatica and Ascaris lumbricoides. Filariasis and Schistosomiasis. Diagnosis of chicken guinea and swine flu.

**MODULE III CLINICAL GENETICS 9**

Chromosomes chemistry and packaging, Cytogenetic, Structural and numerical abnormalities of chromosomes, Chromosome bands, banding techniques , mutation and polymorphism analysis, human genome project, cancer genetics- oncogenes, tumor suppressor genes- gene therapy, genetic counseling, nucleic acid hybridization techniques, Disease linked with mitochondrial DNA Genetic linkage and chromosome and genetic mapping in human diseases, Prenatal

**MODULE IV IMMUNODIAGNOSTICS 9**

Introduction to immunodiagnosics, antigen-antibody reactions, antibody production, antibody markers, CD markers, FACS, Human Leukocyte Antigen (HLA) typing, agglutination (ABO/Bacterial), immunoprecipitation, immunodiffusion, floctometer.

**MODULE V FORENSIC SCIENCE 9**

Introduction to Forensic Science, DNA fingerprinting / DNA Profiling / DNA Testing in Forensic Science.; Ethics, Rules and Procedures in DNA analysis. Autopsy and toxicological diagnosis. Determination of Paternity- Human identification and sex determination. semen analysis , Case study.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Carl A. Burtis, Edward R. Ashwood, Tietz Textbook of Clinical Chemistry, eds. Philadelphia, PA: WB Saunders, 1998
2. Lisa Anne Shimeld , Anne T. Rodgers, Essentials of Diagnostic Microbiology, Delmar Cengage Learning; New edition edition, 1998
3. John Crocker, David Burnett, The Science of Laboratory Diagnosis, Wiley, 2005

**COURSE OUTCOMES:**

**CO1:** Familiar with the theoretical working principles of clinical biochemistry.

**CO2:** Understand the causes and spread of infection and design strategy to stop their spread.

**CO3:** Understand the aspects of genetic disease, their causes and design strategy to diagnose them at earlier stages.

**CO4:** Learners will be able to define basic terminology and describes basic concepts in molecular diagnostics

**CO5:** will know the importance and the relevance of molecular diagnostic techniques and applications of molecular diagnostics in various field including medical, forensic, etc.

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

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CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

### SDG15: Life on Earth

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY202</b>	<b>PLANT AND ANIMAL</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>	<b>BIOTECHNOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To learn about embryogenesis and other type of hybridization techniques.

**COB2:** To know about genetic transformation and techniques about gene delivery.

**COB3:** To have an idea about gene mapping and cloning and different type of biotic and abiotic stress.

**COB4:** To familiar with different animal tissue culture techniques and methods of separation

**COB5:** To understand about artificial breeding in animals, transgenic and their applications

**MODULE I PLANT TISSUE CULTURE 9**

Totipotency, organogenesis, somatic embryogenesis, artificial seed production, Micropropagation, somaclonal variation, Germplasm conservation and cryopreservation. Protoplast Culture and Somatic Hybridization Protoplast isolation- its culture and usage, Somatic hybridization and its applications.

**MODULE II AGROBIOLOGY 9**

Agrobacterium-plant interaction; Virulence; Ti and Ri plasmids; Opines and their significance; T-DNA transfer, Genetic Transformation Agrobacterium-mediated gene delivery, Direct gene transfer - PEG- mediated, electroporation, particle bombardment and alternative methods; Screenable and selectable markers, Characterization of transgenics, Gene targeting.

**MODULE III MOLECULAR MAPPING & MARKER ASSISTED SELECTION (MAS) 9**

Resistance, grain quality and grain yield, Molecular polymorphism, RFLP, RAPD, STS, AFLP, SNP markers; Construction of genetic and physical map, Gene mapping and cloning, strategies for Introducing Biotic and Abiotic Stress Resistance/Tolerance Bacterial resistance; Viral resistance; Fungal resistance; Insects and pathogens resistance; Herbicide resistance; Drought, salinity, thermal stress, flooding and submergence tolerance.



**MODULE IV ANIMAL TISSUE CULTURE 9**

Tissue culture- definition, concept and significance, maintenance of sterility and use of antibiotics, detection of various biological contaminations, cross contamination, formulation of tissue culture media- serum and synthetic media, Balance salt Solution, Primary culture and Types, Cryopreservation of cell lines. role of growth factors in cell culture, various methods of cell separation, Cell cloning, transformation, transfection, micro-manipulation, nuclear transplantation

**MODULE V APPLIED ANIMAL BIOTECHNOLOGY 9**

Biotechnological approaches to obtain blood products: Tissue plasminogen activator and erythropoietin, production of vaccines and proteins of pharmaceutical relevance, recombinant protein production, harvesting and purification. Artificial breeding – in vitro fertilization and embryo transfer technology, artificial insemination, germ cell storage, transgenic animals- fish, mice and sheep, gene targeting and transfer, mouse models for human genetic disorder and diseases, knock-out and knock-in mice.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Edited by BR Jordan, 2nd Edition, The Molecular Biology and Biotechnology of Flowering, CABI, 2006.
2. Denis Murphy, Plant Breeding and Biotechnology: Societal Context and the Future of Agriculture, Cambridge University Press, 2007.
3. R. Ian Freshney. Culture of Animal cells, 5th Edition, 2010. A John Wiley & Sons, Inc., Publications, USA
4. Molecular Biotechnology: 4 edition. (2010), Glick B.R., Pasternak J.J., Patten C. L., ASM press, USA

**COURSE OUTCOMES:**

**CO1:** Understand the principle and concepts related to totipotency, embryogenesis, protoplast culture, applications of somatic hybridization, etc

**CO2:** Get knowledge about agrobacterium mediated creation of transgenic plants.

**CO3:** Understand the effect of biotic and abiotic stress components on different life forms and know the techniques to create plants that can circumvent such conditions

**CO4:** The concept of animal cell culture, growth, media, maintenance, culture conditions

**CO5:** Understand different classes of vaccines and their production, protein productions, and advanced animal breeding and animal transgenic technology.

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY203</b>	<b>PROTEIN ENGINEERING</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To learn about embryogenesis and other type of hybridization techniques.

**COB2:** To know about genetic transformation and techniques about gene delivery.

**COB3:** To have an idea about gene mapping and cloning and different type of biotic and abiotic stress.

**COB4:** To familiar with different animal tissue culture techniques and methods of separation

**COB5:** To understand about artificial breeding in animals, transgenic and their applications

<b>MODULE I</b>	<b>PROTEIN STRUCTURE AND ENGINEERING</b>	<b>9</b>
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Introduction, Overview of protein structure, Higher level structure, protein classification on the basis of structure, Protein structural stability, higher order structure prediction for secondary and tertiary structure, Chau and Fasman rule, protein folding, intrinsically disordered protein.

<b>MODULE II</b>	<b>POST-TRANSLATIONAL MODIFICATION</b>	<b>9</b>
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Introduction: post translational modification, types of post translational derivatives, types of post translational reactions, chaperones involved post translational modification, biological functions of post translational modification: regulation, cross links, covalent cofactors, membrane anchors, other functions.

<b>MODULE III</b>	<b>PROTEIN SOURCES</b>	<b>9</b>
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Recombinant versus non-recombinant production, Heterologous protein production in E.coli, bacteria other than E.coli, yeast and fungi, proteins from plants, animal tissue as protein source, Heterologous protein production in transgenic animals, Heterologous protein production in using cell culture.

<b>MODULE IV</b>	<b>PROTEIN PURIFICATION AND CHARACTERIZATION</b>	<b>9</b>
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Protein detection and quantification, Initial recovery of protein, removal of whole cells and cell debris, Concentration, chromatographic purification, Protein inactivation and stabilization, protein characterization.

## MODULE V STABILIZATION AND MODIFICATION OF PROTEINS 9

Insertion of Foreign DNA into Host Cells; Transformation; Transfection, Transduction, Construction of libraries, Oligonucleotide directed mutagenesis, error prone PCR, random mutagenesis with degenerate, oligonucleotide primers, random insertion/deletion mutagenesis, mutant proteins with unusual amino acids, different methods of engineering the protein.

**L – 45; TOTAL HOURS –45**

### TEXT BOOKS:

1. Proteins: Biochemistry and Biotechnology by Gary Walsh. (2002): John Wiley & Sons Ltd.
2. Proteins Analysis and Design. Ruth Hogue Angeletti, Albert Einstein College of Medicine of Yeshiva University Bronx, New York.
3. Protein Engineering in Industrial Biotechnology, Lilia Alberghina, Harwood academic publishers.

### COURSE OUTCOMES:

**CO1:** This Course will provide theoretical and methodological knowledge in Protein Engineering.

**CO2:** The student will get acquainted with enzyme kinetics, rational protein engineering

**CO3:** directed evolution of enzymes

**CO4:** In addition, student will get expertise in therapeutic protein and industrial enzymes widely used.

**CO5:** understand mechanisms behind protein stability

### Board of Studies (BoS) :

9<sup>th</sup>BoS of SLS held on 20.08.2022

### Academic Council:

19<sup>th</sup> AC held on 29.09.2022

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CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

SDG15: Life on Earth

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY211</b>	<b>MOLECULAR MICROBIOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To familiarize students with the bacterial and viral genome

**COB2:** To provide the students an in-depth knowledge of the process involved in DNA replication

**COB3:** To understand the concept of transcription in bacteria.

**COB4:** To familiarize the students with the concept and mechanism of transcription.

**COB5:** To study concept of transposition and process of gene regulations.

**MODULE I                    MICROBIAL GENOME ORGANIZATION                    9**

Comparative account of organization of Viral, Prokaryotic, eukaryotic and organellar genomes; Chromatin arrangement & nucleosome formation; C value paradox & genome size, cot curves; Operons, pseudogene, gene families, gene cluster, super families.

**MODULE II                    DNA REPLICATION                    9**

DNA replication: Prokaryotic and Eukaryotic DNA replication; mechanism, Initiation, elongation, termination and regulation of replication; DNA replication machinery, polymerases; Uni and bi-directional replication, D loop and rolling Circle model of replication; DNA damage and repair mechanisms: Direct repair, base excision repair, nucleotide excision repair, mismatch repair, recombination repair, SOS repair.

**MODULE III                    TRANSCRIPTION                    9**

The Mechanism of Transcription in Bacteria: RNA Polymerase Structure, Sigma (s) as a Specificity Factor, Promoters, Binding of RNA Polymerase to Promoters, Promoter Structure, Transcription Initiation, Elongation, Structure of the Elongation Complex, Termination of Transcription, Rho-Independent and dependent Termination; Operons: Fine Control of Bacterial Transcription (The *lac*, *era* and *trp* Operons).

**MODULE IV                    TRANSLATION                    9**

Initiation of Translation in Bacteria: tRNA Charging, Dissociation of Ribosomes, Formation of the 30S and 70S Initiation Complex, Bacterial Translational Control; The Direction of Polypeptide Synthesis and of mRNA Translation; The Genetic Code; The Elongation Cycle; Termination; Post-translation.

**MODULE V TRANSPOSITION AND GENE REGULATIONS 9**

Bacterial Transposons: Discovery of Bacterial Transposons, Insertion Sequences, Mechanisms of Transposition; Rearrangement of Immunoglobulin Genes, Recombination Signals, The Recombinase, Mechanism of V(D)J Recombination: Retrotransposons, Retroviruses, Retrotransposons; General aspects of gene regulations, The lactose system and the operon model.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Allison, L. A. (2021). *Fundamental molecular biology*. John Wiley & Sons.
2. Lodish, H., Berk, A., Kaiser, C. A., Kaiser, C., Krieger, M., Scott, M. P., ... & Matsudaira, P. (2008). *Molecular cell biology*. Macmillan.
3. Robert, F. W. (2012). *Molecular biology*, 5<sup>th</sup> Ed. McGraw-Hill.
4. Tang, Y. W., & Sails, A. (Eds.). (2014). *Molecular medical microbiology*. Academic press.

**COURSE OUTCOMES:**

**CO1:** Elucidate bacteria and viruses' genome organization and DNA replication mechanism.

**CO2:** Understand the mechanisms of transcription, translation and transposition.

**COB3:** To get familiar with transcription and translation in details.

**COB4:** To understand the signaling pathways in cell functioning

**COB5:** To understand energy conservation and conversion phenomenon in cells

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

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19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
CO1	H	L	-	-	-	-			-	-	-	
CO2	H	L	-	-	-	-			-	-	-	
CO3	H	L	-	-	-	-			-	-	-	
CO4	H	L	-	-	-	-			-	-	-	
CO5	H	L	-	-	-	-			-	-	-	

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.



<b>LSEY212</b>	<b>FERMENTATION TECHNOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To understand and apply scale-up methods for fermenters.

**COB2:** To understand the basic role of engineering in fermentation applications

**COB3:** To obtain a basic understanding of how cells work and become familiar with the environmental conditions (*i.e.* nutrients, pH, etc.) required for applications of biological components (cells or enzymes) to bio-processing systems

**COB4:** To utilize material balances to evaluate cell growth and substrate/product utilization in bioreactors.

**COB5:** To become familiar with principles of recovery and purification techniques of bioprocesses.

**MODULE I INTRODUCTION TO FERMENTATION TECHNOLOGY 9**

Aerobic and anaerobic fermentation, Production Strains of microorganisms, Production media, Types of Media, Carbon, Nitrogen Sources, Process flow sheeting – block diagrams, pictorial representation.

**MODULE II FERMENTERS 9**

Types & operation of Bioreactors, physico-chemical standards used in bioreactors, limitations of bioreactors, stages of fermentation processes, Media design for fermentation processes, Solid substrate fermentation, Fermenters (Stirred tank, bubble columns, airlift. Bioreactors, Static, Submerged and agitated fermentation), advantages & disadvantages of solid substrate & liquid fermentations.

**MODULE III AERATION AND AGITATION IN FERMENTERS 9**

Sparger and types, development of novel spargers, self-cleaning spargers, Impeller and types, axial and radial flow, baffle installation in bioreactor

**MODULE IV DOWNSTREAM PROCESSING 9**

Cell disruption- physical, chemical and enzymatic methods. Bioseparation, Chromatography, extraction, drying techniques

**MODULE V INDUSTRIAL PRODUCTION OF BIOBASED PRODUCTS 9**

Ethyl alcohol, Acetic Acid (Vinegar), Citric acid, lactic acid,  $\alpha$ -amylase, protease penicillin, tetracycline and vitamin B12, with reference to easily available raw materials, Production of herbal drugs.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Bioprocess Engineering - Basic concepts by M. L. Schuler & F. Kargi, Entice Hall 1992.
2. Bioprocess Engineering Principles by Pauline M. Doran, Academic Press 1995
- 3.

**COURSE OUTCOMES:**

**CO1:** Understand and apply scale-up methods for designing bioreactors.

**CO2:** Understand the basic role of engineering in bio-processing applications

**CO3:** Understand and model enzyme kinetics and apply the models for analysis of immobilized enzymatic bioreactors.

**CO4:** Utilize material balances to evaluate cell growth and substrate/product utilization in bioreactors.

**CO5:** Become familiar with principles of recovery and purification techniques of bioprocesses.

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

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CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

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**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY213</b>	<b>HOST-MICROBE INTERACTIONS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the basic concepts of bacterial taxonomy

**COB2:** To understand the basics of adaptation mechanisms of microbes

**COB3:** To understand various host-microbe interaction phenomenon.

**COB4:** To understand the phenomenon of gene transfer in microbes.

**COB5:** To acquire knowledge of the beneficial role of microbes in industry and environment.

**MODULE I INTRODUCTION AND BACTERIAL TAXONOMY 9**

History; classification, nomenclature, and taxonomy; Morphology and Physiology of Bacteria-Microscopy, Staining Techniques, Shapes of Bacteria, Anatomy, Physiology of bacteria, bacterial nutrition, bacteriocins; Sterilization and Disinfection methods

**MODULE II ADAPTATION & IDENTIFICATION OF MICROBES 9**

Introduction to the organisms of the microbial world; Adaptation of microbes to ecological niches; Methods for identification of bacteria; Bacteriology- Staphylococcus, Streptococcus, Pneumococcus, Neisseria, Mycobacterium, Haemophilus, etc

**MODULE III INTERACTION OF MICROBES 9**

Broad range of host-microbe interaction- types of host-microbe interactions, ecology and symbiosis; Normal flora of human body, microbes, health and disease

**MODULE IV MICROBIAL GENE TRANSFER 9**

Conjugation- mechanism, consequences, significance; Transformation- mechanism, consequences, bacterial competence; Transduction- mechanisms, Generalized transduction and Specialized transduction; Introduction to Viruses.

**MODULE V ROLE OF MICROBES IN INDUSTRY AND ENVIRONMENT 9**

Microbiology of Food- food spoilage, methods to control food spoilage, food borne diseases, detection methods, fermented foods; Industrial Microbiology- antibiotics, organic acids, biosurfactants, etc; Microbes in environment-

sanitization, bioremediation, etc.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. The host – bacterial interaction theory and the risk continuum: caseyhein,; contemporary oral hygiene, 2004.
2. Text book of Periodontology: Carranza 10 edition.
3. Text book of microbiology: Anathanarayna, 7 Edition

**COURSE OUTCOMES:**

**CO1:** Students will gain knowledge of the taxonomy, classification, physiology, anatomy of microbes.

**CO2:** Students will be able to understand the mechanisms of ecological adaptations employed by microbes.

**CO3:** Students will be able to understand the interrelationship of microbes with their environment and host.

**CO4:** Students will know the phenomenon of bacterial gene transfer and its associated consequences

**CO5:** Students will be able to get knowledge to use these microbes in industry and environment for human benefit

**Board of Studies (BoS) :**

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CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

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**SEMESTER III**

<b>LSEY 111</b>	<b>NANOBIOTECHNOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To introduce the basic concepts of Nanoscience relevant to the field of engineering

**COB2:** To provide an exposure about the importance of various synthesis method

**COB3:** To enrich the knowledge of students in various characterisation techniques

**COB4:** To make them to understand nanomaterials synthesis methods

**COB5:** To enrich the knowledge of students in various applications of nanomaterials

**MODULE I INTRODUCTION & CLASSIFICATION OF NANOMATERIALS 9**

Definition - Origin of nanotechnology - Difference between bulk and nanomaterials Top-down and bottom-up processes - Size dependent properties (magnetic, electronic, transport and optical), Classification based on dimensional property - 0D, 1D, 2D and 3D nanostructures

**MODULE II TYPES OF NANOMATERIALS 9**

Metal oxides and metal nano particles - Ceramic nano particles - Semi conducting quantum dots - Core-shell quantum dots - Nanocomposites - Micellar nanoparticles.

**MODULE III PRODUCTION OF NANOPARTICLES 9**

Biosynthesis, Sol-gel, hydrothermal, solvothermal, Solution plasma, Plasma Arcing, Electro deposition, Pulsed laser deposition, Chemical vapour, deposition.

**MODULE IV BIOMATERIALS 9**

Biopolymers – Types and Classification, Structure and functions; Nanostructured polymers, Oligosaccharides, Peptides, Drug Delivery molecule, Targeted Drug delivery.

**MODULE V CHARACTERISATION TECHNIQUES 9**

Basic principles of scanning Electron Microscopy (SEM), Atomic force microscopy (AFM), Scanning tunneling microscopy (STM), Scanning probe microscopy (SPM) and Transmission electron microscopy (TEM), Particle size analyzer, Luminescence techniques.

**L – 45; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Hari Singh Nalwa, —Handbook of Nanostructured Materials and Nanotechnology, Academic Press, 2000.
2. Guozhong Cao, —Nanostructures and Nano materials-Synthesis, Properties and Applications, Imperial College Press (2011).
3. Zhong Lin Wang, —Handbook of Nanophase and Nanomaterials (Vol 1 and II), Springer, 2002.
4. Mick Wilson, KamaliKannangara, Geoff smith, —Nanotechnology: Basic Science and Emerging Technologies, Overseas press, 2005.

**REFERENCES:**

1. A. Nabok, —Organic and Inorganic Nanostructures, Artech House, 2005.
2. C.Dupas, P.Houdy, M.Lahmani, Nanoscience: —Nanotechnologies and Nanophysics, Springer-Verlag Berlin Heidelberg, 2007.
3. Mick Wilson, KamaliKannangara, Michells Simmons and Burkhard Raguse, —Nano Technology – Basic Science and Emerging Technologies, 1st Edition, Overseas Press, New Delhi, 2005.
4. M.S. Ramachandra Rao, Shubra Singh, —Nanoscience and Nanotechnology: Fundamentals to Frontiers, Wiley, 2013.

**COURSE OUTCOMES:**

**CO1:** Apply the knowledge of different types of nanomaterials for various engineering applications.

**CO2:** Acquire the knowledge of various methods of production of nanomaterials

**CO3:** Familiarize with various characterization techniques.

**CO4:** Familiarize with various synthesis of nanomaterials

**CO5:** Familiarize with various applications of nanomaterials

**Board of Studies (BoS) :**

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CO2	H	L	-	-	-	-	L	L	-	-	-	
CO3	H	L	-	-	-	-	L	L	-	-	-	
CO4	H	L	-	-	-	-	M	L	-	-	-	
CO5	H	L	-	-	-	-	H	L	-	-	-	

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

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<b>LSEY112</b>	<b>MEDICAL CODING</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To introduce about medical coding and the role of coder.

**COB2:** To study the Anatomy of upper body parts such as the heart, lung, and skin

**COB3:** To study the Anatomy of upper body parts such as heart, lung and skin.

**COB4:** To study on ICD-10 coding for major diseases.

**COB5:** To introduce HCPCS

**MODULE I OVERVIEW OF CODING-INTRODUCTION 9**

Introduction-Medical coding, Role of Medical coder, Healthcare in India and US, Patient, Provider and Payers Relationship, Importance and significance of coding in today's world, Coding as a Profession, First Aid/CPR and Medical Law and Ethics

**MODULE II OVERVIEW OF ANATOMY-I 9**

Human body – Anatomical Position, Body Cavities, Dermatology, Ophthalmology, Otorhinolaryngology, Pulmonology and Cardiology

**MODULE III OVERVIEW OF ANATOMY-II**

Gastroenterology, Genitourinary system, Gynaecology and Obstetrics, Orthopaedics, Endocrinology, Immunology, Lymphatics, and Hematology

**MODULE IV ICD-10-CM 9**

General Coding Guidelines, Infections and Parasitic Diseases, Neoplasms Diseases of Nervous system, Diseases of Sense organs, Diseases of Circulatory System, Diseases of Respiratory system, Diseases of Digestive system, Diseases of Genitourinary system, Complications of Pregnancy, and Childbirth and Puerperium

**MODULE V HCPCS (HEALTHCARE COMMON PROCEDURE CODING SYSTEM) 9**

Introduction to HCPCS, Significance and Usage, Types of HCPCS, Modifiers Level II HCPCS

**L – 45; TOTAL HOURS –45**



**TEXT BOOKS:**

1. Sandra L Johnson & Robin Linker. Understanding Medical Coding: A Comprehensive Guide. 4<sup>th</sup> Edition. Cengage Learning. 978-1305666122.
2. Buck's Step-by-Step Medical Coding, 2022 Edition 1st Edition. Elsevier. 978-0323790383
3. ICD-10-CM 2022 THE COMP OFF CO: The Complete Official Codebook With Guidelines (ICD-10-CM The Complete Official Codebook) Spiral-bound – Import, 30 September 2021. American Medical Association. 978-1640161559.

**COURSE OUTCOMES: After the completion of the course**

**CO1:** Students understand the basis of medical coding and the role of coder.

**CO2:** Students understand the basic anatomy of heart, lungs, and Ophthalmology

**CO3:** Students understand the basic anatomy of gastrointestinal system, Genitourinary system and Gynaecology

**CO4:** Students understand the ICD-10- coding for major diseases.

**CO5:** Understand about the HCPCS

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CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

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<b>LSEY113</b>	<b>GENE MANIPULATION</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:****COB1:**

To learn about genetic manipulations, principles involved in manipulating genes and DNA.

**COB2:** To know about cloning strategies and expression systems.

**COB3:** To acquire basic understanding of techniques in genetic engineering

**COB4:** To learn PCR and its variations

**COB5:** To understand the principles of gene alteration

**MODULE I                    FUNDAMENTALS OF GENE MANIPULATIONS                    9**

Introduction to gene manipulation, Recombinant DNA by gene manipulation, Structure and functional properties of DNA. Different types of enzymes used in gene manipulation- Restriction Enzymes, DNA ligase, Klenow enzyme, T4 DNA polymerase, Polynucleotide kinase, Alkaline phosphatase; Cohesive and blunt end ligation; Linkers; Adaptors; Homopolymeric tailing; Labeling of DNA: Nick translation, Random priming, Radioactive and non-radioactive probes, Hybridization techniques: Northern, Southern and Colony hybridization, Fluorescence in situ hybridization.

**MODULE II                    CLONING VECTORS                    9**

Plasmids; Bacteriophages; M13mp vectors; PUC19 and Bluescript vectors, Phagemids; Lambda vectors; Insertion and Replacement vectors; Cosmids; Artificial chromosome vectors (YACs; BACs); Expression vectors, Plant based vectors, Ti and Ri as vectors, Yeast vectors, Shuttle vectors.

**MODULE III                    TECHNIQUES OF GENE MANIPULATIONS**

Insertion of Foreign DNA into Host Cells; Transformation; Transfection, Transduction, Construction of libraries; Isolation of mRNA and total RNA; cDNA and genomic libraries; cDNA and genomic cloning; Yeast two hybrid system; Phage display; Principles in maximizing gene expression. Methods to confirm cloning and reporter genes and proteins.

**MODULE IV                    PCR AND ITS APPLICATIONS                    9**

Primer design; Fidelity of thermostable enzymes; DNA polymerases; Types

of PCR — multiplex, nested, reverse transcriptase, real time PCR, touchdown PCR, hot start PCR, colony PCR, cloning of PCR products; Site specific mutagenesis; PCR in molecular diagnostics; Viral and bacterial detection; Automated DNA sequencing; RNA sequencing.

#### **MODULE V APPLICATIONS OF GENE MANIPULATION 9**

Gene cloning and analysis in medicine: Production of recombinant pharmaceuticals, recombinant insulin. Synthesis of growth hormones in *E. coli*, recombinant factor VIII, Recombinant vaccines. Gene therapy and cancer. Gene manipulation in agriculture: plant genetic engineering, The endotoxins of *Bacillus thuringiensis*, Cloning endotoxin gene in maize, Countering insect resistance to endotoxin crops, Herbicide resistant crops, Gene subtraction-Antisense RNA and the engineering of fruit ripening in Tomato. Gene manipulation in Forensic Science- DNA analysis in the identification of crime suspects, Genetic fingerprinting by hybridization probing, DNA profiling by PCR of short tandem repeats

**L – 45; TOTAL HOURS –45**

#### **TEXT BOOKS:**

1. S.B. Primrose, R.M. Twyman and R.W. Old; Principles of Gene Manipulation, 6th Edition, S.B. University Press, 2001.
2. Brown TA: Gene Cloning and DNA Analysis: 6<sup>th</sup> Edition, Wiley-Blackwell, pages 338, 2010.
3. Brown TA, Genomes, 3<sup>rd</sup> ed. Garland Science 2006
4. Selected papers from scientific journals.
5. Desmond S.T. Nicholl An Introduction to Genetic Engineering Cambridge University Press 2008

#### **COURSE OUTCOMES:**

**CO1:** Familiarize with the basic concepts and principles of utilization of different expression vectors for cloning in prokaryotic and eukaryotic organisms

**CO2:** Understand the different strategies of gene cloning and construction of genomic and cDNA libraries for applications of recombinant DNA technology

**CO3:** Familiarize the concepts of structural and functional genomics

**CO4:** Understand utilization and principle of mutagenesis studies and hybridization probes

**CO5:** will be skilled enough to use these techniques in different fields, such as forensic science, agriculture, medicine, etc.

**Board of Studies (BoS) :**

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CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

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SDG 3. Good Health and Well Being

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SDG15: Life on Earth

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<b>LSEY131</b>	<b>ANTIBIOTICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the history and contribution of antibiotics

**COB2:** To understand the classes and mechanisms of antibiotics

**COB3:** To understand the antibiotic resistance and mechanisms

**COB4:** To acquire the knowledge of actions of antibiotics on cells

**COB5:** To understand the production and applications of antibiotics

**MODULE I HISTORY AND INTRODUCTION TO ANTIBIOTICS 9**

Introduction, Definition – Antibiotics, Types, Evolution and History - Alexander Fleming, Lilly, Merck, Sandoz, Chain and Florey.

**MODULE II ANTIBIOTIC CLASSES AND MECHANISMS OF RESISTANCE 9**

Antibiotic classes. Targets for bacterial pathways. Mechanism of antibiotic resistance genes and antibiotics resistant bacteria. Linkage between antibiotic/heavy metal resistance genes and virulence.

**MODULE III OVERVIEW OF ANTIBIOTIC RESISTANCE**

History of antibiotic use since 1945 - Mobile elements [plasmids, transposons, integrons] - Bacterial gene exchange [conjugation, transformation, transduction]. Basic overview of antibiotic resistance, differences between bacteria, viruses, fungi, parasites

**MODULE IV ROLE OF ANTIBIOTICS 9**

Antibiotics that act on cell wall biosynthesis - penicillin, ampicillin, cephalosporins, monobactams, carbapenems, vancomycin, daptomycin. Antibiotics that block bacterial protein biosynthesis - tetracyclines, macrolides, aminoglycosides, clindamycin, chloramphenicol, linezolid. Antibiotics that block DNA replication and repair - fluoroquinolones, trimethoprim/sulfamethoxazole.

**MODULE V ANTIBIOTICS PRODUCTION AND APPLICATION 9**

Antibiotics – Bioprospecting, Screening, Primary and secondary metabolites. Fermentation – Submerged fermentation – Solid state fermentation – Extraction and Purification. Penicillin, Streptomycin, Cephalosporin, Amoxicillin.

**L – 45; TOTAL HOURS –45****REFERENCES:**

1. Marinez JK, Baquero F. 2014. Emergence and spread of antibiotic resistance: setting parameter space. *Upsala J Med Sciences*. 119:68-77. <http://informahealthcare.com/doi/pdf/10.3109/03009734.2014.901444>
2. Heuer H., Smalla K. 2007. Horizontal gene transfer between bacteria. *Environ Biosafety Res*. 6:3-13. <http://dx.doi.org/10.1051/ebr:2007034>
3. C. Walsh, Antibiotics: Actions, Origins, Resistance (<http://www.asmscience.org/content/book/10.1128/9781555817886>)

**COURSE OUTCOMES:**

**CO1:** Apply the knowledge of evolution of antibiotics

**CO2:** Acquire the knowledge of classes of antibiotics and its mechanism.

**CO3:** Familiarize with various antibiotics resistance.

**CO4:** Familiarize with various role of actions of antibiotics

**CO5:** Familiarize with various applications and productions of antibiotics.

**Board of Studies (BoS) :**

9<sup>th</sup>BoS of SLS held on 20.08.2022

**Academic Council:**

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CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

**SDG 3. Good Health and Well Being**

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**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY132</b>	<b>MICROBIAL SYSTEMS BIOLOGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** Developing the basic concept of Mathematical modelling at cell and molecular level

**COB2:** Understanding the dynamic and robustness of metabolic networks among microbes

**COB3:** To know the molecular signalling mechanisms and acquisition of drug resistance, prevention strategies

**COB4:** Understanding bacterial chemotaxis

**COB5:** Understanding establishment of infection

**MODULE I MATHEMATICAL MODELS FOR SYSTEMS BIOLOGY 9**

Introduction to modelling methodologies in life and biomedical science. Modelling strategies in prokaryotic systems relevance to structure, function and dynamics of biological systems.

**MODULE II DYNAMICS AND ROBUSTNESS OF METABOLIC NETWORKS 9**

Microbial Metabolism, genome scale computational modelling approaches with available molecular genetic tools. Metabolic flux determination and applications.

**MODULE III BACTERIAL CHEMOTAXIS**

Chemotaxis signal-transduction pathway, sensing and cell signalling, regulation, gene expression and defence mechanisms, chemoreceptors in *Bacillus subtilis*, and *Rhodobactersphareoides*. Molecular mechanisms of proteins and their interaction, quorum sensing. Bacterial DNA repair, mutagenesis, motility, transport and intercellular competitions. Yeast cell cycle and metabolic control.

**MODULE IV SYSTEMS BIOLOGY OF INFECTION 9**

Drug resistance pathogens- MDR, SXT gene elements, Integrative and Conjugative Elements (ICEs), Class I integrons. Prevention and control strategies. Molecular mechanisms of protein machines, bacterial pathogenesis, cellular homeostasis control and cell population dynamics

**MODULE V HOST-MICROBE INTERACTIONS 9**

Cellular and molecular level changes between host cell and intracellular

pathogens, Mtb, Influenza, Malaria, HIV, SARS and Corona. Gut-microbiome interactions. Dynamic of phospholipids, actin polymerization and myosins in the particle-shape recognition by the amoeba *Dictyostelium*.

**L – 45; TOTAL HOURS –45**

**REFERENCES:**

1. Alon, Uri. *An Introduction to Systems Biology: Design Principles of Biological Circuits*. Chapman & Hall / CRC, 2006. ISBN: 9781584886426.
2. Nowak, M. A. *Evolutionary Dynamics: Exploring the Equations of Life*. Belknap Press, 2006. ISBN: 9780674023383.
3. Becskei, A., and L. Serrano. "Engineering Stability in Gene Networks by Autoregulation." *Nature* 405 (2000): 590–3.
4. Alberts, Bruce. *Essential Cell Biology*. Garland Science, 2009. ISBN: 9780815341291.
5. Strogatz, Steven H. *Nonlinear Dynamics and Chaos: With Applications to Physics, Biology, Chemistry, and Engineering*. Westview Press, 2014. ISBN: 9780813349107.

**COURSE OUTCOMES:**

**CO1:** Familiar with basic knowledge on the microbial systems biology

**CO2:** Understand the microbes and their environment

**CO3:** Understanding the global history of microbes and probing the microbial complexity in the environment.

**CO4:** Understanding systems biology of infection

**CO5:** Understanding beneficial infections

**Board of Studies (BoS) :**

**Academic Council:**

9<sup>th</sup>BoS of SLS held on 20.08.2022

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	PO 12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation



**SDG 3. Good Health and Well Being**

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

**SDG15: Life on Earth**

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

<b>LSEY133</b>	<b>METAGENOMICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 3, 15</b>		3	0	0	3

**COURSE OBJECTIVES:**

**COB1:** Provide focus on next generation DNA sequencing technology to describe the ecological roles of microbial communities in different environments.

**COB2:** To use metagenomic data to describe the taxonomic make-up, functional potential and ecological processes of microbial communities from a range of environments

**COB3:** To apply next generation sequencing technology.

**COB4:** To assemble and annotate genomes by identifying genes.

**COB5:** To identify proteases for treatment of diseases

**MODULE I ENVIRONMENTAL GENOMICS 9**

Environmental Metagenomics – Introduction; Pure culture and in consortium ; Cultivable and Non-cultivable microbial analysis; Recombination DNA technology and DNA cloning; Types of vectors, applications of recombination DNA technology; Molecular fingerprinting techniques (RFLP, T-RFLP, ARISA, DGGE, rDNA library, and FISH); Stable isotope probing (SIP); Suppressive subtractive hybridization (SSH); Differential expression analysis (DEA); Microarrays & Metagenome sequencing; Next-generation sequencing approaches to metagenomics

**MODULE II ISOTOPE PROBING & OLIGONUCLEOTIDE MICROARRAYS 9**

Direct linking of microbial populations to specific biodegradation and biotransformation processes by stable isotope probing of biomarkers- PhyloChip&GeoChip-Detection of xenobiotic-degrading bacteria by using oligonucleotide microarrays

**MODULE III CONSTRUCTION & ANALYSIS OF METAGENOMIC LIBRARIES 9**

Cataloging microbes: phylogenetic tree and construction - Construction of a metagenomic library; Analysis of Metagenomic Libraries; Sequence-based Metagenomics Analysis; Functionbased Metagenomics Analysis; Phylogenetic analysis and Comparative genomics Softwares& Tools

**MODULE IV METAGENOMICS CASE STUDIES 9**

Metagenomic analysis of soil microbial communities; Metagenomic analysis of marine microbial communities; Metagenome of the Microbial Community in Acid Mine Drainage ; Metagenomic Analysis of Bacteriophage; Metagenomics and Its Applications to the Study of the Human Microbiome; Archaeal Metagenomics: Bioprospecting Novel Genes and Exploring New Concepts.

**MODULE V METAGENOMICS IN ENVIRONMENTAL 9**

Application of Metagenomics to Bioremediation; Applications of Metagenomics for Industrial Bioproducts; Escherichia coli host engineering for efficient metagenomic enzyme discovery; Next-generation sequencing approaches to metagenomics; Stable isotope probing: uses in metagenomics; DNA sequencing of uncultured microbes from single cells.

**L – 45; TOTAL HOURS –45**

**REFERENCES:**

1. Diana Marco Universidad Nacional de Cordoba, Argentina, "Metagenomics: Theory, Methods and Applications", Caister Academic Press,2010.
2. Diana Marco Universidad Nacional de Cordoba,Argentina "Metagenomics: Current Innovations and Future Trends",Caister Academic Press,2011.
3. Joanna R. Freeland, Heather Kirk, Stephen Petersen, "Molecular Ecology", Mc Graw Hill, 2nd Edition "2012.
4. Beebee T.J.C., D G. Rowe," An Introduction to Molecular Ecology", Mc Graw Hill, 2004.

**COURSE OUTCOMES:**

**CO1:** Apply the knowledge of omics to biological system of interest to obtain a snapshot of the underlying biology at a great resolution

**CO2:** Able to design drugs at the level of transcriptome

**CO3:** Understand the interaction of drugs at proteome level.

**CO4:** Able to design strategies that can integrate genomics, proteomics, transcriptomics to understand the living systems

**CO5:** Recognize proteases as the next target for treatment of emerging diseases.

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12
CO1	H	L	-	-	-	-	L	L	-	-	-	L
CO2	H	L	-	-	-	-	L	L	-	-	-	L
CO3	H	L	-	-	-	-	L	L	-	-	-	L
CO4	H	L	-	-	-	-	M	L	-	-	-	L
CO5	H	L	-	-	-	-	H	L	-	-	-	L

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

### SDG 3. Good Health and Well Being

Statement: Understanding of the fundamentals of this course can help in maintains systems to promote good health and well being.

### SDG15: Life on Earth

Statement: This course gives knowledge about the living and non living and relation with all the levels of life in the earth.

**OPEN ELECTIVE COURSES**

<b>OEEY 731</b>	<b>ADVANCED MATERIALS FOR</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>	<b>ENERGY APPLICATIONS</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the fundamentals and basics of materials for solar energy.

**COB2:** To provide the knowledge of the synthesis of materials.

**COB3:** To understand about the characterization of materials.

**COB4:** To provide an understanding of energy harvesting materials.

**COB5:** To provide insights in to the energy storage materials.

**MODULE I MATERIALS FOR PHOTOVOLTAICS 9**

First generation solar cell materials; single and polycrystalline Silicon, amorphous silicon: growth and wafer processing, contact materials, materials for surface engineering. Second generation solar cell materials; CdSe, CdTe, Copper Indium Gallium Selenide (CIGS), Gallium Arsenide for applications in photovoltaics, Materials for thin film solar cells, Thin film processing, and properties. Contact materials for second generation solar cells. Third generation solar cell materials; Quantum Dots, Organic materials, Composites, Dyes, Perovskites and their synthesis, characterization and properties, Interface energetics, photoactive layers and their materials, role of electron transport, hole transport, electron blocking and hole blocking materials and their processing. Contact materials and processing of contact layers.

**MODULE II RECHARGEABLE BATTERIES 9**

Primary and secondary batteries, battery potential, charge figure of merit, energy and power in battery, polarization losses, thermodynamics of battery materials, tortuosity and porosity of battery materials, reversible and irreversible interfacial reactions, battery architecture and design guidelines, Lead–acid battery, Nickel–cadmium battery (NiCd), Nickel–metal hydride battery (NiMH), Lithium-ion battery, Lithium-ion polymer battery, Energy density, power density, price and market. Battery Management systems and System Performance

**MODULE III SUPERCAPACITORS AND HYDROGEN 9**

## STORAGE

Basic components of super capacitors like types of electrodes like high surface area activated carbons, metal oxide and conducting polymers, aqueous and organic electrolytes Background and working of Fuel Cell, Hydrogen production processes, Hydrogen storage: Physical and chemical properties, general storage methods, compressed storage-composite cylinders, glass micro sphere storage, zeolites, metal hydride storage, chemical hydride storage and cryogenic storage, Carbon based materials for hydrogen storage.

### MODULE IV MATERIALS FOR ENERGY HARVESTING 9

Piezoelectric, Pyroelectric and Thermo-electrics materials, Electrostatic (capacitive) Energy Harvesting and materials, energy from Magnetic Induction, Metamaterial, energy from atmospheric pressure changes, electroactive polymers (EAPs), nanogenerators, Ambient radiation sources and nanoantenna, energy from noise.

### MODULE V MATERIALS FOR ENERGY STORAGE 9

Electrochemistry and electro-chemical Battery materials, Hydrogen Storage materials for fuel cells: Metal hybrids, Nanostructured metal hydrides, Non-metal hydrides, Carbohydrates, Synthesis of hydrocarbons, Aluminum, Liquid organic hydrogen carriers (LOHC), Ammonia, Amine borane complexes, Nano borohydrides and nano catalyst doping, imidazolium ionic liquids, phosphonium borate, Carbonite substances, Metal Organic frameworks, Activated Carbons, Carbon nanotubes, Clathrate hydrates, Glass capillary arrays.

**L – 45; TOTAL HOURS –45**

#### REFERENCES:

1. Detlef Stolten, "Hydrogen and Fuel Cells: Fundamentals, Technologies and Applications", Wiley, 2010.
2. Jiujun Zhang, Lei Zhang, Hansan Liu, Andy Sun, Ru-Shi Liu,
3. "Electrochemical Technologies for Energy Storage and conversion", John Wiley and Sons, 2012.
4. Francois Beguin and Elzbieta Frackowiak, "Super capacitors", Wiley, 2013.

5. Doughty Liaw, Narayan and Srinivasan, "Batteries for Renewable Energy Storage", The Electrochemical Society, New Jersey, 2010.

### COURSE OUTCOMES:

At the end of the course, the students will be able to

**CO1:** analyze the different energy storage systems.

**CO2:** understand the concepts and design of batteries

**CO3:** get the insights into supercapacitors and hydrogen fuels

**CO4:** comprehend the ideas behind the materials used for energy harvesting.

**CO5:** get familiarized with the different materials used for energy applications.

### Board of Studies (BoS) :

BOS of Physics was held on 30.6.22

### Academic Council:

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PSO1	PS O2	PS O3
CO1	H	M	L	L	M	M	M	L	L	L	M	M	M	M	M
CO2	H	M	M	L	L	M	L	L	L	L	L	M	M	M	M
CO3	H	M	M	L	L	L	L	L	L	L	L	M	M	M	M
CO4	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M
CO5	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M

**Note:** L- Low Correlation    M -Medium Correlation    H -High Correlation

SDG 4 : Ensuring inclusive and equitable quality education for all persons and promote lifelong learning opportunities.

Statement : The modules and topics mentioned in this course are designed to ensure all inclusive and thorough education with equity to all persons and promote learning opportunities at all times.

<b>OEEY 732</b>	<b>ALTERNATIVE ENERGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 7,9</b>	<b>RESOURCES</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

The students will be trained about the

**COB1:** Different types of batteries

**COB2:** Factors affecting battery performance

**COB3:** Selection and application of batteries

**COB4:** Application in photovoltaic cells

**COB5:** Various materials used in solar cells and PEC cells

**MODULE I      BATTERIES      9**

Lithium-ion battery, The Principle carbonaceous anode materials, cathode material- The intercalative reactions, relationships between performance requirements and materials characteristics- Electrolyte, separator. Advanced Ni-MH Batteries: Improvement in hydrogen storage alloys, improvement in Cathode materials, improvement in separator and cell design.

**MODULE II      FACTORS      AFFECTING      BATTERY      9**  
**PERFORMANCE      AND      SELECTION      OF**  
**BATTERIES**

Factors affecting battery capacity, voltage level current drain of discharge, types of discharge continuous, intermittent, constant current, constant load, constant power, temperature of battery during discharge, service life, voltage regulation, changing voltage, effect of all design, battery age and storage condition, effect of battery design.

Major consideration in selecting a battery, battery applications, comparative features and performance characteristics, characteristics of batteries for portable equipment.

**MODULE III      PHOTOVOLTAICS      9**

Basic of photovoltaics, homo and heterojunctions, preparation of single crystals and polycrystalline silicon solar cells, Metal-Insulator-Metal and semiconductors - Insulator-semiconductors solar cells, photovoltaic measurements - I-V characteristics, spectral response and capacitance measurements.

**MODULE IV      FUEL CELLS AND SUPER CAPACITORS      9**

Introduction, Types of Fuel cells, figure of merit, electro catalysts for



hydrogen oxidation and oxygen reduction, electrochemical double layer capacitors, ruthenium oxide as capacitor electrode, manual capacitors with proton conducting solid polymer electrolytes.

Ultra capacitors: Double layer, Metal Oxide, conducting polymers energy and power densities, voltage limitation and self discharge.

## **MODULE V SOLAR CELLS AND PEC CELLS 9**

Preparation of CdS/Cu<sub>2</sub>S solar cells by screen printing technique and their characteristics, amorphous Si solar cells GaAs solar cells, Semiconductors electrolyte interface. Photoelectrochemical (PEC) cells for conversion of light energy to electrical energy, PEC cells based on CdSe Si and GaAs and their output characteristics, Estimation of flat band potential from Mott-Schottky plots.

**L – 45; TOTAL HOURS – 45**

### **REFERENCES:**

1. Energy Storage Systems for Electronics Edited by Tetsuya Osaka, Department of Applied Chemistry, Wasuda University, Tokyo, Japan and Madhav Dutta, Intel Corporation, Hillsboro, USA, 2000.
2. Photoelectrochemical Solar Cell, Edited By K.S.V. Santhanam and M. Sharon, Elsevier Science Publishers, BV New York, 1995.
3. A.F. Fahrenbruch and R.H. Bube, Fundamentals of Solar Cells, Academic Press, London 1983.
4. W.E. Hatfield and J.H. Miller (Editors), High Temperature Superconducting Materials, Marcel Dekker, New York 1988.
5. Lindar D., Handbook on Batteries and Fuel Cells, McGraw Book Co., New York, 2011.

### **COURSE OUTCOMES:**

The students will have

**CO1:** A thorough understanding about batteries and their components

**CO2:** Understand the working mechanism of batteries.

**CO3:** Influence of various factors on performance of batteries and based on which selection of suitable batteries depending on application.

**CO4:** Testing in fuel cells.

**CO5:** Applications in solar cells and PEC cells.

### **Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

### **Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1		M			H										
CO2	H			H	M										
CO3		H				L									
CO4	M			H		L									
CO5	H				M										

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 7: Affordable & Clean Energy

SDG 9: Industry, Innovation and Infrastructure

Statement:

SDG 7: Ensure access to affordable, reliable, sustainable and modern energy for all

SDG 9: Build resilient infrastructure, promote inclusive and sustainable industrialization and foster innovation

<b>OEEY 701</b>	<b>ANALYTICAL TECHNIQUES</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 6, 7</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the students to understand the

**COB1:** basics in data analysis

**COB2:** basics and principles in volumetric and gravimetric analysis

**COB3:** types and principles of electro analytical methods

**COB4:** principles and analysis of spectroscopic techniques

**COB5:** the principle and methods in chromatography and thermal analysis

**MODULE I DATA ANALYSIS 9**

Precision and accuracy, Classification of errors, methods of minimization and elimination of errors Mean and standard deviation; absolute and relative errors; students t-test, F-test, linear regression for deriving calibration plots, covariance and correlation coefficient

Statistics for analytical experimentation: Probability, Regression analysis, Data analysis and signal enhancement.

**MODULE II VOLUMETRIC METHODS OF ANALYSIS 9**

Different methods of expressing concentration terms, Difference between titrimetric and volumetric analysis, Types and roles of indicators - Principle and reactions involved in neutralization, precipitation, complexometric and redox titrations, calculations involving stoichiometry – for all types of systems - Gravimetric analysis (volatilisation and precipitation methods)

**MODULE III ELECTROANALYTICAL METHODS 9**

Types of electrodes - Conductometric Titrations - Potentiometric titrations - pH-metry and ion-selective electrodes - Amperometric titrations - Coulometric Titrations, DM Electrode - polarography - electrogravimetry - voltammetry, cyclic voltammetry, impedance studies - Electrochemical sensors, ISFETs, CHEMFETs.

**MODULE IV SPECTROPHOTOMETRIC TECHNIQUES 9**

Quantitative applications of Colorimetric analysis – UV-Visible spectrophotometry – *Atomic absorption spectroscopy (AAS)* - atomic

emission spectroscopy (AES), *Flame photometry*, ICP-AES - Fluorescence spectroscopy, Stern Volmer Equation and quantum yield calculation.

#### **MODULE V CHROMATOGRAPHIC TECHNIQUES AND 9 THERMAL METHODS**

Chromatography: Paper, TLC and column Chromatography – Detectors in Chromatography - GC, HPLC, (hyphenated techniques GC/MS, LC/MS) and GPC -- ion exchange chromatography – Electrochromatography: Capillary electrophoresis and gel electrophoresis

Thermal analytical techniques: TGA, DTA, DSC, DMA – Chemisorption Techniques – TPD, TPO, TPR, TPS.

**L – 45 ; TOTAL HOURS – 45**

#### **TEXT BOOKS:**

1. Skoog D.A., West D.M., Holler F.J. and Crouch S.R., Fundamentals of Analytical Chemistry, 8<sup>th</sup> Edition, Thomson Brooks/Cole Publication., Singapore, 2004.
2. Willard H.H., Merritt L.L., Dean J.A. and Settle F.A., Instrumental Methods of Analysis, 7<sup>th</sup> Edition, CBS Publication, New Delhi Reprint, 2004.
3. Skoog D.A., Holler F.J. and Nieman T.A., Principles of Instrumental Analysis, 5<sup>th</sup> Edition, Harcourt College Publication., Singapore, 1998.
4. Christian G.D., Analytical Chemistry, 6<sup>th</sup> Edition, John Wiley, Singapore, 2003.
5. Fifield F.W. and Kealey D., Principles and Practice of Analytical Chemistry, 5<sup>th</sup> Edition, Blackwell Publication, London, 2000.
6. Settle F. (Editor), Handbook of Instrumental Techniques for Analytical Chemistry, Pearson Education, Singapore, 2004.

#### **COURSE OUTCOMES:**

The student will be able to

**CO1:** analyse the numerical data without error

**CO2:** perform the volumetric and gravimetric analysis of chemical compounds and interpret the result

**CO3:** perform the electro analytical titrations and analyse the result

**CO4:** identify the appropriate spectral technique and do the spectral

analysis and interpret the data

**CO5:** perform the chromatographic techniques and separate the compounds

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on  
22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1	M	M		M											
CO2	H	M		M											
CO3	H	M		M		H									
CO4	H	M		M	M	H									
CO5	H	M		M	M	H									

**Note:** L- Low Correlation M - Medium Correlation H -High Correlation

SDG 6: Clean Water & Sanitation

SDG 7: Affordable and Clean Energy

Statement: Through various analytical methods, innovative, cheap and affordable materials can be developed and can be employed in the area of clean water, sanitation and energy

<b>OEEY 733</b>	<b>BIOMASS FOR ENERGY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 7</b>	<b>APPLICATIONS</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the student conversant with

**COB1:** Structure, properties and applications of cellulose, hemicelluloses and lignin.

**COB2:** Types of biomass used as feedstock for energy applications

**COB3:** Various biomass pretreatment techniques.

**COB4:** Understand the chemistry and economics of biodiesel

**COB5:** Understand the conversion of lignocelluloses into alcohol and fuel.

**MODULE I STRUCTURE AND PROPERTIES OF BIOMASS 9**

Chemistry of polysaccharides, Structure and properties of cellulose, Addition and substitution reactions, Structure and properties of hemicelluloses, Hydrolysis of cellulose by acid and enzyme, Chemistry of lignin, Biosynthesis of lignin, Structure and properties of lignin, Isolation and application of lignin, Chemistry of extractives.

**MODULE II BIOMASS FEEDSTOCKS 9**

Availability and abundance, photosynthesis, composition and energy potential, virgin biomass production and selection, waste biomass (municipal, industrial, agricultural and forestry) availability, abundance and potential, biomass as energy resources: dedicated energy crops, annual crops (maize, sorghum sugar beet, hemp), perennial herbaceous crops (sugarcane, switchgrass, miscanthus), short rotation woody crops (poplar, willow), oil crops and their biorefinery potential, microalgae as feedstock for biofuels and biochemical, enhancing biomass properties for biofuels, challenges in conversion.

**MODULE III BIOMASS PRETREATMENT 9**

Biomass pretreatment - Physical pretreatment methods – milling, microwave, mechanical extrusion, pulse electric field; Chemical pretreatment methods - acid pretreatment, alkali pretreatment, Organosolv pretreatment, Ionic liquids pretreatment; Physico-chemical pretreatment - Steam explosion pretreatment, Ammonia fiber explosion (AFEX) pretreatment; CO<sub>2</sub> explosion, wet oxidation, sulphite pretreatment; Biological pretreatment

**MODULE IV BIODIESEL 9**

Chemistry and Production Processes; Vegetable oils and chemically processed biofuels; Biodiesel composition and production processes; Biodiesel economics; standards for biodiesel quality; Energetics of biodiesel production and effects on greenhouse gas emissions Issues of ecotoxicity and sustainability with ; expanding biodiesel production

## **MODULE V BIOETHANOL**

**9**

**Biochemical conversion of lignocellulose to alcohol**, Separate hydrolysis and fermentation process (SHF), Simultaneous saccharification and fermentation process (SSF), Consolidated Bioprocess (CBP), Pentose fermentation by yeast and bacteria. **Thermochemical conversion of biomass to liquid fuels**, Combustion, Pyrolysis process of lignocellulose to liquid fuels, Gasification process, Cogeneration and polygeneration. Innovative cycles (such as biomass integrated gasification combined cycles, biomass air turbines, humid air turbines etc) for biomass resources, Bioethanol production.

**L – 45; Total Hours –45**

### **REFERENCES:**

1. Krzysztof J Ptasinski, Efficiency of Biomass Energy: An Exergy Approach to Biofuels, Power, and Biorefineries, John Wiley and Sons, 2015.
2. Kaltschmitt, Martin, Energy from Organic Materials (Biomass), A Volume in the Encyclopedia of Sustainability Science and Technology, Second Edition, 2019.
3. George W. Huber, Sara Iborra, AvelinoCorma, Synthesis of Transportation Fuels from Biomass: Chemistry, Catalysts, and Engineering, Chemical Review 2006, 106, 9, 4044-4098.

### **COURSE OUTCOMES:**

Students will be able to

**CO1:** Identify the structure and properties of cellulose, hemicelluloses and lignin.

**CO2:** Find the significance of different biomass resources.

**CO3:** Illustrate biomass pretreatment techniques.

**CO4:** Synthesize the biodiesel at economical price.

**CO5:** Produce the bioethanol at economical price.

**Board of Studies (BoS):****Academic Council:**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022    19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1	M	H	M	M	M	H									
CO2	M	H	M	M	M	H									
CO3	M	H	M	M	M	H									
CO4	M	H	M	M	M	H									
CO5	M	H	M	M	M	H									

**Note:** L - Low Correlation    M - Medium Correlation    H - High Correlation

SDG 7: Affordable & Clean Energy: Ensure access to affordable, reliable, sustainable and modern energy for all.

Statement: Utilization of biomass for the energy need provide solution for affordable and sustainable energy for all.



<b>OEEY 703</b>	<b>BIOMATERIALS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To enable the students understand importance of and properties of Biomaterials

**COB2:** To familiarize the students with different orthopaedic materials.

**COB3:** To understand different cardiovascular materials.

**COB4:** To help students study about materials in ophthalmology

**COB5:** To make the students understand applications of various biomaterials

**MODULE I      BIOLOGICAL PERFORMANCE OF MATERIALS      9**

Biocompatibility- Introduction to the biological environment – Material response: swelling and leaching, corrosion and dissolution, deformation and failure, friction and wear – Host response: the inflammatory process - coagulation and hemolysis- approaches to thrombo- resistant materials development.

**MODULE II      ORTHOPAEDIC MATERIALS      9**

Bone composition and properties - temporary fixation devices - joint replacement – Biomaterials used in bone and joint replacement: metals and alloys – Stainless steel, cobalt based alloys, titanium based materials – Ceramics: carbon, alumina, zirconia, bioactive calcium phosphates, bioglass and glass ceramics – polymers: PMMA, UHMWPE/HDPE, PTFE – Bone cement – Composites.

**MODULE III      CARDIOVASCULAR MATERIALS      9**

Blood clotting – Blood rheology – Blood vessels – The heart – Aorta and valves – Geometry of blood circulation – The lungs - Vascular implants: vascular graft, cardiac valve prostheses, cardiac pacemakers – Blood substitutes – Extracorporeal blood circulation devices.  
probability-internal conversion- nuclear isomerism.

**MODULE IV      DENTAL MATERIALS      9**

Teeth composition and mechanical properties – Impression materials – Bases, liners and varnishes for cavities – Fillings and restoration materials – Materials for oral and maxillofacial surgery – Dental cements and dental amalgams – Dental adhesives.

**MODULE V MATERIALS IN OPHTHALMOLOGY 9**

Biomaterials in ophthalmology – Viscoelastic solutions, contact lenses, intraocular lens materials – Tissue grafts – Skin grafts – Connective tissue grafts – Suture materials – Tissue adhesives – Drug delivery: methods and materials – Selection, performance and adhesion of polymeric encapsulants for implantable sensors- biomemtic materials-Technology from nature.

**L – 45; TOTAL HOURS –45**

**REFERENCES:**

1. Sujata V. Bhat. Biomaterials, Narosa Publication House, New Delhi, 2002.
2. Jonathn Black. Biological Performance of Materials: Fundamentals of biocompatibility, Marcel Dekker Inc, New York, 1992.
3. D.F.Williams (editor). Materials Science and Technology: A comprehensive treatment, Volume 14. Medical and Dental Materials, VCH Publishers Inc, New York, 1992.
4. F.Silver and C.Doillon. Biocompatibility: Interactions of Biological and implantable materials. Volume I Polymers, VCH Publishers Inc, New York, 1989.
5. L.L.Hench and E.C.Ethridge. Biomaterials: An Interfacial Approach, Academic Press, 1982.
6. Joon Park, R. S. Lakes, Biomaterials. An Introduction, Springer, third edition, 2010. Springer

**COURSE OUTCOMES:**

At the end of the course, the students will be able to

**CO1:** importance and properties of biomaterial..

**CO2:** different classes of orthopaedic materials

**CO3:** different types of cardiovascular materials.

**CO4:** various types of materials used in ophthalmology.

**CO5:** applications of various biomaterials

**Board of Studies (BoS) : Academic Council:**

BOS of Physics was held on 19<sup>th</sup> AC held on 29.09.2022  
30.6.22

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1	H	M	L	L	M	M	M	L	L	L	M	M	M	M	M
CO2	H	M	M	L	L	M	L	L	L	L	L	M	M	M	M
CO3	H	M	M	L	L	L	L	L	L	L	L	M	M	M	M
CO4	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M
CO5	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M

**Note:** L- Low Correlation    M -Medium Correlation    H -High Correlation

SDG 4 : Ensuring inclusive and equitable quality education for all persons and promote lifelong learning opportunities.

Statement : The modules and topics mentioned in this course are designed to ensure all inclusive and thorough education with equity to all persons and promote learning opportunities at all times.

<b>OEEY 704</b>	<b>BIOMEDICAL INSTRUMENTATION</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To understand the human physiological systems.

**COB2:** To know the different aspects of biosignal acquisition.

**COB3:** To understand the basics in biopotential recorders.

**COB4:** To know the importance methods, instruments available for biomedical field.

**COB5:** To analyze the special biomedical instrumentation systems.

**MODULE I HUMAN PHYSIOLOGICAL SYSTEMS 9**

Cells and their structure – Nature of Cancer cells – Transport of ions through the cell membrane – Resting and action potentials – Bio-electric potentials – Nerve tissues and organs – Different systems of human body. Biopotential Electrodes and Transducers Design of Medical instruments – components of the biomedical instrument system – Electrodes – Transducers.

**MODULE II BIOSIGNAL ACQUISITION 9**

Physiological signal amplifiers – Isolation amplifiers – Medical preamplifier design – Bridge amplifiers – Line driving amplifier – Current amplifier – Chopper amplifier – Biosignal analysis – Signal recovery and data acquisition – Drift Compensation in operational amplifier – Pattern recognition – Physiological Assist Devices. Pacemakers – Pacemakers batteries – Artificial heart valves – Defibrillators – nerve and muscle stimulators Heart – Lung machine – Kidney machine.

**MODULE III BIOPOTENTIAL RECORDERS 9**

Characteristics of the recording system – Electrocardiography (ECG) – Electroencephalography (EEG) – Electromyography (EMG) – Electroethnography (ERG) and Electroculography (EOG) – Recorders with high accuracy – recorders for OFF line analysis.

**MODULE IV OPERATION THEATRE EQUIPMENT 9**

urgical diathermy- shortwave diathermy – Microwave diathermy – Ultrasonic disathermy – Therapeutic effect of heat – Range and area of irritation of different techniques – Ventilators – Anesthesia machine – Blood flowmeter –

Cardiac Output measurements – Pulmonary function analyzers – Gas analyzers – Blood gas analyzers – Oximeters – Elements of intensive care monitoring.

## **MODULE V SPECIALISED MEDICAL EQUIPMENTS**

**9**

Blood Cell counter – Electron microscope – Radiation detectors – Photometers and colorimeters – digital thermometer – audiometers – X-rays tube – X-ray machine – image intensifiers – Angiography – Application of X-ray examination. Safety instrumentation: Radiation safety instrumentation – Physiological effects due to 50Hz current passage – Microshock and macroshock – electrical accident Hospitals – Devices to protect against electrical hazards – Hospitals architecture.

**L – 45; TOTAL HOURS –45**

### **REFERENCES:**

1. Arumugam M., Biomedical Instrumentation, Anurada Agencies Publishers, 1992.
2. Khandpur R.S., Handbook of Biomedical Instrumentation, Third Edition, Tata McGraw-Hill Education, 2014.
3. Shakti Chatterjee and Aubert Miller, Biomedical Instrumentation Systems, Cengage Learning Publisher, 2010.
4. Gromwell L., Fred J. Weibell, Erich A. Pfeiffer, Biomedical Instrumentation and Measurements, Second Edition, Prentice Hall, 1980.

### **COURSE OUTCOMES:**

At the end of the course, the students will be able to

**CO1:** the human physiological systems.

**CO2:** the different aspects of biosignal acquisition.

**CO3:** different biopotential recorders such as EEG, ECG, EMG, EOG

**CO4:** biomedical instruments involved in advanced operation theatres

**CO5:** the application of biomaterials towards specialized medical equipment such as electron microscope and radiation detectors

### **Board of Studies (BoS) :**

BOS of Physics was held on 30.6.22

### **Academic Council:**

19<sup>th</sup> AC held on  
29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PSO1	PSO2	PSO3
CO1	H	M	L	L	M	M	M	L	L	L	M	M	M	M	M
CO2	H	M	M	L	L	M	L	L	L	L	L	M	M	M	M
CO3	H	M	M	L	L	L	L	L	L	L	L	M	M	M	M
CO4	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M
CO5	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M

**Note:** L- Low Correlation    M -Medium Correlation    H -High Correlation

SDG 4 : Ensuring inclusive and equitable quality education for all persons and promote lifelong learning opportunities.

Statement : The modules and topics mentioned in this course are designed to ensure all inclusive and thorough education with equity to all persons and promote learning opportunities at all times.

<b>OEEY 705</b>	<b>BIOPHOTONICS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To know the role of light and its interaction in the cells and tissues.

**COB2:** To understand the different imaging techniques for the biological systems.

**COB3:** To know the concepts of spectroscopy in biological applications.

**COB4:** To understand the optical force spectroscopy.

**COB5:** To understand the role of Biophotonic materials in applications.

**MODULE I INTERACTION OF LIGHT WITH BIOLOGICAL SYSTEMS 9**

Interaction of light with cells, tissues, nonlinear optical processes with intense laser beams, photo-induced effects in biological systems.

**MODULE II IMAGING TECHNIQUES 9**

Imaging techniques: Light microscopy, wide-field, laser scanning - confocal, multiphoton, fluorescence lifetime imaging, FRET imaging - Frequency-Domain lifetime imaging. Cellular Imaging - Imaging of soft and hard tissues and other biological structures.

**MODULE III SINGLE MOLECULE SPECTROSCOPY 9**

Single molecule spectroscopy: UV-VIS spectroscopy of biological systems, single molecule spectra and characteristics – IR and Raman spectroscopy and Surface Enhanced Raman Spectroscopy for single molecule applications.

**MODULE IV OPTICAL FORCE SPECTROSCOPY 9**

Optical Force Spectroscopy: Generation optical forces – Optical trapping and manipulation of single molecules and cells in optical confinement - Laser trapping and dissection for biological systems - single molecule biophysics, DNA protein interactions.

**MODULE V BIOSENSORS 9**

Biosensors, Principles- DNA based biosensors – Protein based biosensors– materials for biosensor applications- fabrication of biosensors.

**L – 45; TOTAL HOURS –45**

**REFERENCES:**

1. Prasad. P.N., Introduction to Biophotonics, John Wiley & Sons, 2003
2. Michael P. Sheetz, Laser Tweezers in Cell Biology (Methods in Cell Biology), Vol.55, Academic Press Publishers, 1997.
3. Ranier .W, Nanoelectronics and Information Technology, Wiley Publishers, 2012.
4. Drexler. K.E., Nanosystems: Molecular Machinery, Manufacturing and Computation, Wiley Publishers, 1992.

#### **COURSE OUTCOMES:**

At the end of the course, the students will be able to

**CO1:** Make clear insights into the applications of light interaction with biological systems.

**CO2:** Compare different imaging techniques

**CO3:** Understand and analyse the various spectroscopic techniques used in biological system.

**CO4:** Effectively grasp the usage of the optical force spectroscopy.

**CO5:** Get clear ideas and communicate about the importance of use of spectroscopy in design of bio-photonic devices.

#### **Board of Studies (BoS) :**

BOS of Physics was held on 30.6.22

#### **Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PSO1	PSO2	PSO3
CO1	H	M	L	L	M	M	M	L	L	L	M	M	M	M	M
CO2	H	M	M	L	L	M	L	L	L	L	L	M	M	M	M
CO3	H	M	M	L	L	L	L	L	L	L	L	M	M	M	M
CO4	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M
CO5	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M

**Note:** L- Low Correlation M -Medium Correlation H -High Correlation

SDG 4 : Ensuring inclusive and equitable quality education for all persons and promote lifelong learning opportunities.

Statement : The modules and topics mentioned in this course are designed to ensure all inclusive and thorough education with equity to all persons and promote learning opportunities at all times.



<b>OEEY 734</b>	<b>CORROSION AND CORROSION</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 9</b>	<b>CONTROL</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the student conversant with the:

**COB1:** Causes and theories of corrosion.

**COB2:** Different types of corrosion.

**COB3:** Basic concepts to prevent corrosion and testing of corrosion by various diagrams.

**COB4:** Factors influencing corrosion.

**COB5:** Control of corrosion using various methods.

**MODULE I CORROSION 9**

Causes and effects of corrosion — theories of corrosion – Dry corrosion — oxidation — direct atmospheric effect – Hydrogen corrosion, liquid metal corrosion and corrosion by other gases-electrochemical corrosion – hydrogen evolution – presence and absence of oxygen – corrosion by gaseous reduction.

**MODULE II FORMS OF CORROSION 9**

Eight forms of corrosion- Galvanic bimetallic corrosion – differential aeration corrosion – concentration cell corrosion – erosion corrosion – pitting corrosion – underground soil corrosion – intergranular corrosion – stress corrosion: Types - seasonal cracking of alloys and caustic embrittlement – corrosion fatigue.

**MODULE III CORROSION TESTING 9**

Rate of corrosion – calculation of  $\Delta G$  and other related thermodynamic parameters – potential measurement – electrochemical series – redox reactions – EMF measurement and corrosion current – anodic and cathodic behaviour of metals – passivity – testing of virgin metals – alloy – Pourbaix and Evans diagrams.

**MODULE IV FACTORS INFLUENCING CORROSION 9**

Nature of metal — over voltage — areas of anodic/cathodic — purity of metal — physical state of metals — passive nature of metal — solubility — volatility of corrosion products – corroding environment – influence of pH – Pourbaix diagrams- ions – formations of cells – polarization of electrodes.

**MODULE V                    CORROSION CONTROL                    9**

Design – selection of materials – pure metals and alloys – annealing – elimination of galvanic action – cathodic protection – sacrificial anodic protection and impressed current cathodic protection – modification of environment: deaeration and dehumidification – corrosion inhibitors – protective coatings : preparation of surface before applying coatings – Classification: Inorganic coatings- metallic and non-metallic – organic coatings – special paints – varnish, enamel and lacquers.

**L – ; TOTAL HOURS –45**

**TEXT BOOKS:**

1. C.G. Munger and Louis D. Vincent, “Corrosion Prevention by Protective Coatings”, Third Edition (e-Book), 2014.

**REFERENCES:**

2. M.G. Fontana and N.G. Green, Corrosion Engineering, McGraw Hill BookCompany, New York, 1984.
3. J.H. Brophy, R.M. Rose and J. Walf, The Structure and Properties of Materials, Wiley Inter Science Inc., New York, 1984.
4. B.T. Kelly, Irradiation Diamagneto Solids, Pergamon Press, New York,1992.
5. D.R. Cross, Principles and Applications of Electrochemistry, Chapman andHall, UK, 1988.

**COURSE OUTCOMES:**

Students will become familiar with the

**CO1:** basic concepts of corrosion

**CO2:** Different types of corrosion and their mechanism of corrosion

**CO3:** Testing and evaluation of corrosion

**CO4:** Factors which influence the corrosion

**CO5:** Control of corrosion in real situation.

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1					L										
CO2															
CO3		H						M							
CO4															
CO5										H					

Note: L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 9 : Build resilient Infrastructure, promote inclusive and sustainable industrialization and foster innovation.

The holistic understanding of corrosion and its prevention leads to construction of resilient infrastructure and sustainable industrialization.

<b>OEEY 735</b>	<b>CORROSION SCIENCE AND</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>	<b>TECHNOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To enable the students understand principles behind corrosion science.

**COB2:** To expose the students to various instrumental techniques.

**COB3:** To familiarize the students with methods of coating

**COB4:** To help the students in the corrosion in selected environments

**COB5:** To make the students to understand various corrosion processes and engineering applications.

**MODULE I CORROSION PROCESSES 9**

Basic principles of electrochemistry and aqueous corrosion processes - Electrochemical Thermodynamics and Electrode Potential - Electrochemical Kinetics of Corrosion Cathodic and anodic behavior - Faraday's Law - Nernst equation; standard potentials Pourbaix diagram - Tafel equations, corrosion rate - Evans diagram - pitting, crevice and exfoliation corrosion; influence of deposits and anaerobic conditions; corrosion control; high temperature oxidation and hot corrosion; corrosion/mechanical property interactions.

**MODULE II ORTHOPAEDIC MATERIALS 9**

ray diffraction, TEM, SEM and EDX, WDX analysis, surface analysis by AES, XPS and SIMS, overview of other techniques.

**MODULE III CARDIOVASCULAR MATERIALS 9**

Electrodeposition; flame and plasma spraying; thermal, HV of detonation gun, gas dynamic spray, physical vapour deposition; chemical vapour deposition; HIP surface treatments devices, probability-internal conversion- nuclear isomerism.

**MODULE IV DENTAL MATERIALS 9**

Atmospheric Corrosion, Corrosion in Automobiles, Corrosion in Soils, Corrosion of Steel in Concrete, Corrosion in Water, Microbiologically Induced Corrosion, Corrosion in the Body, Corrosion in the Petroleum Industry, Corrosion in the Aircraft Industry, Corrosion in the Microelectronics Industry

**MODULE V MATERIALS IN OPHTHALMOLOGY 9**

Abrasive, erosive and sliding wear. The interaction between wear and corrosion.

Coating systems for corrosion and wear protection; new coating concepts including multi-layer structures, functionally gradient materials, intermetallic barrier coatings and thermal barrier coatings.

**L – 45; TOTAL HOURS –45**

**REFERENCES:**

1. D.A. Jones, Principles and Prevention of Corrosion, 2nd Edition, Macmillan Publishing Co., 1995.
2. J.O.M. Bockris, B.E. Conway, E. Yeager and White, Electrochemical Materials Science in Comprehensive Treatise of Electrochemistry, Volume 4, Plenum press, 2001.
3. M.G. Fontanna and N.D. Greene, Corrosion Engineering, McGraw-Hill publishing, 1978
4. I.M. Hutchings, Tribology: Friction and Wear of Engineering Materials, CRC press, Boca Raton, 1992 D.O. Sprowds, Corrosion Testing and Evaluation, Corrosion Metals Hand book, vol. 13, 1986.

**COURSE OUTCOMES:**

At the end of the course, the students will be able to

**CO1:** various corrosion process involved in electrochemistry

**CO2:** working mechanism of various instrumentation techniques

**CO3:** various coating process,

**CO4:** applications of coatings towards environmental safety

**CO5:** industrial applications of coatings

**Board of Studies (BoS) :**

BOS of Physics was held on 30.6.22

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PSO1	PS O2	PS O3
CO1	H	M	L	L	M	M	M	L	L	L	M	M	M	M	M
CO2	H	M	M	L	L	M	L	L	L	L	L	M	M	M	M
CO3	H	M	M	L	L	L	L	L	L	L	L	M	M	M	M
CO4	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M
CO5	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M

**Note:** L- Low Correlation M -Medium Correlation H -High Correlation

SDG 4 : Ensuring inclusive and equitable quality education for all persons and promote lifelong learning opportunities.

Statement : The modules and topics mentioned in this course are designed to ensure all inclusive and thorough education with equity to all persons and promote learning opportunities at all times.

<b>OEEY 736</b>	<b>ENVIRONMENTAL CHEMISTRY</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 13</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the student conversant with

**COB1:** Understand the issue of chemicals based pollution.

**COB2:** Understand the chemicals mobility in aquatic systems.

**COB3:** Understand contaminating chemicals in air and their fate.

**COB4:** Understand the type of soil contaminants and provide remediation.

**COB5:** Identify emerging environmental contaminants including speciation

**MODULE I FUNDAMENTALS 9**

Stoichiometry and mass balance-Chemical equilibria, acid base, solubility product ( $K_{sp}$ ), heavy metal precipitation, amphoteric hydroxides,  $CO_2$  solubility in water and species distribution – Ocean acidification, Chemical kinetics, First order- 12 Principles of green chemistry.

**MODULE II AQUATIC CHEMISTRY 9**

Water and wastewater quality parameters- environmental significance and determination; Fate of chemicals in aquatic environment, volatilization, partitioning, hydrolysis, photochemical transformation– Degradation of synthetic chemicals - Metals, complex formation, oxidation and reduction, pE – pH diagrams, redox zones – sorption- Colloids, electrical properties, double layer theory, environmental significance of colloids, coagulation

**MODULE III ATMOSPHERIC CHEMISTRY 9**

Atmospheric structure – chemical and photochemical reactions – photochemical smog. Ozone layer depletion – greenhouse gases and global warming,  $CO_2$  capture and sequestration – acid rain- origin and composition of particulates. black carbon, air quality parameters determination.

**MODULE IV SOIL CHEMISTRY 9**

Nature and composition of soil - Clays- cation exchange capacity-acid base and ion-exchange reactions in soil – agricultural chemicals in soil-reclamation of contaminated land; salt by leaching- Heavy metals by electrokinetic remediation.

**MODULE V EMERGING POLLUTANTS 9**

Heavy metals-chemical speciation –Speciation of Hg & As- endocrine

disturbing chemicals- Pesticides, Dioxins & Furan, PCBs ,PAHs and Fluro compounds toxicity- Nano materials, CNT, titania, composites ,environmental applications.

**L – 45; Total Hours –45**

**REFERENCES:**

1. Sawyer, C.N., Mac Carty, P.L. and Parkin, G.F., "Chemistry for Environmental Engineering and Science", Tata McGraw – Hill, Fifth edition, New Delhi 2003.
2. Colin Baird,, Environmental Chemistry, Freeman and company, New York, 5th Edition,2012.
3. Manahan, S.E., "Environmental Chemistry", Ninth Edition, CRC press, 2009.
4. Ronald A. Hites ,"Elements of Environmental Chemistry", Wiley, 2nd Edition,2012.

**COURSE OUTCOMES:**

Students will be able to

**CO1:** In solving environmental issues of chemicals based pollution.

**CO2:** To determine chemicals mobility in aquatic systems.

**CO3:** To identify contaminating chemicals in air and their fate.

**CO4:** Understand the type of soil contaminants and provide remediation.

**CO5:** Identify emerging environmental contaminants including speciation

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO1 0	PO1 1	PO1 2	PSO 1	PSO 2	PSO 3
CO 1	M	H	M	M	M	H									
CO 2	M	H	M	M	M	H									
CO 3	M	H	M	M	M	H									
CO 4	M	H	M	M	M	H									
CO 5	M	H	M	M	M	H									

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 13: Climate Action: Take urgent action to combat climate change and its impacts

Statement: Understanding of environmental chemistry will lead to take necessary changes for maintaining a healthy environment.



<b>OEEY 737</b>	<b>FUEL CELLS FOR SUSTAINABLE</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 7,11</b>	<b>ENERGY PRODUCTION</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

The student will

**COB1:** familiar with the types of fuel cell

**COB2:** familiar with the components of fuel cells

**COB3:** understand the performance for fuel cells

**COB4:** learn the methods of production storage of hydrogen

**COB5:** learn the sustainability and applications of fuel cells

**MODULE I INTRODUCTION AND TYPES OF FUEL CELLS 9**

Introduction - definition - history - difference between batteries and fuel cells - chemistry of fuel cells - classification of fuel cell (based on temperature and electrolyte) - types of fuel cell: polymer electrolyte membrane or proton exchange membrane fuel cell (PEMFC), direct methanol fuel cell (DMFC), alkaline fuel cell (AFC), phosphoric acid fuel cell (PAFC), molten carbonate fuel cell (MCFC) and solid oxide fuel cells (SOFC)

**MODULE II FUEL CELL COMPONENTS 9**

Membrane electrode assembly components: membranes and ionomers, fuel cell electrodes and gas diffusion layer, fuel cell electrocatalysts (type and synthesis) - bi-polar plates, humidifiers and cooling plates - *phase-change materials* (PCMs) for thermal packaging - fuel cell stack - Balance of plant - Seals and insulation – Safety.

**MODULE III FUEL CELLS PERFORMANCE AND APPLICATIONS 9**

Thermodynamics of fuel cells - electrochemical kinetics of fuel cells - Fuel cell efficiency - performance characteristics: voltage efficiency - effect of voltage with current density for low and high temperature fuel cells - causes for voltage losses – introduction to fuel cycle analysis - Mass balance

**MODULE IV PRODUCTION AND STORAGE OF HYDROGEN FUEL 9**

Hydrogen as energy source - its merit as a fuel - hydrogen production: steam reforming, partial oxidation, coal gasification/thermal reforming, fuel cell technology based on bio-mass - hydrogen storage: compressed hydrogen, liquid hydrogen, metal hydrides, carbon fibers - safety and management of hydrogen

**MODULE V FUEL CELL APPLICATIONS AND SUSTAINABILITY 9**

Fuel cell material recycle, durability, lifetime issues - Critical issues, adoption, future technologies - distributed power generation - grid-connect applications - non-grid connect applications - combined heat and power (CHP) - economic and environmental analysis - Control of contaminants: CO and sulphur - future trends of fuel cells - Sustainability of Hydrogen Fuel Cell Electric Vehicles.

**L – 45; TOTAL HOURS – 45**

**TEXT BOOKS:**

1. R.H. Thring (Editor), Fuel Cells for Automotive Applications, Professional Engineering Publishing UK, 2004.
2. Gregor Hoogers (Editor), Fuel Cell Technology Handbook, SAE International, CRC Press, 2003.
3. Vladimir S. Bagotsky, Fuel Cells: Problems and Solutions, 2<sup>nd</sup> Edition, John Wiley and Sons, 2012.
4. B. Viswanathan and M. Aulice Scibioh, Fuel Cells: Principles and Applications, Taylor and Francis Group, 2007.
5. Supramaniam Srinivasan, Fuel cells: From Fundamentals to Applications, Springer, 2006.
6. Prospects for Hydrogen and Fuel Cells, International Energy Agency, OECD Publishing, 2005.

**COURSE OUTCOMES:**

The student will be able to

**CO1:** classify fuel cells and elaborate the different types of fuel cells.

**CO2:** explain the components of the fuel cells and can synthesise electrocatalysts for the system

**CO3:** calculate the open circuit voltage, efficiency and voltage losses, explain fuel cycle analysis and mass balance

**CO4:** suggest the suitable methods of production and storage of hydrogen for fuel cells.

**CO5:** find application of fuel cells for variety of application and practice on sustainable environment

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1	L	M													
CO2		M			M										
CO3		M		L											
CO4	M		L			M									
CO5			M												

**Note:** L- Low Correlation    M -Medium Correlation    H -High Correlation

SDG 7: ensure access to affordable, reliable, sustainable and modern energy for all.

SDG 11: Make cities and human settlements inclusive, safe, resilient and sustainable

<b>OEEY 738</b>	<b>GREEN AND SUSTAINABLE</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4, 7, 9</b>	<b>CHEMISTRY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the student conversant with

**COB1:** understand the principle and concepts of green chemistry

**COB2:** various alternative (non-traditional) reagents and chemicals for green synthesis.

**COB3:** understand the non-conventional energy sources for green synthesis

**COB4:** understand the uses of eco-benign solvents - alternative to organic solvents

**COB5:** synthesis of nanomaterials using green chemistry approaches

**MODULE I INTRODUCTION, PRINCIPLE AND CONCEPTS OF GREEN CHEMISTRY 9**

Need for green chemistry; Inception and evolution of green chemistry; Twelve principles of green chemistry with their explanations and examples; Designing a green synthesis using these principles; Green chemistry in day to day life.

**MODULE II NON-TRADITIONAL GREENER ALTERNATIVE APPROACHES 9**

Different approaches to green synthesis: (a) Uses of green reagents in organic synthesis - Dimethyl carbonate, polymer supported reagents - peracids and chromic acid; (b) Green catalysts, role of catalysis in sustainable development, homogeneous and heterogeneous catalysts; Introduction, advantages and applications of (i) Nanocatalysts, (ii) Phase transfer catalysts, (iii) Biocatalysts, (iv) Organocatalysts, in organic synthesis.

**MODULE III APPLICATIONS OF NON-CONVENTIONAL ENERGY SOURCES 9**

Introduction of microwave induced synthesis: Microwave activation, equipment, time and energy benefits, and limitations. Organic transformations under microwaves - Fries rearrangement, Diels-Alder reaction, decarboxylation, saponification of ester, alkylation of reactive methylene compounds; Heterocyclic synthesis - pyrrole, quinoline. Introduction of ultrasound assisted green synthesis: Instrumentation,

physical aspects, applications in organic transformations.

#### **MODULE IV ENVIRONMENTALLY BENIGN SOLUTIONS TO ORGANIC SOLVENTS 9**

Ionic liquids as green solvents: Introduction, properties and types of ionic liquids. Synthetic applications Diels-Alder reaction, epoxidation and Heck reaction.

Aqueous phase reactions: Enhancement of selectivity, efficiency. Synthetic applications - 1,3-Dipolar Cycloadditions, Carbon-Carbon bond-forming processes and bromination reactions.

Fluorous solvents in green chemistry: Scope, definition and their synthetic applicability. Role of supercritical carbon dioxide in green chemistry.

Ethyl lactate as a renewable green solvent: Properties and applications.

#### **MODULE V GREENER SYNTHESIS OF NANOMATERIALS 9**

Greener synthesis of Nanomaterials— Microwave assisted synthesis of Quantum Dots (QD) and nano catalysts in aqueous medium, Magnetic Nanoparticles. Synthesis of Nanoparticles using Bacteria, Yeast, Algae and Fungus.

**L – 45; TOTAL HOURS –45**

#### **TEXT BOOKS:**

1. Green Chemistry – Environmentally benign reactions – V. K. Ahluwalia. Ane Books India (Publisher). (2006).
2. Green Chemistry – Designing Chemistry for the Environment – edited by Paul T. Anastas & Tracy C. Williamson. Second Edition, (1998).
3. Green Chemistry – Frontiers in benign chemical synthesis and processes- edited by Paul T. Anastas & Tracy C. Williamson. Oxford University Press, (1998).
4. Green Chemistry – Environment friendly alternatives- edited by Rashmi Sanghi & M. M. Srivastava, Narora Publishing House, (2003).
5. Sheldon, R.A., Arends, I., and Hannefed, U., Green Chemistry and Catalysis, Wiley-VCH Verlag GmbH and Co. (2007).
6. Anastas, P., and Williamson, T. C., Green Chemistry Frontiers in Benign Chemical Synthesis and Processes, Oxford University Press (1999).
7. Ahluwalia, V. K., and Kidwai, M., New Trends in Green Chemistry, Anamaya Publishers (2004)

**COURSE OUTCOMES:**

The students will be able to

**CO1:** understanding of the 12 principles of green chemistry to improve the sustainability performance of the products/ materials

**CO2:** use various alternative reagents and chemicals for green synthesis.

**CO3:** apply non-conventional energy sources for the synthesis of organic compounds and materials.

**CO4:** use eco-begin solvents for the synthesis of organic compounds and materials

**CO5:** understand the synthesis of nanomaterials using greener methods

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1	H				L										
CO2			M												
CO3		H						M							
CO4		H													
CO5				L						H					

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 4 & Ensure inclusive and equitable quality education and promote lifelong learning opportunities for all.

SDG 7 & Ensure access to affordable, reliable, sustainable and modern energy for all.

SDG 9 : Build resilient Infrastructure, promote inclusive and sustainable industrialization and foster innovation

Statement : The holistic understanding of green chemistry principles and concepts to sustainable development in the field of synthetic and materials chemistry.

<b>OEEY 739</b>	<b>INDUSTRIAL POLLUTION CONTROL</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG:</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>
<b>6,7,9,11,</b>					
<b>12,13 and 15</b>					

**COURSE OBJECTIVES:**

This course will enable students to:

**COB1:** understand the environmental pollution, environmental emission standards and the laws and rules.

**COB2:** understand the concept of pollution prevention.

**COB3:** Understand various air pollution control methods.

**COB4:** Understand various water pollution control methods of primary and secondary treatment.

**COB5:** understand the biological treatment, tertiary treatment and solid wastes disposal.

**MODULE I EMISSION STANDARDS AND ENVIRONMENTAL LAWS 8**

Environment and environmental pollution from chemical process industries- Air pollutants and pollution and its effects- , characterization of emissions, water pollutants and pollution- and its effects- characterization of effluents- standards for ambient air, noise emission and effluents-Environmental Laws and rules : Air act 1981 and 1987, water act 1974,1977,1987, environmental protection act 1986, The hazardous wastes (management and handling) rules, 1989 & 2000, The manufacture, storage and import of hazardous chemical rules, 1989 & 2000, Public liability insurance act, 1991.The national environment tribunal act, 1995,The chemical accidents (emergency planning, preparedness and response rules, 1996, The recycled plastic manufacture and usage rules, 1999,The batteries (management and handling (draft) rules, 2000.

**MODULE II POLLUTION PREVENTION 10**

Process modification: process change, technology change, better process control and product modification- alternative raw material - recovery of by-product from industrial emission effluents- waste reduction techniques: recycle and reuse of waste and volume reduction- energy recovery and waste utilization- Material and energy balance for pollution minimization- Water use minimization- Fugitive emission/effluents and leakages and their control- LDAR programmes- housekeeping and maintenance.

**MODULE III AIR POLLUTION CONTROL****9**

Introduction to air pollution control- Particulate emission control by mechanical separation: gravitational settling chambers, cyclone separators, fabric filters and electrostatic precipitator and wet gas scrubbing, gaseous emission control by absorption and adsorption, Design of cyclones, ESP, fabric filters and absorbers.

**MODULE IV WATER POLLUTION CONTROL****10**

Introduction to Water Pollution and Control - Physical treatment, pre-treatment, solids removal by setting and sedimentation, filtration centrifugation, coagulation and flocculation Secondary treatment: Biological treatment- Anaerobic and aerobic treatment - Trickling filter, activated sludge and lagoons, aeration systems.

**MODULE V TERTIARY TREATMENT AND SOLID DISPOSAL****8**

Tertiary treatment: colour and odour removal - **Solids Disposal:** Sludge separation and drying- Solids waste disposal – composting, landfill, briquetting / gasification and incineration.

**L – 45; TOTAL HOURS – 45****TEXT BOOKS:**

1. Paul N Cheremisinoff, Air pollution control and design for industry, 2018.
2. Rao. C.S, Environmental Pollution control Engineering, 2007.

**REFERENCES:**

1. Thomas T. Shen, Industrial Pollution Prevention, Springer, 1999.
2. Nancy J. Sell, Industrial Pollution Control: Issues and Techniques, 2nd Edition, Wiley, 1992.
3. Pollution Control Law Series: Pollution Control Acts, Rules and Notification Issued There under, Central Pollution Control Board, Ministry of Environment and Forest, Government of India, 2006.
4. www.moef.nic.in.

**COURSE OUTCOMES:**

Students will be able to describe

**CO1:** environmental pollution and the environmental standards.



**CO2:** the concept of pollution prevention.

**CO3:** various air pollution control methods.

**CO4:** The pre and secondary treatment of water pollution control methods.

**CO5:** various tertiary treatment and solid wastes disposal methods.

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on  
22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1			H												
CO2			H			M									
CO3					M										
CO4					M										
CO5					M										

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 7 : Ensure access to affordable, reliable, sustainable and modern energy for all

Statement: The holistic understanding of recycling materials and technology leads to provide modern renewable energy and sustainable industrialization.

<b>OEEY 740</b>	<b>INTRODUCTION TO EMBEDDED</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4,9</b>	<b>SYSTEM</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To describe the embedded system concepts with its hardware and software architectures.

**COB2:** To analyze the significance of memory and interrupts in an embedded system

**COB3:** To discuss the software development tools necessary for embedded systems.

**COB4:** To interpret the programming model in embedded system

**COB5:** To compare the concepts of OS and RTOS.

**PREREQUISITES:** Basics concepts of Microprocessor

**MODULE I INTRODUCTION OF EMBEDDED SYSTEM 9**

Introduction –Embedded Systems vs. General computing systems- Fundamental Components of embedded systems- Characteristics- Challenges-Examples- Embedded System design process.

**MODULE II EMBEDDED COMPUTING PLATFORM 9**

Overview of Processors and hardware units in an embedded system-CPU buses – Memory devices –Memory types- I/O devices – Interrupts and types.

**MODULE III EMBEDDED SOFTWARE DEVELOPMENT PROCESS 9 and TOOLS**

Development process of an embedded system-software modules and tools for implementation of an embedded system- Integrated development environment- Host and target machines-cross compiler-cross assembler-Debugging mechanisms.

**MODULE IV PROGRAM MODELING IN EMBEDDED SYSTEMS 9**

Program Models – Data Flow Graph model-control DFG model-Synchronous DFG model- Finite state machines- UML modeling – UML Diagrams.

**MODULE V REAL TIME OPERATING SYSTEMS (RTOS) 9**

Overview of Operating Systems (OS) concepts – Real time systems –Types -Need for RTOS in Embedded Systems -Compare OS and RTOS- Multiple Tasks and Multiple Processes-Priority-Based Scheduling- Real time scheduling algorithm – Inter process Communication Mechanisms- Case study.

**L –45 ; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Marilyn Wolf, "Computers as components ", Elsevier, 2016.
2. K.V. Shibu , Introduction to Embedded Systems, McGraw Hill Education India Private Limited; Second edition, 2017.

**REFERENCES:**

1. Qing Li and Carolyn Yao, "Real-Time Concepts for Embedded Systems", CMP Books, 2003.

**COURSE OUTCOMES:**

At the end of the course, the student will be able to:

**CO1:** identify the suitable processor and peripherals for embedded applications

**CO2:** discuss the software development tools and process.

**CO3:** draw the programming model for embedded systems

**CO4:** analyze the role of RTOS for embedded applications

**CO5:** design real time embedded application.

**Board of Studies (BoS) :**

24<sup>th</sup> BOS of ECE held on 08.02.2023.

**Academic Council:**

20<sup>th</sup> AC held on 13.04.2023

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PSO 1	PSO 2	PSO 3
CO1	H	H	H	H	H	H	M	M	M	M	M	M	H	H	H
CO2	H	H	H	H	H	H	M	M	M	M	M	M	H	H	H
CO3	H	H	H	H	H	H	M	M	M	M	M	M	H	H	H
CO4	H	H	H	H	H	H	M	M	M	M	M	M	H	H	H
CO5	H	H	H	H	H	H	M	M	M	M	M	M	H	H	H

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

**SDG 4:** Ensure inclusive and equitable quality education and promote lifelong learning opportunities for all.

Statement: Understanding of the real time systems will bring practical knowledge on quality education.

**SDG 9:** Build resilient Infrastructure, promote inclusive and sustainable industrialization and foster innovation

Statement: capable of promoting industrialization through the application of real-time system design principles.

<b>OEEY 741</b>	<b>MATLAB PROGRAMMING</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 9</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To create the mathematical model for specific systems

**COB2:** Discuss the computer tools to solve mathematical models for specific systems.

**COB3:** Develop models to solve linear and non linear differential equations

**COB4:** To define the system or process through MATLAB

**COB5:** To describe MATLAB graphic feature and its applications.

**PREREQUISITES:** Mathematics

**MODULE I INTRODUCTION TO MATLAB AND DATA PRESENTATION 9**

Introduction to MATLAB-Vectors, Matrices -Vector/Matrix Operations & Manipulation Functions vs scripts- Making clear and compelling plots-Solving systems of linear equations numerically and symbolically.

**MODULE II ROOT FINDING AND MATLAB PLOT FUNCTION 9**

Linearization and solving non-linear systems of equations- The Newton-Raphson method- Integers and rational numbers in different bases- Least squares regression - Curve fitting-Polynomial fitting and exponential fitting.

**MODULE III LINEAR AND NON-LINEAR DIFFERENTIAL EQUATIONS 9**

Numerical integration and solving first order, ordinary differential equations (Euler's method and Runge-Kutta)- Use of ODE function in MATLAB- Converting second order and higher ODEs to systems of first order ODEs- Solving systems of higher order ODEs via Euler's method and Runge-Kutta)- Solving single and systems of non-linear differential equations by linearization-Use of the function ODE in MATLAB to solve differential equations - Plot Function –Saving & Painting Plots.

**MODULE INTRODUCTION OF SIMULINK 9**  
**IV**

Simulink & its relations to MATLAB – Modeling a Electrical Circuit- Modeling a fourth order differential equations- - Representing a model as a subsystem- Programme specific Simulink demos.

**MODULE V APPLICATION OF MATLAB 9**

Linear algebraic equations – elementary solution method – matrix method for linear equation – Cramer’s method – Statistics, Histogram and probability – normal distribution – random number generation – Interpolation – Analytical solution to differential equations – Numerical methods for differential equations.

**L –45 ; TOTAL HOURS –45**

**TEXT BOOKS:**

1. Attaway, Stormy. Matlab: a practical introduction to programming and problem solving. Butterworth-Heinemann, 2013.
2. Chapra, Steven. EBOOK: Applied Numerical Methods with MATLAB for Engineers and Scientists. McGraw Hill, 2011.
3. Singh, Harendra, Devendra Kumar, and DumitruBaleanu, eds. Methods of mathematical modelling: fractional differential equations. CRC Press, 2019.

**REFERENCES:**

1. Hossain, Eklas. "Introduction to Simulink." In MATLAB and Simulink Crash Course for Engineers, Springer International Publishing, 2022.
2. Sumathi, Sai, and Surekha Paneerselvam. Computational intelligence paradigms: theory & applications using MATLAB. CRC Press, 2010.

**COURSE OUTCOMES:**

On completion of the course, students will be able to

**CO1:** analyze mathematical model for specific systems

**CO2:** develop a code for specific tasks.

**CO3:** write MATLAB programs for solving linear and nonlinear systems

**CO4:** model a specific system using simulink

**CO5:** apply built in functions for the wide range of applications

**Board of Studies (BoS) :**

24<sup>th</sup> BOS of ECE held on 08.02.2023.

**Academic Council:**

20<sup>th</sup> AC held on 13.04.2023

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO11	PO 12	PSO1	PSO2	PSO3
CO 1	H	H	M	M	L	L	L	L	L	L	L	L	H	H	H
CO 2	M	H	M	M	L	L	L	L	L	L	L	L	H	H	H
CO 3	M	M	L	M	L	L	L	L	L	L	L	L	H	H	H
CO 4	H	M	M	M	L	L	L	L	L	L	L	L	H	H	H
CO 5	H	H	M	M	L	L	L	L	L	L	L	L	H	H	H

**Note:** L- Low Correlation    M -Medium Correlation    H -High Correlation

SDG 9 : Build resilient Infrastructure, promote inclusive and sustainable industrialization and foster innovation

Statement : Describes the methodology to apply modern tools for solving the mathematical models which promotes sustainable industrialization and foster innovation

<b>OEEY 710</b>	<b>NANOTECHNOLOGY AND CATALYSIS</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 6,7,9,15</b>		<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the student conversant with

**COB1:** basic knowledge on nanoscience and nanotechnology which includes the exotic properties of materials at nanoscale including various techniques for the processing of nanomaterials

**COB2:** various techniques available for the characterization of nanostructured materials

**COB3:** applications in selected fields and impacts of nanotechnology in ecosystem

**COB4:** Impart the basic concepts involved in catalytic processes.

**COB5:** Understand the importance of heterogeneous catalysis.

### **MODULE I INTRODUCTION AND PREPARATION OF 9 NANOMATERIALS**

Introduction to nanomaterials, Properties of nanomaterials, Nanostructures: Zero-, One-, Two- and Three-dimensional structures, Surface Plasmon Resonance, Change of bandgap; Methods of preparation of nanomaterials, top-down approach and bottom-up: Chemical precipitation and coprecipitation; Sol-gel synthesis; Ball milling synthesis; lithography, Plasma Laser deposition (PLD) techniques, Thermolysis routes (Solvothermal, Hydrothermal and pyrolysis), Microwave assisted synthesis; Sonochemical synthesis; Electrochemical synthesis.

### **MODULE II CHARACTERIZATION TECHNIQUES 9**

Structural Characterization: X-ray diffraction, Scanning Electron Microscopy (SEM/HR-SEM/FE-SEM) with EDS, TEM (HR-TEM) and SAED analysis, Atomic force Microscopy (AFM). X-ray Photoelectron spectroscopy (XPS), Raman analysis. Introduction to advanced Scanning Probe Microscopy Techniques Scanning Tunnelling Mode (STM), Piezoelectric force microscopy (PFM). DLS and zeta potential analysis. BET surface area analysis, CHNSO micro analysis.

### **MODULE III APPLICATIONS AND ENVIRONMENTAL IMPACTS 9**

Current applications - Short-term Applications - Long - term Applications –



Energy filed - solar cells, military battle suits. Biomedical applications – Photodynamic therapy in targeted drugs - quantum dot technology in cancer treatment, MRI applications. Nanosensors: pH, heat, humidity, gas, toxic chemicals sensors and sensors for aerospace and defence – biosensors – water remediation - Environmental Impacts: toxicological health effects, relevant parameters in nanoparticles toxicology, integrated concept of risk assessment of nanoparticles.

#### **MODULE IV CONCEPTS OF CATALYSIS 9**

Acid-base catalysis – catalysis by transition metal ions and their complexes – supported transition metal complexes as catalysts – catalysis by enzymes – phase transfer catalysis - photocatalysis – adsorption – chemisorption on metals, metal oxides and semiconductors - kinetics of unimolecular and bimolecular surface reactions - Contact time - WHSV - time on stream - Catalyst deactivation and regeneration, TOF, TON.

#### **MODULE V HETEROGENEOUS CATALYSTS 9**

Metals, metal oxides, mixed metal oxides, supported metals, spinels, perovskites, super acids, hydrotalcites, zeolites and zeotypes (small, medium, large), shape selective catalysts, mesoporous materials (SBA, MCM, KIT, AIPOs, MOFs, COFs) Hydrothermal synthesis, sol-gel process, impregnation method, ion-exchange method - Operations in catalyst manufacture - drying, calcination, spray drying, Reactors- fixed bed and flow reactors.

**L – 45; TOTAL HOURS – 45**

#### **REFERENCES:**

1. T. Pradeep, Nano: The Essentials, Tata McGraw-Hill, New Delhi, 2007.
2. G. Cao, Nanostructures and Nanomaterials –Synthesis, Properties and Applications, Imperial College Press, London, 2004.
3. C. N. R. Rao, A. Muller and A. K. Cheetham, The Chemistry of Nanomaterials, Volume 1, Wiley –VCH Verlag GmbH & Co. KgaA, Weinheim, 2004.
4. G. A. Ozin, A. C. Aresnault, L. Cadematriri, Nanochemistry: A chemical approach to nanomaterials, RSC Publishing, 2008
5. J. Rajaram and J.C. Kuriacose, Kinetics and Mechanisms of Chemical Transformations, Macmillan Publishers India Limited, 2000.
6. B. Viswanathan, S. Sivasanker and A.V. Ramaswamy (Editors), Catalysis

**COURSE OUTCOMES:**

The students will be able to

**CO1:** differentiate the nanomaterials based on their dimensions and acquire knowledge of various synthetic methods

**CO2:** understand the components of instrumental techniques of and characterization techniques for structural and properties of nanomaterials

**CO3:** select the appropriate nanomaterials for specific applications in the interested arena

**CO4:** Find the fundamentals of catalysis

**CO5:** Evaluate significance of heterogeneous catalysts.

**Board of Studies (BoS):** 12<sup>th</sup> BoS of  
Chemistry held on 22.07.2022

**Academic Council:** 19<sup>th</sup> AC  
held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1		L		M	H	H									
CO2	M			H	M	H									
CO3					H	M									
CO4															
CO5															

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 6: Clean Water and Sanitation

SDG 7: Affordable & Clean Energy

SDG 9 : Industry and Innovation

SDG 15 : Life on Land

Statement:

SDG 6, 7 & 9: Foundation to work in R&D of renewable energy and sensors sector and for teaching career.

SDG 15: R&D labs in API labs in the production novel materials for various applications

<b>OEEY 715</b>	<b>STRUCTURAL INTERPRETATION</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4, 9</b>	<b>OF MATERIALS</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To use the concepts (basic and advanced level) of analytical methods for structure elucidation of materials and the students will be trained for the

**COB1:** Interpretation of electronic spectral data of materials

**COB2:** Interpretation of magnetic spectral data of materials

**COB3:** Interpretation of structural and morphological data of materials

**COB4:** Interpretation of thermos analytical data of materials

**COB5:** Interpretation of electrochemical and XPS data of materials

**MODULE I ELECTRONIC DATA 9**

UV-visible, fluorescence and phosphorescence: Characteristic absorption of simple chromophoric groups, conjugated/ aromatic/ ligand systems, metal complexes and materials. FT-IR and Raman: Characteristic group frequencies of organic, inorganic molecules and various materials (polymer, nano, semiconducting) Interpretation of organic and inorganic and hybrid materials using combination of the spectral data.

**MODULE II MAGNETIC AND MASS DATA 9**

Solid-state nuclear magnetic resonance spectroscopy: Compounds containing  $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^{19}\text{F}$ ,  $^{27}\text{Al}$ ,  $^{29}\text{Si}$ , and  $^{31}\text{P}$  nuclei. Electron spin resonance (ESR): Simulation of ESR spectra of paramagnetic species, spin dynamics in solid and liquid. Mass spectrometry: The production and analysis of positive ions, molecular ions, application of isotopic abundance measurements, fragmentation modes and rearrangement of ions. Interpretation of organic, inorganic compounds and materials using combination of the spectral data.

**MODULE III STRUCTURAL AND MORPHOLOGICAL DATA 9**

Fundamental theoretical framework for diffraction (XRD) and imaging methods (SEM, TEM and AFM) used in structural and compositional characterization of materials in solid, film state etc.

**MODULE IV THERMOANALYTICAL DATA AND SURFACE AREA 9**

Interpretation of Differential Thermal Analysis (DTA), Thermo-gravimetric Analysis (TGA), Differential Scanning Calorimetry (DSC) data of various materials including inorganic complex, organic polymeric materials, composite, nano-composites etc; Surface area analysis; isotherms, types, BET surface area, pore dimensions, pore

volume, etc.

## **MODULE V                      ELECTROCHEMICAL AND XPS DATA                      9**

Cyclic voltammetry for oxidation and reduction potentials, TAFEL polarization and Impedance spectroscopy for corrosion inhibitor behavior, chronoamperometry for charge or discharge of battery. X-ray photoelectron spectroscopy: Study the chemical composition and oxidation state of elements at the surface and interface. Applications of XPS in various arenas.

**L – 45; TOTAL HOURS – 45**

### **TEXT BOOKS:**

1. R. S. Drago, Physical Methods for Chemists, W. B. Saunders, 1992.
2. R. M. Silverstein, C. G. Bassler and T. C. Morrill, Spectrophotometric Identification of Organic Compounds, 5th edition, Wiley, 1991.
3. D. H. Williams and I. Fleming, Spectroscopic Methods in Organic Chemistry, 3rd edition, McGraw Hill, 1980.
4. W. Kemp, Organic Spectroscopy, ELBS, 1979.
5. W. L. Jolly, The synthesis and characterization of inorganic compounds, Prentice-Hall, 1970.
6. John Wertz, Electron Spin Resonance: Elementary Theory and Practical Applications, Springer Science & Business Media, 2012.
7. R. F. Speyer, Thermal Analysis of Materials, CRC Press, 1994.
8. P.J. Goodhew, J. Humphreys and R. Beanland, Electron Microscopy and Analysis, Taylor & Francis, 2001.
9. John F Watts, John Woistenhoime, An introduction to surface analysis by XPS and AES, John Wiley and Sons, 2nd edition, 2003.
10. James, B. Condon, Surface Area and Porosity Determinations by Physisorption Measurement and Theory, Elsevier, 1<sup>st</sup> edition, 2006.

### **COURSE OUTCOMES:**

The students will be able to

**CO1:** Interpret electronic spectral data of materials

**CO2:** Interpret magnetic spectral data of materials

**CO3:** Interpret structural and morphological data of materials

**CO4:** Interpret thermo analytical data and porous nature of materials

**CO5:** Interpret electrochemical and XPS data of materials

### **Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

### **Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO1 1	P O 1 2	PSO1	PSO2	PSO3
CO1	H	M		H	M	H									
CO2	H	M		H	M	L									
CO3	H	L		H	M	M									
CO4	H	L		H	M	H									
CO5	H	L		H	M	L									

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 4: Quality Education

SDG 9: Industry and Innovation

Statement:

SDG9: Foundation to work in R&D laboratory, chemical industry, independent researcher and for teaching career.

SDG4: Ensure inclusive and equitable quality education and promote lifelong learning opportunities.

<b>OEEY 742</b>	<b>SURFACE COATING</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 9</b>	<b>TECHNOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

To make the student conversant with

**COB1:** basic principles of surface chemistry

**COB2:** various coating techniques including CVD

**COB3:** industrial coatings and sputtering techniques

**COB4:** surface coating resins and emulsions

**COB5:** techniques like laser alloying and electron beam coating

**MODULE I SURFACE CHEMISTRY OF ALLOYS 9**

Basic physical chemistry, surface chemistry, pretreatment principle - technology and control of electro deposition systems such as alloy plating, electrolysis, composites.

**MODULE II METHODS OF COATING I 9**

Hot dip coatings - principle, surface preparation, methods, applications, Diffusion coatings - Principle - Cementation - Cladding - case hardening - structures.

Chemical vapor deposition - classification-techniques, metal organic type, plasma assisted, layer assisted, applications.

**MODULE III METHODS OF COATING II 9**

Industrial coatings like Enamels, Primers, Putties, Lacquers, Water based paints, Inks, HDPCs, Conversion coatings.

Sputtering techniques, methods, applications, plasma treatments, nitriding, carbonizing, boriding, titanizing methods and applications.

**MODULE IV SURFACE COATING RESINS 9**

Synthesis & characterization of various surface coating resins like Hard resins, Alkyds, Varnishes, Polyesters, Epoxies, Polyamides, Acrylics, Amino resins, CNSL resin, emulsions & water reducible resins.

**MODULE V LASER ALLOY AND ELECTRON BEAM COATING 9**

Laser alloying - sources, variables, methods, applications, Electron beam coating- evaporation materials, methods, applications.

**L – 45; TOTAL HOURS – 45**

**REFERENCES:**

1. G. Braco, Surface Science Techniques, Springer-Verlag Berlin and Heidelberg GmbH & Co. K, 2000.
2. T.S. Sudarsan, Surface Modification Technologies, Marcel Dekker Inc., 1989
3. D.R. Gabe, Principles of Metal Surfaces Treatment and Protection, Pergmon Press 1972.
4. Tracton, Coatings Technology, CRC press, 2006.

**COURSE OUTCOMES:**

The students will be familiar with

**CO1:** the pretreatment methods of electrodeposition

**CO2:** coating methods like, hot dipping, cementation, cladding. Advanced techniques like CVD, plasma assisted, layer assisted methods are also learnt by the students.

**CO3:** composition and characteristics of industrial coatings including enamels, primers etc.

**CO4:** synthesis and characterization of resins, emulsions etc

**CO5:** in detail knowledge about laser alloying, and electron beam coating and their applications in various fields.

**Board of Studies (BoS):**

12<sup>th</sup> BoS of Chemistry held on 22.07.2022

**Academic Council:**

19<sup>th</sup> AC held on  
29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PSO1	PSO2	PSO3
CO1		H				M									
CO2		H			M										
CO3			L			H									
CO4	M				L										
CO5				M		H									

**Note:** L- Low Correlation    M - Medium Correlation    H -High Correlation

SDG 9 : Industry, Innovation and Infrastructure

SDG9: Build resilient infrastructure, promote inclusive and sustainable industrialization and foster innovation

<b>OEEY 743</b>	<b>THIN FILM SCIENCE AND</b>	<b>L</b>	<b>T</b>	<b>P</b>	<b>C</b>
<b>SDG: 4</b>	<b>TECHNOLOGY</b>	<b>3</b>	<b>0</b>	<b>0</b>	<b>3</b>

**COURSE OBJECTIVES:**

**COB1:** To familiarize with preparation and properties of thin films.

**COB2:** To understand the different kinetics of thin film nucleation.

**COB3:** To understand the characterization tools for thin films.

**COB4:** To study the different properties of thin films.

**COB5:** To apply the knowledge of thin film technology into applications.

**MODULE I PREPARATION OF THIN FILMS 9**

Kinetic aspects of gases in a vacuum chamber – classifications of vacuum ranges – production of vacuum - pressure measurement in vacuum systems– thin film (epitaxy) – definition – types of epitaxy. Different Growth Techniques: Liquid phase epitaxy – vapour phase epitaxy – molecular beam epitaxy – metal organic vapour phase epitaxy – sputtering (RF & DC) – pulsed laser deposition. Thickness Measurement: Microbalance technique – photometry-ellipsometry– interferometry.

**MODULE II KINETICS OF THIN FILMS 9**

Nucleation Kinetics: types of nucleation – kinetic theory of nucleation – energy formation of a nucleus – critical nucleation parameters; spherical and non spherical (cap, disc and cubic shaped) Growth Kinetics: Kinetics of binary (GaAs, InP, etc.), ternary ( $\text{Al}_{1-x}\text{Ga}_x\text{As}$ ,  $\text{Ga}_{1-x}\text{In}_x\text{P}$ ,  $\text{InAs}_{1-x}\text{Px}$ , etc.) and quaternary ( $\text{Ga}_{1-x}\text{In}_x\text{As}_1 - y\text{Py}$ , etc.) semiconductors – derivation of growth rate and composition expressions.

**MODULE III CHARACTERIZATION 9**

X-ray diffraction – photoluminescence – UV-Vis-IR spectrophotometer – Atomic Force Microscope – Scanning Electron Microscope – Hall effect – Vibrational Sample Magnetometer – Secondary Ion Mass Spectrometry – X-ray Photoemission Spectroscopy.

**MODULE IV PROPERTIES OF THIN FILMS 9**

Dielectric properties – experimental technique for the determination of dielectric properties – optical properties – experimental technique for the determination of optical constants – mechanical properties – experimental technique for the determination of mechanical properties of thin films – magnetic and superconducting properties.



**MODULE V APPLICATIONS****9**

Optoelectronic devices: LED and Solar cell – Micro Electromechanical Systems (MEMS) – Fabrication of thin film capacitor – application of ferromagnetic thin films; data storage, Giant Magnetoresistance (GMR) – sensors – fabrication and characterization of thin film transistor and FET – quantum dot - Cryptography.

**L – 45; TOTAL HOURS –45****REFERENCES:**

1. Goswami. A, Thin Film Fundamentals, New Age International (P) Limited, New Delhi, 1996.
2. AichaEishabini-Riad, Fred D. Barlow and ISHN, Thin film Technology Handbook, McGraw-Hill Professional Publishers, 1997.
3. Krishna Seshan, Handbook of Thin Film Deposition, William Andrew Publishers, 2012.
4. Donald Smith, Thin-Film Deposition: Principles and Practice, McGraw-Hill Professional Publishers, 1995.
5. K.L.Chopra, "Thin Film Phenomena", Malabar: Robert E. Krieger Publishing Company, 1979.

**COURSE OUTCOMES:**

At the end of the course, the students will be able to

**CO1:** the basic concepts about the thin film technology

**CO2:** the different kinetics of thin film nucleation.

**CO3:** the characterization tools for thin films.

**CO4:** Structural, optical, dielectric and mechanical properties of thin films

**CO5:** applications of thin films in optoelectronics

**Board of Studies (BoS) :**

BOS of Physics was held on 30.6.22

**Academic Council:**

19<sup>th</sup> AC held on 29.09.2022

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PSO1	PS O2	PS O3
CO1	H	M	L	L	M	M	M	L	L	L	M	M	M	M	M
CO2	H	M	M	L	L	M	L	L	L	L	L	M	M	M	M
CO3	H	M	M	L	L	L	L	L	L	L	L	M	M	M	M
CO4	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M
CO5	H	M	M	L	M	M	M	L	L	L	M	M	M	M	M

**Note:** L- Low Correlation    M -Medium Correlation    H -High Correlation

SDG 4 : Ensuring inclusive and equitable quality education for all persons and promote lifelong learning opportunities.

Statement : The modules and topics mentioned in this course are designed to ensure all inclusive and thorough education with equity to all persons and promote learning opportunities at all times.